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Queen et al.

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[45] Date of Patent:

Dec. 17, 1996

[54] HUMANIZED IMMUNOGLOBULINS

[75] Inventors: Cary L. Queen, Los Altos; Harold E.

Selick, Belmont, both of Calif.

[73] Assignee: Protein Design Labs, Inc., Mountain

View, Calif.

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[22] Filed: Jun. 7, 1995

Related U.S. Application Data

[63] Continuation of Ser. No. 634,278, Dec. 19, 1990, Pat. No. 5,530,101, which is a continuation-in-part of Ser. No. 590, 274, Sep. 28, 1990, abandoned, and Ser. No. 310,252, Feb. 13, 1989, abandoned, which is a continuation-in-part of Ser. No. 290,975, Dec. 28, 1988, abandoned.

[51] Int. Cl.⁶ C07K 16/18; A61K 39/395

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Primary Examiner-Lila Feisee

Attorney, Agent, or Firm—Townsend and Townsend and Crew LLP

[57] ABSTRACT

Novel methods for producing, and compositions of humanized immunoglobulins having one or more complementarity determining regions (CDR's) and possible additional amino acids from a donor immunoglobulin and a framework region from an accepting human immunoglobulin are provided. Each humanized immunoglobulin chain will usually comprise, in addition to the CDR's, amino acids from the donor immunoglobulin framework that are, e.g., capable of interacting with the CDR's to effect binding affinity, such as one or more amino acids which are immediately adjacent to a CDR in the donor immunoglobulin or those within about 3 Å as predicted by molecular modeling. The heavy and light chains may each be designed by using any one or all of various position criteria. When combined into an intact antibody, the humanized immunoglobulins of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen, such as a protein or other compound containing an epitope.

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FIGURE 1B

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FIGURE 2A

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FIGURE 2B

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FIGURE 3A

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FIGURE 3B

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FIGURE 4A

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FIGURE 4B

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FIGURE 5A

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FIGURE 6A

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FIGURE 6B

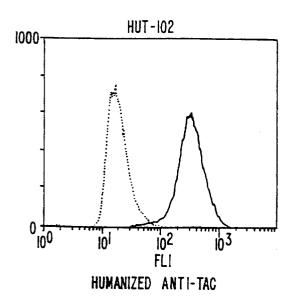


FIGURE 7A

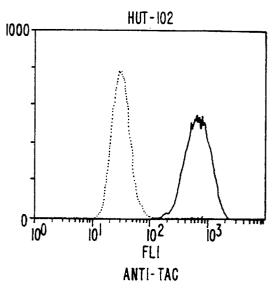


FIGURE 7B

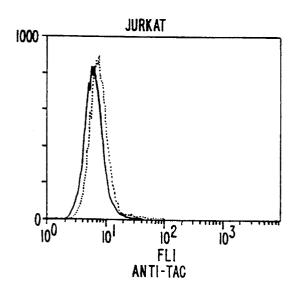


FIGURE 7C

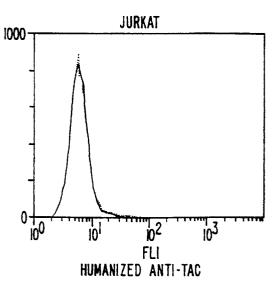


FIGURE 7D

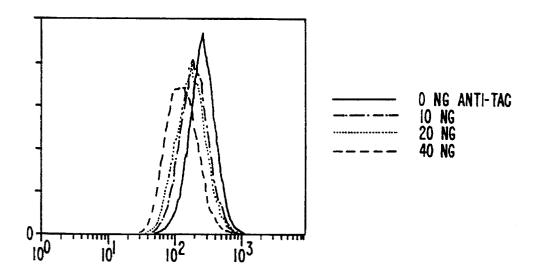


FIGURE 8A

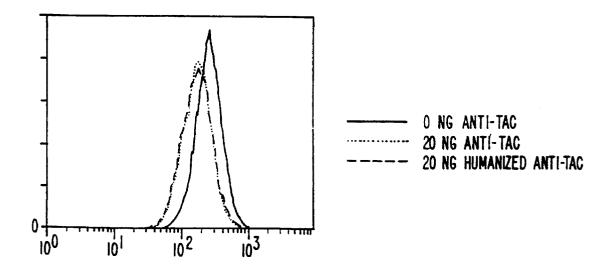
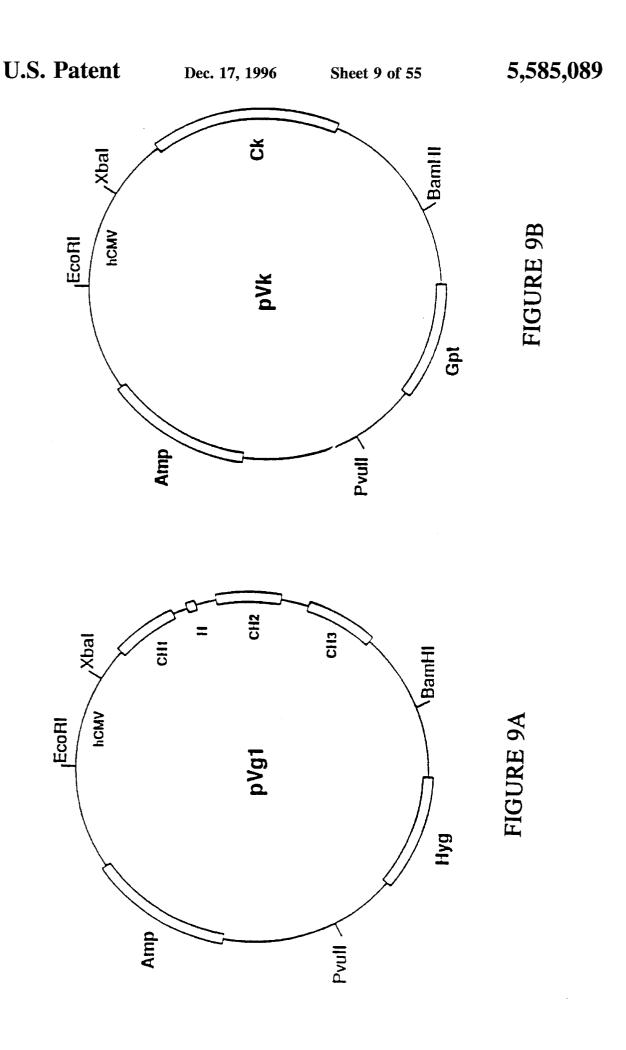


FIGURE 8B



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Is	80 TCCAGCTTGT AGGTCGAACA	90 CCAGTCTGGG GGTCAGACCC	GCTGAAGTCA CGACTTCAGT	AGAAACCTGG TCTTTGGACC	CTCGAGCGTG GAGCTCGCAC	130 AAGGTCTCCT TTCCAGAGGA	140 CCAAGGCTTC
• 1	150 TGGCGGGACC ACCGCCTGG	160 TTTTCTAGCT AAAAGATCGA	ACAGGATGCA TGTCCTACGT	180 CTGGGTAAGG GACCCATTCC	190 CAGGCCCTG GTCCGGGGAC	200 GACAGGGTCT CTGTCCCAGA	210 GGAATGGATG CCTTACCTAC
	220 GGATATATTA CCTATATAAT	230 ATCCGTCGAC TAGGCAGCTG	240 TGGGTATACT ACCCATATGA	250 GAATACAATC CTTATGTTAG	260 AGAAGTTCAA TCTTCAAGTT	270 GGACAGGGTC CCTGTCCCAG	280 ACAATTACTG TGTTAATGAC
,	290 CAGACGAATC GTCTGCTTAG	300 CACCAATAGA GTGGTTATGT	310 GCCTACATGG CGGATGTACC	320 AACTGAGCAG TTGACTCGTC	330 CCTGAGATCT GGACTCTAGA	340 GAGGACACCG CTCCTGTGGC	350 CATTCTATTT GTAAGATAAA
•	360 CTGTGCAGG GACACGTCCC	370 GGTGGGGGAG CCACCCCTC	380 TCTTTGACTA AGAAACTGAT	390 CGAATACAAT GCTTATGTTA	400 gaAgggcTgg ccTcccgAcc	410 TCACAGTCTC AGTGTCAGAG	420 CTCAGGTGAG GAGTCCACTC
	430 TCCTTAAAAC AGGAATTTTG	430 TCCTTAAAAC CTCTAGACGA AGGAATTTTG GAGATCTGCT	TAT				

FIGURE 11A

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CAAATCTAGA GTTTAGATCT	TGGAGACCGA	TACCCTCCTG ATGGGAGGAC	CTATGGGTCC GATACCCAGG	TCCTGCTATG AGGACGATAC	GGTCCCAGGA	TCAACCGGAG AGTTGGCCTC
80	06	100	110	120	130	140
ATATTCAGAT	GACCCAGTCT		TCTCTGCTAG	COTCGGGGAT	AGGGTCACCA	TAACCTGCTC
TATAAGTCTA	CTGGGTCAGA	GGTAGATGGG	AGATGGG AGAGGATC GCAGGCCCTA	GCAGGGGGTA	TCCCAGTGGT	ATTGGACGAG
150	160	170	180	190	200	210
TGCCAGCTCA	AGTATAAGTT	ACATGCACTG	GTACCAGCAG	AAGCCAGGCA	AAGCTCCCAA	GCTTCTAATG
ACCCTCCACT	TCATATTCAA	TGTACGTGAC	CATGGTCGTC	TTCGGTCCGT	TTCGAGGGTT	CGAAGATTAC
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0.3.1	2	013	003	007	017	097
TATACCACAT	CCAACCTGGC	TTCTGGAGTC	CCTTCTCCCT	TCATTGGCAG	TCGATCTCCC	ACCGAGTTCA
ATATGGTGTA	CCTTCCACCC	AAGACCTCAG	GGAAGAGCGA	AGTAACCGTC	ACCTAGACCC	TGCCTCAAGT
290	300	310	320	330	.	250
CCCTCACAAT	CACCTCTCTG	CAGCCAGATG	ATTTCGCCAC	TTATTACTEC	CATCAAAGGA	GTACTTACCC
GGGAGTGTTA	GTCGAGAGAC	GTC	TAAAGCCCTC	AATAATGACG	GTAGTTTCCT	CATGAATGGG
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ACTCACGTTC	GGTCAGGGGA	CCAAGGTGGA	GGTCAAACGT	AAGTACACTT	TTCTAGATAT	<
TGAGTGCAAG	CCAGTCCCCT	GGTTCCACCT	CCAGTTTGCA	TTCATGTGAA	AAGATCTATA	

FIGURE 11B

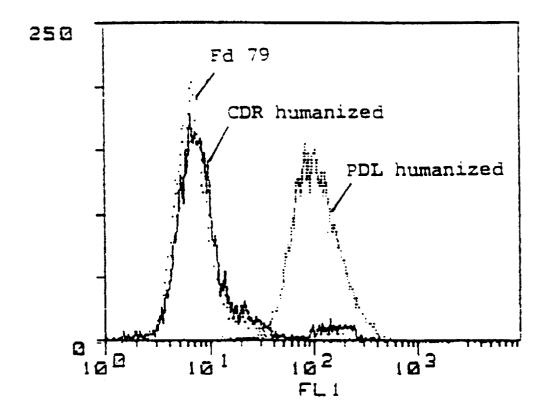


FIGURE 12

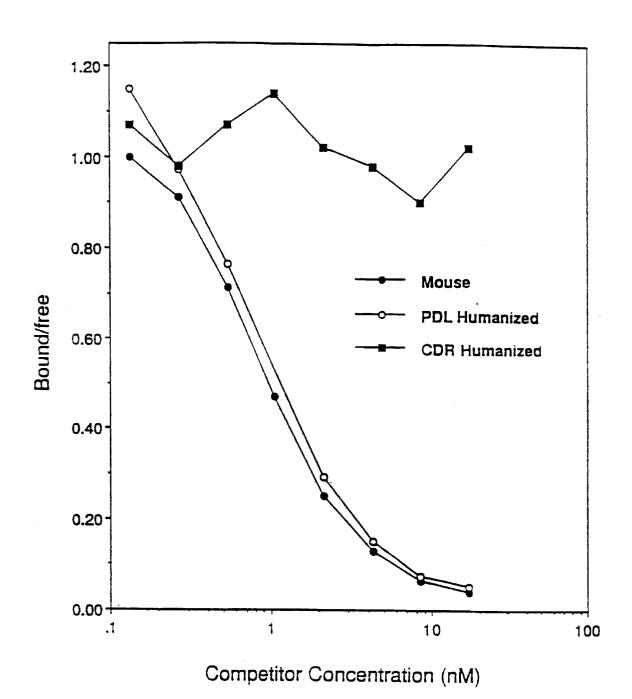


FIGURE 13

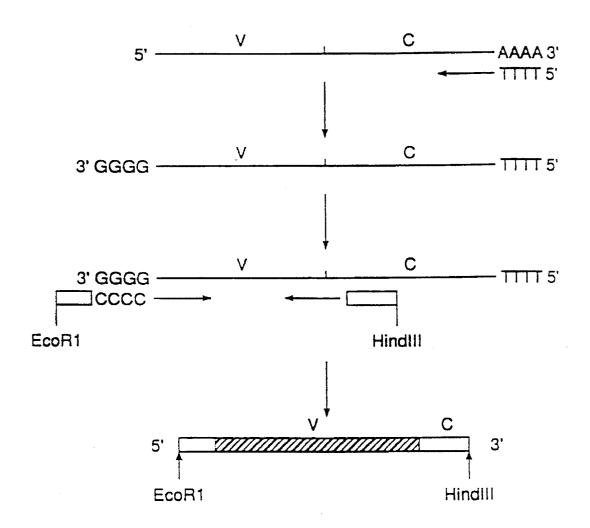


FIGURE 14

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V S C	K A S	G Y T	FTSY	R M H W	V R Q
CCCCTGG	190 A CAGGG TC TG	200 GAATGGATTO	22 22 22 22 22 22 22 22 22 22 22 22 22 2	O 230 CGTCGACTGGGT	240 A T A C T C A A T
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A CATGG A	310 Actgaggagg	320 CTGAGATCT	330 34 GAGGACACCGCAG	10 350 STCTATTACTGTG	360 CAAGAGGG
Y M E	L S S	L R S	E D T A		A R G
GGGGGGT	370 CTTTG ACTAC	380 TGGGGCCAA	390 40 GGAACCCTGGTCA	00 410 NCAGTCTCCTCAG	420 GTGAGTCCT
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TAAAACC	430 TCTAGA				

FIGURE 17

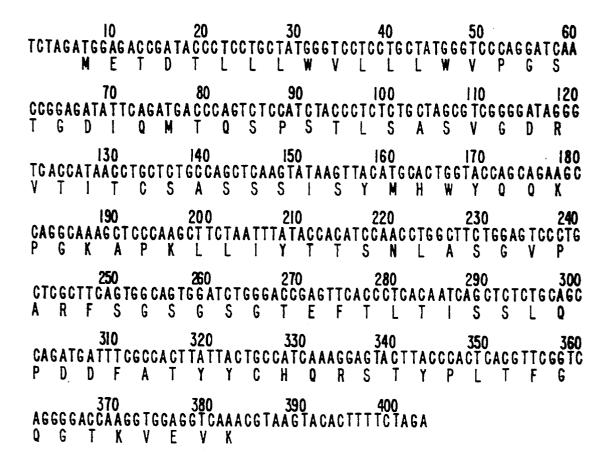
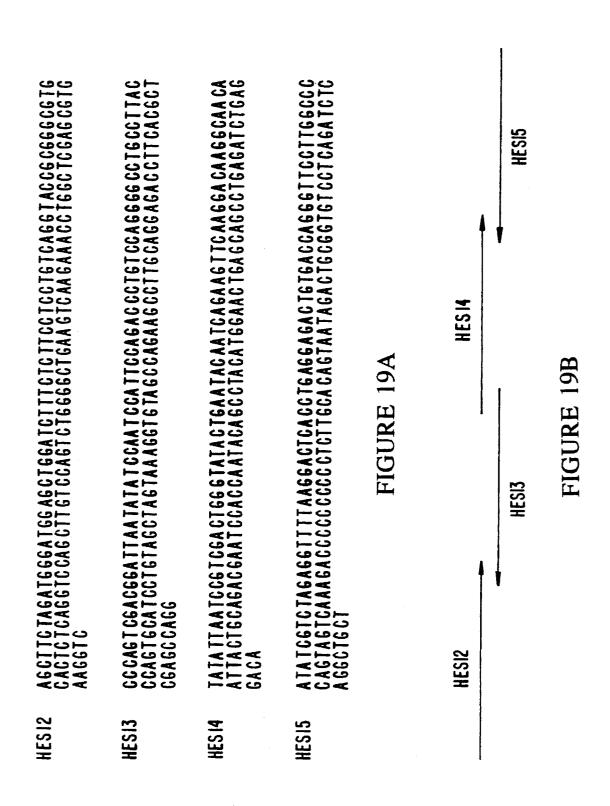
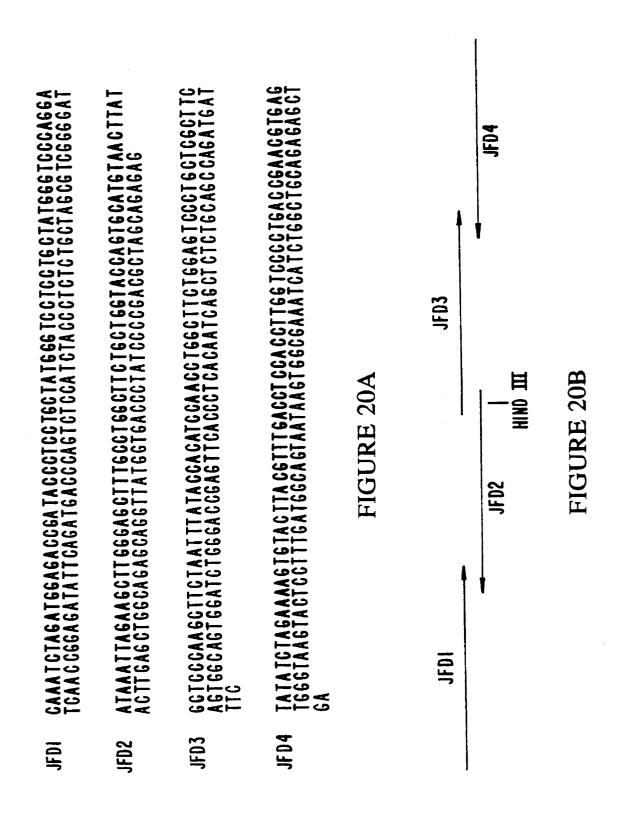


FIGURE 18

Dec. 17, 1996





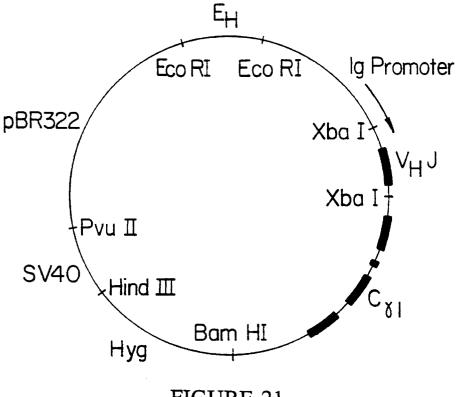


FIGURE 21

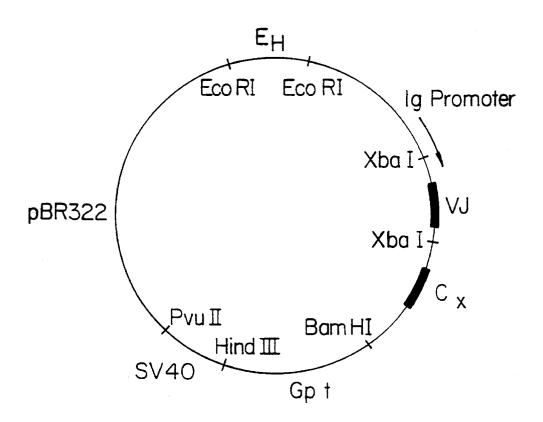


FIGURE 22

ATGGATTTTCAAGGCAGATTTTCAGCTTCCTGCTAATCAGTGCCTCAGTCATATCTGTCC M D F Q V Q I F S F L L I S À S V I L S AGAGGACAAATTGTTCTCACCCAGTCTCCAGCAATCATGTCTGCGTCTCCAGGGGCGAAG R G Q I V L T Q S P A I M S A S P G E K GTCACCATGACCTGCAGTGGCAAGCCTAAGTGTAAGTTTCATGTCTGCAGCAGAGG V T M T C S G S S S V S F M Y W Y Q Q R CCAGGATCCTCCCCCAGACTCCTGAATTTATGACACATCCAACCTGGATCCCAGAGGG V T M T C S G S S S V S F M Y W Y Q Q R CCAGGATCCTCCCCCAGACTCCTGATTTATGACACATCCAACCTGGCTTCTGGAGTCCCT P G S S P R L L I Y D T S N L A S G V P GTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCTCACAATCAGCCGAATGGAG V R F S G S G S G T S Y S L T I S R M E GCTGCAAGATGCTGCCACTTATTACTGCCAGCAGTGGAGTACTTACCCGGTCCGGT A E D A A T Y Y C O Q W S T Y P L T F G GCTGGGACCAAGCTGGAGCTGAAA A G T K L E L K FIGURE 23A ATGGCTGTTTGGGGCTGCTCTTTCTCCCTGGTGACCTCCCACAGCCCTCACAGCCTGTCCCTACCCGTCCGGT M A V L G L L F C L V T F P S C V L S Q GTGCAAGCTGAGCAGCTCAGGACCTGGCCTCACAGGCCTGTCCCTACCCCC V Q L K Q S G P G L V Q P S Q S L S I T GCACCAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCCCAA C T V S G F S V T S Y G V H W I R Q S P GGAAAAGGGTCTGGAGTGGAGACTAACAAGTTATGGTGTACACTGGATTCCCAA C T V S G F S V T S Y G V H W I R Q S P GGAAAAGGGTCTGGAGTGGCTGGCAGAGTGATATAGGAGGACTATAATCA C K G L E W L G V I W S G G S T D Y N A GCTTTCATATCCAGACCTGGCCTGACCACAAGAGCCAAGACTATAATCA C K G L E W L G V I W S G G S T D Y N A GCTGCAACAGTCTCGACTGGCCTGACCACAAGGACCAAGACTATAATCA C K G L E W L G V I W S G G S T D Y N A GCTTCATATCCAGACTGACCATCACCAACACACCAACAACCTATAATCAA C K G L E W L G V I W S G G S T D Y N A A F I S R L T I S K D N S K S Q V F F K AATTACGACGGTTTCCTAACCACACACCACCAACACCAAACCTATAACCTATATACTAC				•			•			30	·= c c	· m » »	, m C 1	• * (~T)	~~~	רר זו	• •	מידי	ריין בי	60 TCC
AGAGGACAAATTGTTCTCACCCAGTCTCCAGCAATCATGTCTGCGTCTCCAGGGGCGAAG R G Q I V L T Q S P A I M S A S P G E K 150						_												I		
R G Q I V L T Q S P A I M S A S P G E K 150				•			•			90				•			•	~~~		
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GTCACCATGACCTGCAGTGGCAGCTCAAGTGTAAGTTTCATGTACTGGTACCAGCAGAGG V T M T C S G S S S V S F M Y W Y Q Q R 210	R	G	Q	I	V	L	T	Q	S	P	A	1	M	5	A	3	2	G	Ē.	K
GTCACCATGACCTGCAGTGGCAGCTCAAGTGTAAGTTTCATGTACTGGTACCAGCAGAGG V T M T C S G S S S V S F M Y W Y Q Q R C 240 CCAGGATCCTCCCCCAGACTCCTGATTATGACCACCTGGCTTCTGGAGTCCCT P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S P R L L I Y D T S N L A S G V P G S S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G T S Y S L T I S R M E S G S G S G S T S D T S Y S L T T S S G S G S G S T S S T S S G S G S										150				•			•			180
V T M T C S G S S S V S F M Y W Y Q Q R 210 240 CCAGGATCCTCCCCCAGACTCCTGATTTATGACACATCCAACCTGGCTTCTGGAGTCCCT P G S S P R L L I Y D T S N L A S G V P GTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCTCACAATCAGCCGAATGGAG V R F S G S G S G T S Y S L T I S R M E GCTGGAGATGCTGCCACTTATTACTGCCAGCAGTGGAGTACTTACCCGCTCACGTTCGGT A E D A A T Y Y C Q Q W S T Y P L T F G GCTGGGACCAAGCTGGAGCTGAAA A G T K L E L K FIGURE 23A ATGGCTGTCTTGGGGCTGCTTTCTGCCTGGTGACATTCCCAAGCTGTGTCCTATCCCAG M A V L G L L F C L V T F P S C V L S Q GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T GCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA C T V S G F S V T S Y G V H W I R Q S P GGAAAAGGGTCTGGAGTGGGAGTGGATATTGGAGTGGAAGCACAGACCAGACCTTCCAC C T V S G F S V T S Y G V H W I R Q S P GGAAAAGGGTCTGGAGTGGGAGTGGATATTGGAGTGGAAGCACAGACCAGACCTATAATGCA G K G L E W L G V I W S G G S T D Y N A GCTTTCATATCCAGACTGGACCTGACAACAGAGCCAAGACCCAAGACCCAACAACACTTTCTTT	GTC	ACC	ATG	ACC	TGC	AGTO	GGC.	AGC'			STAR	AGT	TTC.	ATG	TAC'	rgg:	TAC	CAG	CAG	AGG
CCAGGATCCTCCCCCAGACTCCTGATTTATGACACATCCAACCTGGCTTCTGGAGTCCCT P G S S P R L L I Y D T S N L A S G V P CTO																W	Ā	Q	Q	R
CCAGGATCCTCCCCCAGACTCCTGATTTATGACACATCCAACCTGGCTTCTGGAGTCCCT P G S S P R L L I Y D T S N L A S G V P CTO				_						210										240
P G S S P R L L I Y D T S N L A S G V P . 270	CCA	CCA	ጥርር	ቸርር	ררר:	AGA(- ሞር	רייה:			SAC	ACA:	rcc.	AAC	CTG	GCT'	TCT	GGA		
GTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCACAATCAGCCGAATGGAG V R F S G S G S G T S Y S L T I S R M E															<u>L</u>				V	
GTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCACAATCAGCCGAATGGAG V R F S G S G S G T S Y S L T I S R M E																	_			200
	C-mm	~~~	mmc	•	~~~	3 C TT	•	m C m		270	T C T'	די א ריי	$r \subset r$	• •		a ጥር	AGC	CGA	атс	
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GCTGGGGCCACTTATTACTGCCAGCAGTGGAGTACTTACCCGCTCACGTTCGGT A E D A A T Y Y C O O W S T Y P L T F G GCTGGGACCAAGCTGGAGCTGAAA A G T K L E L K FIGURE 23A A G T K L E L K FIGURE 23A ATGGCTGTCTTGGGGCTGCTCTTCTGCCTGGTGACATTCCCAAGCTGTGCCTATCCCAG M A V L G L L F C L V T F P S C V L S Q GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA C T V S G F S V T S Y G V H W I R Q S P GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGAAGCACAGACTATAATGCA G K G L E W L G V I W S G G S T D Y N A GCTTTCATATCCAGCACTGACCATCAGCAAGGGACCAAGACTATAATGCA A F I S R L T I S K D N S K S Q V F F K GTGAACAGTCTGGAACCTGCCATCACCAGAGGGACCAAGACTTTCTTT	•		-	٥	C	J	J	J	•	-	_	_								
GCTGGGACCAAGCTGAACAAAAAAAAAAAAAAAAAAAAA				•			•							•			•	300	- en en C	
GCTGGGACCAAGCTGGAGCTGAAA A G T K L E L K FIGURE 23A ***STATE C L K BETT CTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT									TGC	CAG	CAG'	TGG.	AGT c	ACT	TAC	و ا		ACC T	F -	,GG 1
FIGURE 23A ***TAGGETGTCTTGGGGGCTGCTCTTCTGCCTGGTGACATCCCAAGCTGTGTCCTATCCCAG** **M A V L G L L F C L V T F P S C V L S Q** **GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC** **V Q L K Q S G P G L V Q P S Q S L S I T** **TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA** **C T V S G F S V T S Y G V H W I R Q S P** **GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA** **G K G L E W L G V I W S G G S T D Y N A** **GCTTTCATATCCAGACTGACCATCAGCAAGGACCAAGACTATTAATGCA** **A F I S R L T I S K D N S K S Q V F F K** **GTGAACAGTCTGCAACCTGCTGACCACAGCCAATATACTATTGTGCCAAGAGCCTAGGGGACCTAT** **V N S L Q P A D F A I Y Y C A R A G D Y** **AATTACGACGGTTTTGCTTACTGGGGGCCAAGGGGACTCTGGTCACTGTCTCTGCG**	A	드	ט	A	A	Т	ž	ĭ	C	0	<u> </u>	VV	<u> </u>	- _					-	•
FIGURE 23A ***TAGGETGTCTTGGGGGCTGCTCTTCTGCCTGGTGACATCCCAAGCTGTGTCCTATCCCAG** **M A V L G L L F C L V T F P S C V L S Q** **GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC** **V Q L K Q S G P G L V Q P S Q S L S I T** **TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA** **C T V S G F S V T S Y G V H W I R Q S P** **GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA** **G K G L E W L G V I W S G G S T D Y N A** **GCTTTCATATCCAGACTGACCATCAGCAAGGACCAAGACTATTAATGCA** **A F I S R L T I S K D N S K S Q V F F K** **GTGAACAGTCTGCAACCTGCTGACCACAGCCAATATACTATTGTGCCAAGAGCCTAGGGGACCTAT** **V N S L Q P A D F A I Y Y C A R A G D Y** **AATTACGACGGTTTTGCTTACTGGGGGCCAAGGGGACTCTGGTCACTGTCTCTGCG**				•			•													
FIGURE 23A	GCT	GGG	ACC	AAG	CTG	GAG	CTG	AAA												
ATGGCTGTCTTGGGGGCTGCTCTTCTGCCTGGTGACATTCCCAAGCTGTGTCCTATCCCAGG M A V L G L L F C L V T F P S C V L S Q GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA C T V S G F S V T S Y G V H W I R Q S P GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA G K G L E W L G V I W S G G S T D Y N A GCTTTCATATCCAGACTGACCATCAGCAAGGACCAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGACTATACTATTCTTTAAA A F I S R L T I S K D N S K S Q V F F K AATTACGACGGTTTTGCTTACTGGGGCCCAAGGGACTCTGGTCACTGGTCACTGGGACCTAGCAAGGGACTCTGGTCACTGTCAC	A	G	T	K	L	Ε	L	K												
ATGGCTGTCTTGGGGCTGCTCTTCTGCCTGGTGACATTCCCAAGCTGTGTCCTATCCCAG M A V L G L L F C L V T F P S C V L S Q									F	GU	JRI	Ξ 2:	3A							
ATGGCTGTCTTGGGGGCTGCTCTTCTGCCTGGTGACATTCCCAAGCTGTGTCCTATCCCAG M A V L G L L F C L V T F P S C V L S Q																				
M A V L G L L F C L V T F P S C V L S Q OUTGOURN STREET S							•			30				•			•			60
GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T 150	ATG	GCI	GTC	·	GGG	CTG	·	TTC	TGC		GTG	ACA	TTC	CCA	\AGC	TGT	GTC	CTA	ATCO	
GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T 150										CTG					LAGC S			CT <i>I</i> L	ATC(S	CCAG
V Q L K Q S G P G L V Q P S Q S L S I T 150										CTG L	V				AGC S			CT <i>I</i> L	ATC(S	CCAG Q
TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA C T V S G F S V T S Y G V H W I R Q S P	М	A	V	L •	G	L	L •	F	С	CTG L 90	V	T	F	P •	S	С	٧.	L	S	CCAG Q 120
TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA C T V S G F S V T S Y G V H W I R Q S P	M GTC	A CAC	V GCTC	L • •	G SCAG	L TCA	L • .GGA	F \CCI	C :GGC	CTG L 90	V .GTG	T CAG	F CCC	P • CTC#	S ACAG	C SAGO	V CTC	L STC	S	CCAG Q 120
C T V S G F S V T S Y G V H W I R Q S P . 210	M GTC	A CAC	V GCTC	L • •	G SCAG	L TCA	L • .GGA	F \CCI	C :GGC	CTG L 90	V .GTG	T CAG	F CCC	P • CTC#	S ACAG	C SAGO	V CTC	L STC	S	CCAG Q 120 CACC T
GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA G K G L E W L G V I W S G G S T D Y N A CONTROL S R L T I S K D N S K S Q V F F K GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGACTAT V N S L Q P A D F A I Y Y C A R A G D Y AATTACGACGGTTTTGCTTACTGGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG	M GTC V	A GCAC Q	V GCTC L	L SAAC K	G CAG Q	L TCA S	L	F CCT	C G G	CTG 90 CTA L	V .GTG V	T CAG Q	F CCC	P TCA S	S ACAG Q	C SAGC S	V CTC	L STC S	S CATO	CAG Q 120 CACC T
GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA G K G L E W L G V I W S G G S T D Y N A CONTROL STANDARD S	M GTC V	A GCAC Q	V GCTC L	L SAAC K	G GCAG Q TGGT	L TCA S	L .GGA G	F ACCI P	C G G	20TG L 90 CTA L 150 AAGT	V .GTG V	T CAG Q	F CCC P	P CTCA S	S ACAG Q CTGG	C SAGC S SATT	V CTC L	E STC(S ,S	S CATO I GTO!	CAG Q 120 CACC T
• 270 • 300 GCTTTCATATCCAGACTGACCATCAGCAAGGACCAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K • 330 GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGGACTAT V N S L Q P A D F A I Y Y C A R A G D Y AATTACGACGGTTTTGCTTACTGGGGCCCAAGGGACTTTGGTCACTGCGG AATTACGACGGTTTTGCTTACTGGGGCCCAAGGGACTTTGGTCACTGTCTCTGCG	M GTC V	A GCAC Q	V GCTC L	L SAAC K	G GCAG Q TGGT	L TCA S	L .GGA G	F ACCI P	C G G	20TG L 90 CTA L 150 AAGT	V .GTG V	T CAG Q	F CCC P	P CTCA S	S ACAG Q CTGG	C SAGC S SATT	V CTC L	E STC(S ,S	S CATO I GTO!	CAG Q 120 CACC T
. 270 . 300 GCTTTCATATCCAGACTGACCATCAGCAAGGACAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K	M GTG V TGC C	A GCAC Q CAC T	V GCTC L AGTC V	L GAAC K CTCT	G G Q G G	L STCA S TTTC F	L .GGA G .TCA	F ACCI P AGTA V	C G G AAC# T	2000 PORTON SERVICE SE	V GTG V TAT	T CAG Q GGT G	F CCC P GTA	P CTC# S ACAC	S ACAG Q CTGG	C SAGC SATT	V CTC L CCGC R	ETC(,S ,S CCA(S CATO I GTO:	CCAG Q 120 CACC T 180 TCCA P
GCTTTCATATCCAGACTGACCATCAGCAAGGACAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K	M GTG V TGC C	A Q CAC! T	V GCTC L AGTC V	EAAC K	G CAG Q G G G G	L S S TTCA TTC	L .GGA G .TCA S	F CCT P V GTA V	C G G LACA T	210	GTG V TAT	T CAG Q GGT G	GTF	P STCF S ACAC H	S ACAG Q ETGG W AAGG	C S S SATI I	V CTC L CCGC R	ETCO STCA CCAC	S CATO I GTO: S	CCAG Q 120 CACC T 180 TCCA P 240 TGCA
GCTTTCATATCCAGACTGACCATCAGCAAGGACAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K	M GTG V TGC C	A Q CAC! T	V GCTC L AGTC V	EAAC K	G CAG Q G G G G	L S S TTCA TTC	L .GGA G .TCA S	F CCT P V GTA V	C G G LACA T	210	GTG V TAT	T CAG Q GGT G	GTF	P STCF S ACAC H	S ACAG Q ETGG W AAGG	C S S SATI I	V CTC L CCGC R	ETCO STCA CCAC	S CATO I GTO: S	CCAG Q 120 CACC T 180 TCCA P 240 TGCA
A F I S R L T I S K D N S K S Q V F F K	M GTG V TGC C	A Q CAC! T	V GCTC L AGTC V	EAAC K	G CAG Q G G G G	L S S TTCA TTC	L .GGA G .TCA S	F CCT P V GTA V	C G G LACA T	90 CTA L 150 AAGT SATA	V GTG V TAT Y ATGG	T CAG Q GGT G	GTF	P STCF S ACAC H	S ACAG Q ETGG W AAGG	C S S SATI I	V CTC L CCGC R	ETCO STCA CCAC	S CATO I GTO: S	CCAG Q 120 CACC T 180 TCCA P 240 TGCA A
. 330 360 GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGGACTAT V N S L Q P A D T A I Y Y C A R A G D Y . 390 AATTACGACGGTTTTGCTTACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG	M GTG V TGC C	A GCAC Q CAC T T AAAC K	V ECTO L AGTO V EGGS	L GAAC K CTCTC S CTCTC	G Q Q TGGT G G GGAG	L S TTTC F GTGG W	L .GGA G .TCF S .GCTC	F ACCI P AGTA V GGGA G	C G G ACA T AGTO	90 CTA L 150 AAGT S 210 GATA	V GTG V TAT Y V	T CAG Q GGT G SAGT	F CCC P GT# V	P CTCA S ACAC H G G	S ACAG Q CTGG W AAGG	C SAGO SATI I CAC	V CTC L CCGC R	L STCO	S CATO S GTO: S TAA	CCAG Q 120 CACC T 180 TCCA P 240 TGCA A 300
GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGGACTAT V N S L Q P A D T A I Y Y C A R A G D Y	M GTG V TGC C GG#	A GCAC Q CACA T T AAAAC K	V GCTC L AGTC V GGGT	EAAC K STCT S FCTC	G GCAG Q TGGT G GGAG	L STCA STTTC F STGG W	L GGGA G G TCA S GCTC L GAGGA GAGA GAGGA G	F ACCT AGTA V AGGA G	C GGC G AACA T V CAGTO	210 CAAC	GTG V TAT Y ATGG	T CAG Q CGGT G SAGT S	F CCCC P CGT/	P STCA S ACAC H IGGA	S ACAG Q ETGG W AAGG S	C SAGO	V COTO L COGO R AGAO D AGT	L STCCONS	SCATO STAA	CCAG Q 120 CACC T 180 TCCA P 240 TGCA A 300 TAAA
V N S L Q P A D T A I Y Y C A R A G D Y • • 390 • • • 390 • AATTACGACGGTTTTGCTTACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG	M GTG V TGC C GG#	A GCAC Q CACA T T AAAAC K	V GCTC L AGTC V GGGT	EAAC K STCT S FCTC	G GCAG Q TGGT G GGAG	L STCA STTTC F STGG W	L GGGA G G TCA S GCTC L GAGGA GAGA GAGGA G	F ACCT AGTA V AGGA G	C GGC G AACA T V CAGTO	90 CCTA L 1500 AAGT S 210 EATF 1 270 CAAG	V GTG V TAT Y ATGG W GGAC	T CAG Q CGGT G SAGT S	F CCCC P CGT/	P STCA S ACAC H IGGA	S ACAG Q ETGG W AAGG S	C SAGO SATI CACA	V COTO L COGO R AGAO D AGT	L STCCONS	SCATO STAA	CCAG Q 120 CACC T 180 TCCA P 240 TGCA A 300 TAAA K
• • 390 • • AATTACGACGGTTTTGCTTACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG	M GTG V TGC C GGA GC:	A GCAC Q CACA T AAAC K CTTTC	V GCTC L AGTC V GGGGGG G	L S CTCT S ATCC S	G GCAG Q CGGT G E E CAGA	L TCA S TTTC F W ACTG	L GGGA G G TTCA S GCTC L GACCO T	F ACCI P AGTA V GGGGA G	C G G LACA T V V CAGO S	900 CTA L 1500 AAGT S 2100 CAAAC K 3300 K 3300	V GTG V TAT Y ATGG W OGGAC D	T CAG Q Q CAG S S CAAC N	F CCCC P CGGT V CGGT S	P CTCA S ACAC H CCAAC K	S ACAG Q ETGG W AAGG S GAGG	C SAGO S SATT I CACA T Q	V CTC L CCGG R AGAG D AGGT V	L GTCC S CCA Q CTA Y	S CATO	120 CACC T 180 TCCA P 240 TGCA A 300 TAAA K
AATTACGACGGTTTTGCTTACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG	M GTG C GGA A GTG	A GCAC Q LACA T AAAC K F TTTC	V GCTC L AGTC V GGGGT G G CATA	L S CTCT S ATCC S TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCTCT TCT TC	G GCAG Q CGGT G E E CAGA	L TCA S TTCO F GTGG W ACTC	L GGGA G G S GCTC L GACC T GGCT	F ACCI P AGTA V GGGGA G I	COCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCO	900 CTA L 1500 AAGT L 210 CAAC K 3300 AAGCC	V GTG V TTAT Y ATGG W GGGAC D CATA	T CAG Q CGGT G AGT S AGT N	F CCCC P CGT/	P CTCA S ACAC H CCAAC K TTTG	S ACAG Q ETGG _ W AAGG S GAGG	C GAGG	V CCTC L CCGC R AGAC D AGT V AGCC AGCC	L STCO	S CATO	120 CACC T 180 TCCA P 240 TGCA A 300 TAAA K
	M GTG C GGA A GTG	A GCAC Q LACA T AAAC K F TTTC	V GCTC L AGTC V GGGGT G G CATA	L S CTCT S ATCC S TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCT TCTCT TCTCT TCT TC	G GCAG Q CGGT G E E CAGA	L TCA S TTCO F GTGG W ACTC	L GGGA G G S GCTC L GACC T GGCT	F ACCI P AGTA V GGGGA G I	COCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCOCO	900 CCTA 1500 AAGT 2700 K 3300 AAGCO AAGCO AAGCO A	V GTG V TTAT ATGG W CGGAC D CATA	T CAG Q CGGT G AGT S AGT N	F CCCC P CGT/	P CTCA S ACAC H CCAAC K TTTG	S ACAG Q ETGG _ W AAGG S GAGG	C GAGG	V CCTC L CCGC R AGAC D AGT V AGCC AGCC	L STCO	S CATO	120 CACC T 180 TCCA P 240 TGCA A 300 TAAA K
NYDGFAYWGQGJLV, SA	M GTG C GGA GCTG V	A GCAC Q CACA T AAAC K FTTC F GAAC	V GCTC L AGTC V GGGGT G CATA CAGG	L GRANCE S GRANCE ATCC TCTC L	G GCAG Q TGGT G E GGAG E CAGA	L STTCA STTGG W ACTG	L	F ACCI P AGTA V GGGA G G I GAC I I GAC	C C C C C C C C C C C C C C C C C C C	900 CCTA 1500 AAGTI 2700 K 3300 AAGCO AAGCO AAGCO A	V GTG V TTAT TTGG W CATA CATA ()	T CAG Q CGGT G G G G G G G G G G G G G G G G G	F CCCC P CGT/V CGGT S CTCC S	P CTCA S ACAG H CAAG K CTTG C CTTG C	S ACAG Q CTGG - W AAGG S GAGG S	C SAGO SATT I CCAA Q CAGA R	V CCTC L CCGC R AGAC V AGCC AGCC A	L STCCAGE S S S S S S S S S S S S S S S S S S S	S CATO I GTC' S TAA' N CTT F	CCAG Q 120 CACC T 180 TCCA P 240 TGCA A 300 TAAAA K 360 CTAT
	M GTC V TGC C GGA A GTC V AAA	A GCAC Q LACI T AAAC K CTTC F GAAC N LTA	V GCTC L AGTC V GGGGG G CATA CAGGGGG CAGGGGGGGGGGGGGGGG	L GAAC ATCC ATCC CGG CGG	G GCAG Q CGGG G E CAGA R GCAA Q CTTTT	L TCA S TTCO F GTGG W ACTG L ACCT	L	F ACCI P AGTA V AGTA I IGAC D	C C C C C C C C C C C C C C C C C C C	PORTOL STATE OF THE PROPERTY O	V GTG V TAT LTGG W OGGAC D AGGAC	T CAG Q CGGT GGAG AGAC N ATAC Y	F CCCC P PGTA CGG S CTA Y	P CTCA S ACAG H CAAG K CTTG C CTTG C	S ACAG Q CTGG - W AAGG S GAGG S	C SAGO S SATT I COAM Q CAGA R	V .CTC .CCGC R .AGAC V .CCGC AGAC V .CCGC AGAC AGTC .CCGC AGAC AGCC AGCC AGCC AGCC AGCC AG	L STCO	S CATO I GTO S TAAA N CTT F GGA G	120 CACC T 180 TCCA P 240 TGCA A 300 TAAA K

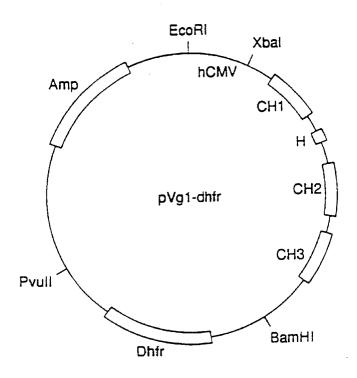


FIGURE 24A

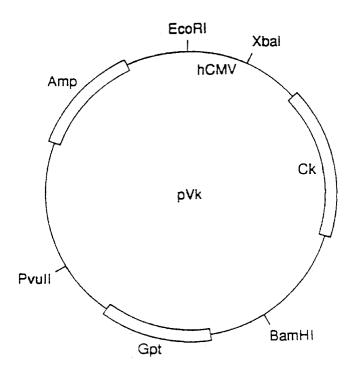


FIGURE 24B

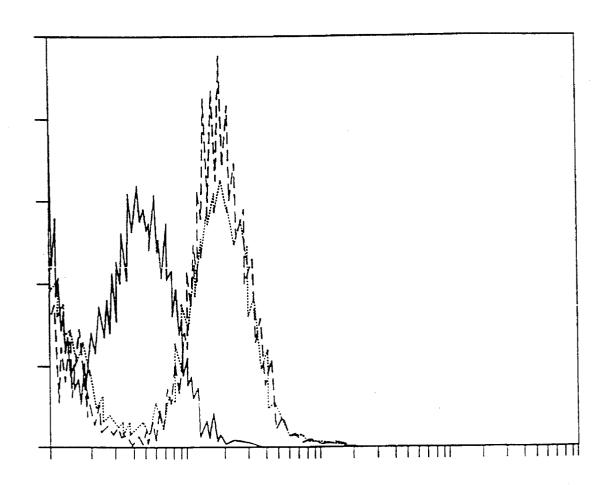


FIGURE 25

```
S
S
                                     S
S
                                        L
                                           S V
S A
                                                   S
S
1
                         Q S P
        D I Q M
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                           S
                               Ρ
          I Q M
                         Q
        I T C Q I T C S
                                                L
                                                   N
                                                      W
21
                    A S Q
                               N
                                      N A
                                         S
21
                  P
P
                                                T
                                                      Ε
                                                            G
41
        G <u>K</u>
                        L
L
                                            S
                     K
                               Ι
                               Ι
                                  Y D
                     K
                            L
                                            S
                                                N
40
              A
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S
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D
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61
        R
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              S
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        R
                     S
                         G
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                                                F
60
              S
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                               C Q Q
C <u>Q</u> Q
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Λ
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81
                                                Ν
                  Α
80
        E D I
                                         W
101
        G T K V
                     Ε
                         V
100
        GTKV
                     Ε
```

FIGURE 26A

1	A E	V	Q Q	L L	L L	E E	s s	G G	G G	G G	L L	V V	Q Q	P P	G G	G G	s s	ī.	R R	LL.
21 21	S S	СС	A A	A A	S S	G G	F F	T	F V	S Ţ	A <u>S</u>	S Y	A G	M V	S H	W W	V V	R R	Q Q	A A
41 41	יט יט	GG	K	GG	1 . 1	E	M	V V	A G	W	A K	Y -	E W	И S	G	N G	2 3	K.	H D	¥ • <u>'</u>
61 60	A <u>N</u>	D A	S A	V F	N I	G S	R R	F F	T T	I I	S S	R R	N D	D <u>N</u>	\$ \$	K K	N	6.3.4	L	ž Ž
81 80	L L	Q Q	M M	N N	G S	L L	Q Q	A A	Z E	<u>D</u>	S	A A	I I	Y Y	Y Y	CC	A A	2 2 2	D	A A
101 99	G G	P D	Y Y	V	S N	0, >	T D	F G	F 11.	А <u>А</u>	H Y	W	G G	Q Q	GG	1.3 1.3	. 1 . 1	V V	E T	⊽ V
121 118	S S	S S																		

FIGURE 26B

U.S. Patent Dec. 17, 1996 Sheet 28 of 55

vc13

10 20 30 40 50 60 TTCTGCTGGT ACCAGTACAT GAAACTTACA CTTGAGCTGC CACTGCAGGT GATGGTGACG 70 80 90 CGGTCACCCA CTGAGGCACT GAGGCTAGAT GGAGACTGGG TCATTTG

vc14

20 30 40 50 CATGTACTGG TACCAGCAGA AGCCAGGAAA AGCTCCGAAA CTTCTGATTT ATGACACATC 90 100 CAACCTGGCT TCTGGAGTCC CTTCCCGCTT CAGTGGCAGT GGGTCTGGGA CCGATTACAC 130 CTTTACAATC TCTTCA

vc15

30 40 20 50 TGTGTCTAGA AAAGTGTACT TACGTTTTAC CTCGACCTTG GTCCCTTGAC CGAACGTGAG 90 100 CGGGTAAGTA CTCCACTGCT GGCAGTAATA AGTGGCTATA TCTTCCGGCT GAAGTGAAGA 130 GATTGTAAAG GTGTAAT

vc16

10 20 30 40 50 CACATCTAGA CCACCATGGA TTTTCAAGTG CAGATCTTCA GCTTCCTGCT AATCAGTGCC 90 TCAGTCATAC TGTCCAGAGG AGATATTCAA ATGACCCAGT CTCCATCT

FIGURE 27A

wps57

10 20 30 40 50 60
ACACTCTAGA AGTTAGGACT CACCTGAAGA GACAGTGACC AGAGTCCCTT GGCCCCAGTA

70 80 90 100 110
AGCAAAACCG TCGTAATTAT AGTCCCCAGC TCTGGCACAA TAATATATGG CTGTGTCC

FIGURE 27B

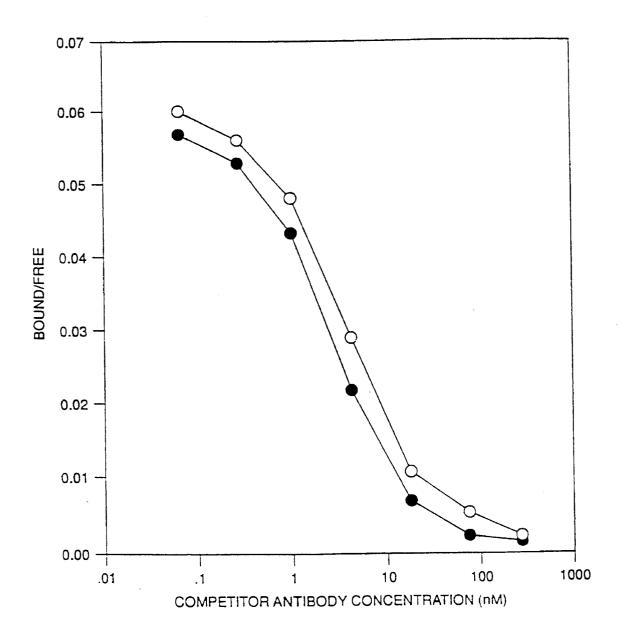


FIGURE 28

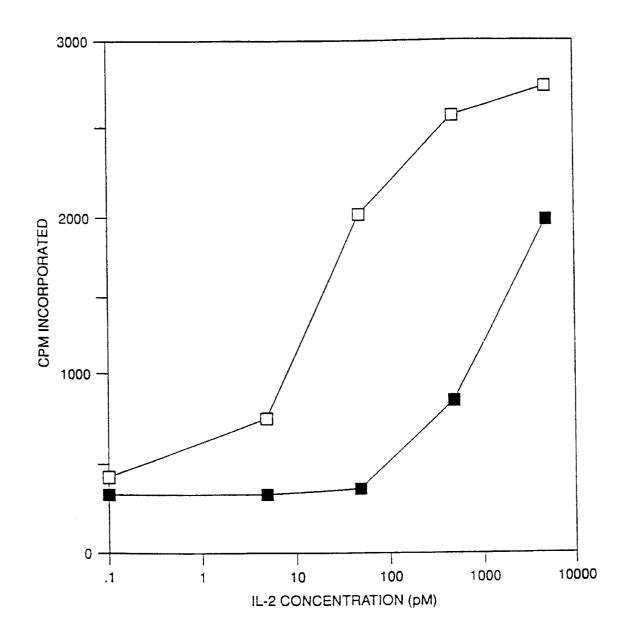


FIGURE 29

FIGURE 30B

FIGURE 30D

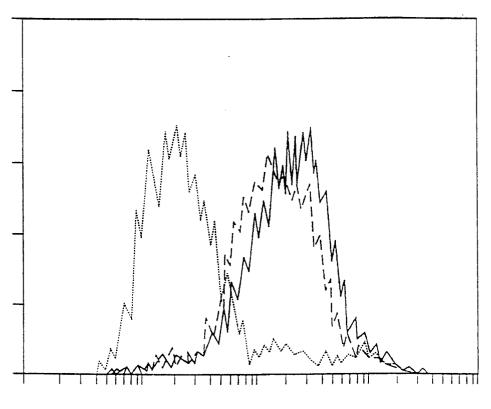


FIGURE 31A

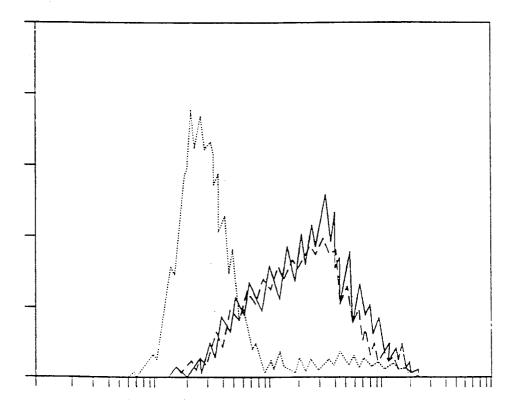


FIGURE 31B

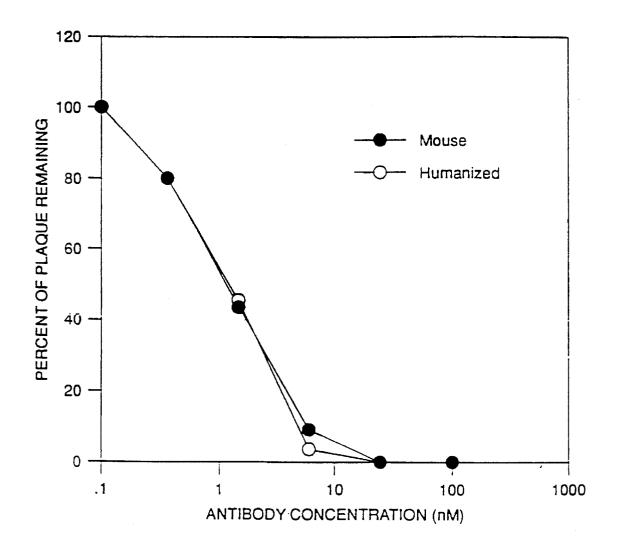


FIGURE 32A

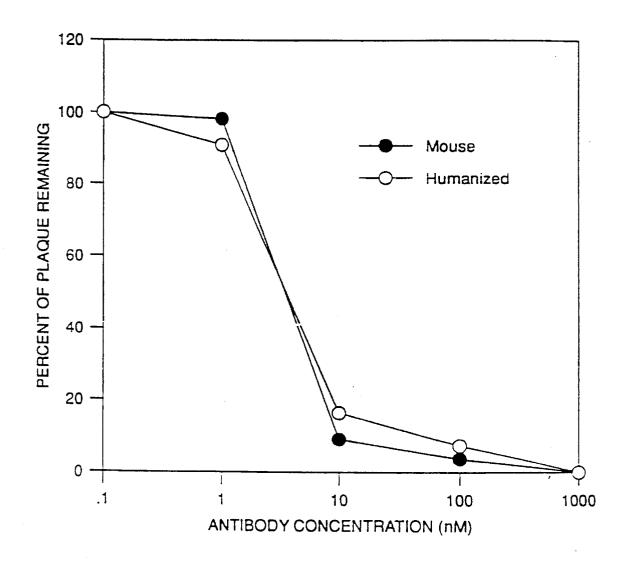


FIGURE 32B

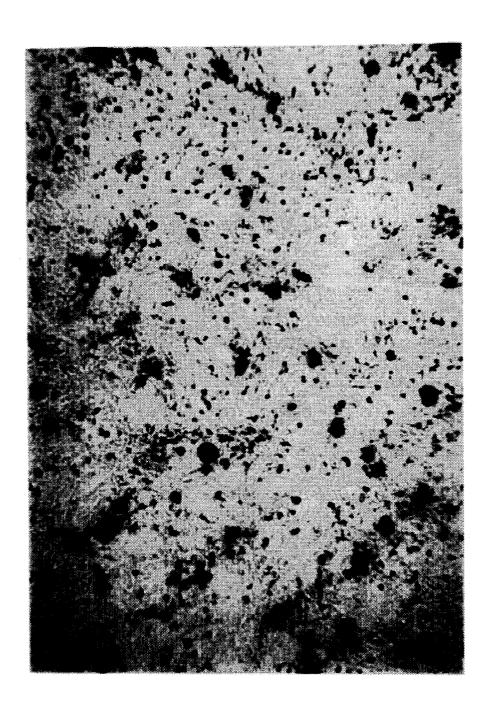


FIGURE 33A





• 30	
ATGGAGAAAGACACTCCTGCTATGGGTC M E K D T L L W V	L L L W V P G S T G
• • 90	
GACATTGTGCTGACCCAATCTCCAGCTTCT D I V L T Q S P A S	TTGGCTGTCTCTAGGGCAGAGGGCCACC L A V S L G Q R A T
• • 150	• 180
ATCTCCTGCAGAGCCAGCGAAAGTGTTGAT	TAATTATGGCATTAGTTTTATGAACTGGTTC NYGISFMNWF
• • 210	
CAACAGAAACCAGGACAGCCACCCAAACTC	CCTCATCTATGCTGCATCCAACCAAGGATCC L I Y A A S N Q G S
• • 270	• • 300
	STCTGGGACAGACTTCAGCCTCAACATCCAT S G T D F S L N I H
• • • 33	
CCTATGGAGGAGGATGATACTGCAATGTA'	TTTCTGTCAGCAAAGTAAGGAGGTTCCGTGG
P M E Ë D D T A M Y	
39 ACGTTCGGTGGAGGCACCAAGCTGGAAAT	=
T F G G G T K L E I	K IDE 244
FIGU	RE 34A
	60
• • 3 ATGGGATGGAGCTGGATCTTTCTCTTCCT	0 • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG
	0 • 60
ATGGGATGGAGCTGGATCTTTCTCTTCCT M G W S W I F L F L • 9	0 • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG L S G T A G V H S E 0 • 120
ATGGGATGGAGČTGGATCTTTCTCTTCCT M G W S W I F L F L • 9	0 • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG L S G T A G V H S E 0 • 120
ATGGGATGGAGCTGGATCTTTCTCTTCCT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L 15	O • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG L S G T A G V H S E O • 120 GGTGAAACCTGGGGCCTCAGTGAAGATATCC V K P G A S V K I S O • 180
ATGGGATGGAGCTGGATCTTTCTCTTCCT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA	0 • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG L S G T A G V H S E 0 • 120 GGTGAAACCTGGGGCCTCAGTGAAGATATCC V K P G A S V K I S
ATGGGATGGAGCTGGATCTTTCTCTTCTT M G W S W I F L F L O STCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D O STCCAGCTTCTGGATACACATTCACTGA C K A S G Y T F T D	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTCTT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D GGAAAGAGCCTTGAGTGGATTGGATATAT	O • 60 CCTGTCAGGAACTGCAGGCGTCCACTCTGAG L S G T A G V H S E O • 120 GGTGAAACCTGGGGCCTCAGTGAAGATATCC V K P G A S V K I S O • 180 CTACAACATGCACTGGGTGAAGCAACAT Y N M H W V K Q S H
ATGGGATGGAGCTGGATCTTTCTTCCT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D GGAAAGAGCCTTGAGTGGATATAT G K S L E W I G Y I 27	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTTCTT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D GGAAAGAGCCTTGAGTGGATTGGATATAT G K S L E W I G Y I CAGAAGTTCAAGAGCAAGGCCACATTGAC	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTTCTT M G W S W I F L F L O	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTCTT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D GGAAAGAGCCTTGAGTGGATTGGATATAT G K S L E W I G Y I CAGAAGTTCAAGAGCAAGGCCACATTGAC Q K F K S K A T L T GACGTCCGCAGCCTGACATCTGAGGACTC	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTCTT M G W S W I F L F L O	O O O O O O O O O O O O O O O O O O O
ATGGGATGGAGCTGGATCTTTCTTCCT M G W S W I F L F L GTCCAGCTTCAGCAGTCAGGACCTGAGCT V Q L Q Q S G P E L TGCAAGGCTTCTGGATACACATTCACTGA C K A S G Y T F T D GGAAAGAGCCTTGAGTGGATTGGATATAT G K S L E W I G Y I CAGAAGTTCAAGAGCAAGGCCACATTGAC Q K F K S K A T L T GACGTCCGCAGCCTGACATCTGAGGACTC D V R S L T S E D S GCTATGGACTACTGGGGTCAAGGAACCTC	O O O O O O O O O O O O O O O O O O O

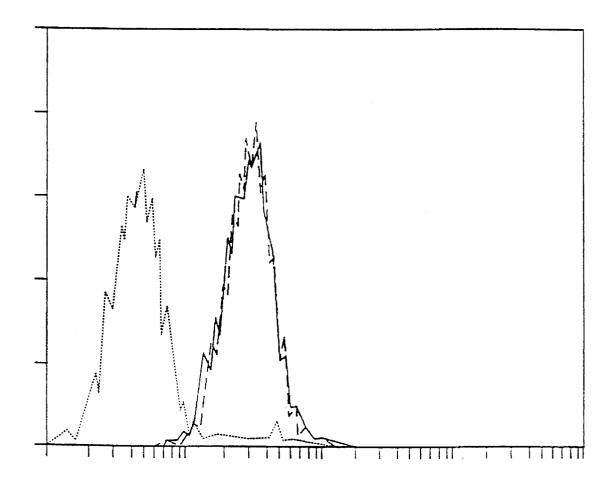


FIGURE 35

1	D D	I I	Q Q	M M	T T	QQ	S S	P P	S S	T S	L L	S S	A A	S S	V V	G . G			V	T T
21 21	I		C C	R R	A A	s s	Q E	S S	v	I D	N N	Ā	G	I	T S			A N	W W =	Y E
37 41	Q Q	Q Q	K K	P P	G G	G G	A A	P P	K K	L L	L L	М <u>І</u>	Y Y	К <u>А</u>			S N	L Q		S S
57 61	G G	V V	P P	S S	R R	F	I S	G G	S S	G G	s s	G G	T T	E D	F	T	L L	Ť	I I	S S
77 81	s s	L L	Q Q	P P	D D	D D	F F	A A	T T	Y Y	Y Y	C C	Q Q	Q Q	Y S	N K	S E	D V	S P	к <u>w</u>
97 101	M T	F	G G	Q Q	G G	T T	K K	V V	E E	V <u>I</u>	K									

FIGURE 36A

1	Q Q		QQ	L L	V V	Q Q	S S	G G	A A	E E	V V	K K	K K	P P	G G	S S		V V		A A
21 21	S S	СС	K	A A	S S	G G	G Y	T T	F F F	S T	R D	S Y	A N	I M	I <u>H</u>		V V	R R		A A
4 <u>1</u> 4 <u>1</u>	יט יט	G G	QQ	GG	.1.1	E	W	М <u>І</u>	G G	G Ÿ	I	Ā Ā	ם מ	M 7	F N	G G	p, G	טייני	И G	ř Ž
61 61	A N	Q O						V A										T T	A A	
81 81	M M	E E	L	S S	S S	L L	R R	S S	E E	D D	T T	A A	F V	. Ā	E y	C	A A	G R	_	Y —
101 100	G R	Į.	Y A	S M	D D	E	E W	Y G	<u>о</u>	G G	G E	_ L	A A	14.1	7	S S	S			

FIGURE 36B

mal

20 30 40 50 60 TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCT 80 90 100 110 120 GGCGTCCACT CTCAGGTTCA GCTGGTGCAG TCTGGAGCTG AGGTGAAGAA GCCTGGGAGC 130 TCAGTGAAGG TT

ma2

20 30 40 50 80 90 100 110 120 CCTGCCTCAC CCAGTGCATG TTGTAGTCAG TGAAGGTGTA GCCAGAAGCT TTGCAGGAAA 130 CCTTCACTGA GCT

ma3

20 30 40 50 TGGTGGTACC GGCTACAACC AGAAGTTCAA GAGCAAGGCC ACAATTACAG CAGACGAGAG 30 90 100 110 TACTAACACA GCCTACATGG AACTCTCCAG CCTGAGGTCT GAGGACACTG CA

ma4

10 20 30 40 50 60 TATATCTAGA GGCCATTCTT ACCTGAAGAG ACAGTGACCA GAGTCCCTTG GCCCCAGTAG 80 90 100 TOCATAGOGG GGOGGCCTOT TGCGCAGTAA TAGACTGCAG TGTCCTCAGA C

FIGURE 37A

ma5 10 20 30 40 50 60 TATATCTAGA CCACCATGGA GAAAGACACA CTCCTGCTAT GGGTCCTGCT TCTCTGGGTT 70 80 90 100 110 CCAGGTTCCA CAGGTGACAT TCAGATGACC CAGTCTCCGA GCTCTCTGTC CGCATCAGTA GG ma6 10 20 30 40 50 60 TCAGAAGCTT AGGAGCCTTC CCGGGTTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 70 80 90 100 110 120 AATTGTCGAC ACTTTCGCTG GCTCTGCATG TGATGGTGAC CCTGTCTCCT ACTGATGCGG AC ma7 10 20 30 40 50 TCCTAAGCTT CTGATTTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 70 80 90 100 110 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT ma8 10 20 30 40 50 TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 100 90 TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG

FIGURE 37B

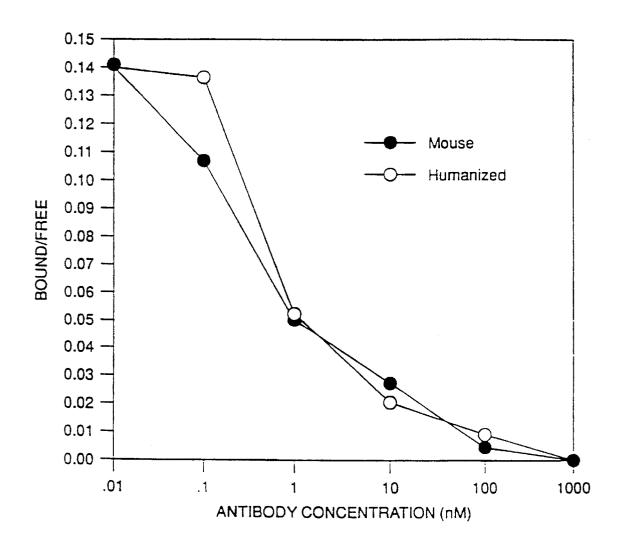


FIGURE 38

	ATG M	GTT V		ACA			ATA	_		30 CTT.		CTT	TTI	TGG	ATT	TCA			AGF	60 AGGT
1	141	٧	Ē	1.	D.	Q	Ī	L	G	L	M	L	F	W	-	S	A	S	R	G
¥		ATT		CTA		CAG			.GCC	90 32A:	CTG		GTG	·	CCG	GGA	• GAT	AGC	GTC	120 CAGT
	D	-	V	L	Ţ	Q	S	Đ	A	T	L	S	V	T	5	G	D	S	V	S
	CTT	TCC	TGC	• AGG	GCC	AGC	CAA	AGT	ATT	150 'AGC	AAC	AAC	CTA	• .CAC	TGG	TAT	CAA	CAA	AAZ	180 TCA
	L	S	С	R	A	S	0	S	Ţ	S	N		L		W	Y	Q	Q	K	S
	CAT	GAG'	тст	• CCA	AGG:	اناستات	_ •	ኔ ጥ ር	አአር	210	CC^{m}	Tr.C.C.	C N C	•	ካ ጥር	ىلىپ ئىل	•	አ m c		240 TCC
	Н		s	P	R	L	L	I	K		A	S		S		S	G	I	P	S
	7.00	mme.	3 Cm				•			270				•			•			300
	R		S	GGC2 G	AGT(S	GGA G	TCA S	GGG. G	ACA T	GAT D	TTC. F	ACT T	CTC L	AGT S	GTC. V	AAC N	GGT G	GTG V	GAC E	ACT T
				•			•			330				•			•			360
	GAA(GAT' D	TTT E	GGAI G	ATG: M	TAT' Y	TTC F	TGT C	CAA Q	CAC.		AAC N		TGG W	CCT	CAT H	ACG T	TTC F	GGA G	G G
	GGG2	ACCI T	AAG K	CTG(GAA/ E	ATAI	AAA K													
				_	_	-	••		FI	GU	DI	7 20	Λ.							
										U U	I	יכ ב	УA							
				•			•			30	KI	יכ בי	ЭΑ	•			•		į	60
	ATG(GGA'	TGG. W	• AGC: S	rgg2 W	ATC:	TTT F	CTC				TCA	GGA	•			• GTC V			GAG
								_	TTC	30 CTC L	CTG			• ACT	GCA A	GGT G		CAC H	TCI S	GAG E
	M	G CAG	W	S · CAA	W CAG	I CT	F • GGA	L CCT	TTC F GAG	30 CTC L 90	CTG L GTG.	TCA(S AAG(GGA G CCT	ACT T	A GCT'	G TCA	V • ATG			CGAG E 120
	М	G	W	s •	W			L	TTC F	30 CTC L 90 CTG	CTG L	TCA(S	GGA G	ACT T						CGAG E 120 TCC S
	M GTC: V TGC:	G CAG(Q AAG(W CTG L GCT	S CAAC Q TCTC	W CAG Q	I CTC S	F GGA G	L CCT	TTC F GAG E	30 CTC L 90 CTG L	CTG L GTG. V	TCAG S AAGG K	GGA G CCT P	ACT T GGA G	A GCT' A	G TCA S	V ATG. M	H AAG. K	S ATA I	CGAG E 120 TCC S
	M GTC: V TGC:	G CAG Q	W CTG L GCT	S CAAC Q TCTC	W CAG Q	I CTC S	F GGA G	L CCT P	TTC F GAG E ACT	30 CTC L 90 CTG L 150 GGC	CTG L GTG. V	TCAG S AAGG K	GGA G CCT P	ACT T GGA G	A GCT' A	GTG	V ATG M	H AAG. K CAG.	S ATA I	CGAG E 120 ATCC S 180 CCAT
	M GTCO V TGCA C	G CAGO Q AAGO K	W CTG L GCT A	S CAAC Q TCTC S CTTC	W DAGI Q STTI	TACTO	F GGA G TCA S	CCT P TTC. F	TTC F GAG E ACT	30 CTC L 90 CTG L 150 GGC	CTG L GTG. V IAC.	TCAG S AAGG K ACCI	GGA G CCT P ATG M	ACT T GGA G AAC N	GCT' A TGG' W	TCA S GTG. V	V ATG. M AAG K	H AAG. K , CAG. Q	ATA I AGC	120 TCC S 180 CAT H
	M GTCO V TGCA C	G CAGO Q AAGO K	W CTG L GCT A	S CAAC Q TCTC	W DAGI Q STTI	TACTO	F GGA G TCA S	CCT P TTC. F	TTC F GAG E ACT	30 CTC L 90 CTG L 150 GGC	CTG L GTG. V IAC.	TCAG S AAGG K ACCI	GGA G CCT P ATG M	ACT T GGA G AAC N	GCT' A TGG' W	TCA S GTG. V	V ATG. M AAG K	H AAG. K , CAG. Q	ATA I AGC	120 TCC S 180 CAT H
	M GTC: V TGC! C GGA:	G CAGO Q AAGO K CAGO	W CTG L GCT A	S CAAC Q TCTC S CTTC	W Q GTTT V GAGT	I S IACI Y IGGI W	GGAGGGATCA	CCTOPTC.	TTC F GAG E ACT T CTT	30 CTC L 90 CTG L 150 GGC 210 ATT	CTG L GTG. V	TCA(SAAGGAKKACCI	GGA GCCT PATG M	ACT T GGA G AAC N AAT	A GCT A TGG W	G S GTG V	V ATG. M AAG. K ACT.	H AAG. K CAG. Q AGC.	S ATA AGC S	120 TCC S 180 CAT H 240 AAC N
	M GTCO V TGCA C	G CAGO AAGO K CAGO AAGO	W CTG L GCT A AACO	CAAC Q TCTC S CTTC	W CAGG	I CCTC S IAC: Y	F GGAGG G G G G G G G G G G G G G G G G	L CCT P TTC: F GGA(G	TTC F GAG E ACT T CTT	30 CTC L 90 CTG L 150 GGC 210 ATT	CTG L GTG. V IAC. Y	TCA(S AAGG K ACCI T	GGA GCT P ATG M TAC	ACT T GGA G AAC N AAT TCA	GCT' A TGG(W GGT(G TCA S GGTG V	AAG AAG ACT	H AAG. K CAG. Q AGC.	S ATA AGC S TAC	120 120 1CC S 180 CAT H 240 AAC N 300 ATG
	M GTCG V TGCA C GGAGA Q	G CAGG Q AAAGG K CAGA K	W CTG L GCT' A AACO N ITC:	CAAC Q • TCTC S • CTTC • AAGC	W CAGE C STTT V SAGE E G G G G W	I CCTC S TACT Y CGGA W	GGA GGA GGA	CCTTC. F GGAG G ACAT	TTC F GAG E ACT T TTA L	30 CTC L 90 CTG L 150 GGC 210 ATT 1 270 ACT	CTG L GTG. V IAC. Y AAT? N	TCA(S AAGG K ACCI T CCTT GACI D	GGA GCT P ATG M TAC Y AAAG K	ACT GGA G AAC N AAT N TCA	A GCTT A TTGGG W	G TCA S G G G AAAC.	V ATG. M AAAG K ACT. T ACA	H AAAG. K CAG. Q AGC. S	ATA AGC S TAC	120 120 120 180 180 180 180 180 180 180 180 180 18
	M GTCG V TGCA C GGAG G	G CAGO Q AAAGO K CAGO K AAGO K	W CTG L GCT' A AACO N FTC:	CAAC Q • TCTC S • CTTC • AAGC	W CAGO CAGO STTT V SAGI	ICTO S IACO Y CGG: W	F . GGA . TCA . TCTC	L CCTI P TTC: F GGA' G G G G G G G G G G G G G G G G G	TTC F GAG E ACT TTA L GAC	30 CTC L 90 CTG L 150 GGC 210 ATT I 270 ACT T	CTG L GTG. V IAC. Y AAAT(N STA(V	TCA(S AAGG K ACCI T CCTT CCTT CACCI CCTT CCTT	GGA GCT P ATG M TAC Y AAGG K	ACT GGA G AAC N AAT N TCA S	A GCT A TGGG G G TGCC	G TCA S GTG V GGT. N ACA	V ATG. AAAGK ACT. ACAACA	H AAAG K CAGG Q AGCC S GCCC A	ATA AGC S TAC Y	120 120 1CC S 180 CAT H 240 AAC N 360 ATG
	M GTCG V TGCA C GGAGA Q GAGA	G CAGO Q AAAGO K CAGO AAGO CTCO	W CTG L GCT A AACO N FTC:	CAAC Q TCTC S CTTC AAGC AGTC	W CAGO	I SCTO S PACO W AAGO K	GGA . CTG	CCTI P TTC. F GGAACA'	TTC F GAG E ACT TTA L GAC	30 CTC L 90 CTG L 150 GGC 210 ATT 270 ACT S 330 TCT	CTG L GTG. V IAC: Y AAATO V GTAO V	TCA(S AAGO K ACCI T CCTT GACI GACI V	GGA G CCT P ATG M TAC Y AAG K	ACT GGA AAC AAT TCA TCA TAC	GGTCGG	G TCA S GTG. V GGT. N AACA	V ATG. AAGG. K ACT. ACA. T AGGA. AGA.	H AAAG. CAG. Q AGC. S GCC. A CGGG.	ATA AGC S TAC Y	120 120 1CC S 180 CAT H 240 AAC N 360 ATG
	M GTCG V TGCA C GGAGA Q	G CAGG	W CTG L GCT A AACO N ITC. E CTC:	CAAC Q TCTC S CTTC AAGC AGTC	W CAGO	I CTO S FACT Y LAGO K LCAT	GGA OTO	CCTP TTC. F GGA ACA TGGG	TTC GAG E ACT TTA GAC GGT GGT GGT	30 CTC L 90 CTG L 150 GGC 210 ATT 270 ACT S 330 TCT	CTG L GTG. V IAC. Y AATO V GTAO A	TCA(S AAGG K ACCI T GACI GACI T ACCI V ACCI	GGA GCT P ATG M FAC Y AAG K TAT	ACT GGA AAC AAT TCA TCA TAC	GGTCGG	G TCA S GTG. V GGT. N AACA	V ATG. AAGG. AAGT. AAGA. T AAGA. T CC	H AAGG CAGG Q AGCC S GCCC A CGGG TCA	ATA AGC S TAC Y	120 120 1CC S 180 CAT H 240 AAC N 360 ATG

1	E			Ŀ	T	Q	S	ρ.	G G	T	L L	S	Ľ	S S	P P	G G	E	R R	A A	T
1	Ε	I	V	L	T	Q	S	P	G	T	سل	۵	L	3	Ę	G	5.	ĸ	A	Т
21 21	L L	s s	C	R R			Q Q	S S	V		S S	G N		L L	G H	W	Ž Ž	Q Q	Q Q	K K
41 40	P P	G G	Q Q	A A	P P	R R	L L	L L	I	Y K	G Y	A A		S O	R S	A			I	
61 60	D D	R R	F	S S	G G	S S	G G	s s	G G	T T	D D	11 II	T T	L L	T T	I I	s s	R R	L L	E E
81 80	P P	E E	D D	F F	A A	V V		Y Y	C	Q	Q 0	Y S	G N	S S	L W	Gρ	R H	ŭ T	H H	G G
101 100	Q Q	G G	T T	K K	V V	E E	I I	K K												

FIGURE 40A

```
P G K G L E W P G K G L E W
                       V
V
                                        S F
S F
             S V V R V S V
F K G R V T V
                              S L K
S L K
61
     N P G
     <u>N O K</u>
                       S E D T
S E D T
18
     G F D T S D Y Y Y Y W G Q G T L V T V G F R D Y S M D Y W G Q G T L V T V
101
100
121
      S
        S
      S S
118
```

FIGURE 40B

jb16 10 20 30 40 50 60 TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTAG GACTCATGCT CTTCTGGATT 70 80 90 100 110 120 TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG GGAGAAAGG jb17 10 20 30 40 50 60 TAGACAGAAT TCACGCGTAC TTGATAAGTA GACGTGGAGC TTGTCCAGGT TTTTGTTGGT 70 80 90 100 110 120 ACCAGTGTAG GTTGTTGCTA ATACTTTGGC TGGCCCTGCA GGAAAGTGTA GCCCTTTCTC CCGGTGAT jb18 20 30 40 AAGAGAATTC ACGCGTCCCA GTCCATCTCT GGAATACCCG ATAGGTTCAG TGGCAGTGGA 80 90 100 TCAGGGACAG ATTTCACTCT CACAATAAGT AGGCTCGAGC CGGAAGATTT TGC jb19 20 30 40 50 TAGATCTAGA GTTGAGAAGA CTACTTACGT TTTATTTCTA CCTTGGTCCC TTGTCCGAAC

FIGURE 41A

GTATGAGGCC AACTGTTACT CTGTTGACAA TAATACACAG CAAAATCTTC CGGCTC

70 80 90 100

jb20 10 20 30 40 50 60 TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCA 70 80 90 100 110 GGTGTCCACT CTCAAGTCCA ACTGGTACAG TCTGGAGCTG AGGTTAAAAA GCCTGGAAGT TCAGTAAGAG TTTC jb21 20 30 40 50 60 TATATAGGTA CCACCATTGT AAGGATTAAT AAGTCCAACC CACTCAAGTC CTTTTCCAGG 90 110 TGCCTGTCTC ACCCAGTTCA TGGTATACCC AGTGAATGAG TATCCGGAAG CTTTGCAGGA 130 AACTCTTACT GAAC

jb22

10 20 30 40 TATATAGGTA CCAGCTACAA CCAGAAGTTC AAGGGCACAG TTACAGTTC TTTGAAGCCT 70 80 90 100 TCATTTAACC AGGCCTACAT GGAGCTCAGT AGTCTGTTTT CTGAAGACAC TGCAGT

jb23

10 20 30 40 50 TATATCTAGA GGCCATTCTT ACCTGAGGAG ACGGTGACTA AGGTTCCTTG ACCCCAGTAG 90 100 110 80 TCCATAGAAT AGTCTCGAAA CCCCCGTCTT CTACAGTAAT AGACTGCAGT GTCTTC

FIGURE 41R

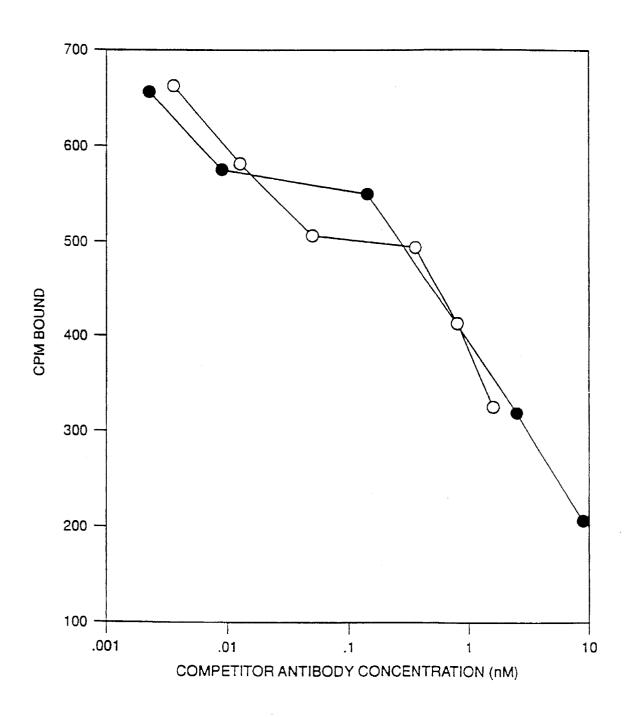


FIGURE 42

			•			•			30				•			•			60
ATG	CAT	CAG	ACC	AGC	ATG	GGC	ATC	AAC	ATG	GAA	TCA	CAG	ACT	CTG	GTC	TTC	ATA	TCC	ATA
M	H	Q	\mathbf{T}	S	M	G	I	K	M	Ε	S	0	T	L	V	F	Ţ	S	T
		~		-		_	_			_	Ū	*	_	_		_	_	•	-
			•						90										120
CTG	\sim	ሞሮር	א נוזינה	an sean	CCM	ccm	~ a m	~~~				3 m c		~ A A	m C m	-	בבב	m ~ ~	
T.	C 1 C		TIM				CAT		AAC	ATT				CAA				TCC	ATG
ب	1	W	L	Y	G	A	D	G	N	I	V	M	T	Q	S	Ρ	K	S	М
			•			•			150				•			•			180
TAC	GTG	TCA	ATA	.GGA	GAG.	AGG	GTC	ACC	CTTG	AGC	TGC	AAG	GCC	AGT	GAA	AAT	GTG	GAI	ACT
Y	V	S	I	G	E	R	V	T	L	S	С	K	Α	S	Ε	N	V	D	T
			•			•			210							•			240
TAT	ста	TCC	TCC	יי איי	(A A	$C \lambda C$	אאא	CC^{2}	GAG	~ > ~	т Ст	\sim	מממי	CTG	СТ С	АТА	ТАТ		GCA
V	V		W	A											- L G				
_ _		<u>_S</u>	W	ĭ	Q	Q	K	P	E	Q	S	P	K	L.	ىك .	Ι	Y	<u>G</u>	<u>A</u>
			•			•			270				•			•			300
TCC	AAC	CGG	TAC	ACT	GGG	GTC	CAC	GA1	CGC	TTC	ACG	GGC	AGI	'GGA	TCT	'GCA	ACA	GAI	TTC
_ <u>S</u>	N	R	Y	T	G	V	H	D	R	F	Ţ	G	S	G	S	A	Ţ	D	F
												-							
			•			•			330										360
ACT	CTC	ACC	ልጥር	אכר	707	\sim m \sim	$C \lambda C$	-	'GAA	C N C	ئىس ىك	-	GAT	ጥልጥ	CAC	ىك ب	CC 3	$C \times C$	AGT
Ţ	T,	7100	- T	S	S	U 17					LIJ.			IAI		101			
-		*	<u> </u>	٥	5	V	Q	A	E	D	1	A	D	ž	Η	ن	<u>G</u>	Q	S
			•			•			390				•						
TAC	AAC	TAT	CCA	TTC	ACG	TTC	GGC	TCG	GGG:	ACA	AAG	TTG	GAA	ATA	AAG	;			
Ϋ́	N	Y	P	F	Ţ.	Ē	G	S	G	Т	K	L	Ε	Ξ	K				
							_	_	. –	-		_	_	_					

FIGURE 43A

			•			•			30				•			•			60
ATG	ACA	TCA	CTG'	TTC	CTC	CTA	CAG	TTA	.CCG2	AGC.	ACA	CAG	GAC	CTC	GCC.	ATG	GGA'	TGG	AGC
M	T	S	L	F	S	L	Q	L	P	S	T	Q	D	L	A	M	G	W	S
			•			•			90				•			•			120
TGT		ATC	CTC	TTC:	rtgo	STA	GCA	ACA	GCT	ACA	GGT	GTC	CTC	TCC				CTG	CAG
С	I	Ι	L	F	L	V	A	T	A	T	G	V	L	S	Q	V	Q	L	Q
C3 C4		~~~	•			•			150				•	C.E.C	mcc	•		~ ~ ~	180
				GAC	CTTC				GGG				AAG	CTG	TCC	TGC		<u>ت</u> ت	TCI
Q	P	G	A	D	L	V	M	Ď	G	A	P	V	K	سن	S	С	÷	A	S
			•			•			210				•			•			240
GGC:	rac.	ATC	TTC	ACC	AGC:	ICC'	TGG.	ATA	AAC'	rgg	GTG	AAG	CAG	AGG	CCT	GGA	CGA	GGC	CTC
G	Y	I	F	T	<u>s_</u>	S	W	I	N	W	V	K	Q	R	P	G	R	G	L
			•			•			270				•			•			300
GAG:		ATT	• GGA	AGG:	ATT	• GAT	CCT		270 GAT	GGT	GAA						GAT		AAG
GAG	IGG. W	ATT I	• GGA G	AGG	ATT(• GAT	CCT P	TCC S		GGT G	GAA E	GTT V	· CAC H	TAC Y	AAT N	CAA Q	GAT D	TTC F	• • •
					ATT(GAT										AAG
	W	Ī	G •	R	ATT(D ·	P	S	GAT(G	Е		<u>н</u> •				D	F	:AAG _K
Ε	W	Ī	G •	R	I	D ·	P	S	330	G	Е	V.	<u>н</u> •				D	F	360
E GAC	W AAG	I GCC	G •	R	I ACT(D GTA	P GAC	S AA <i>P</i>	GATO D 330	G TCC	E AGC	V.	H .GCC			Q • CAA	D	F AAC	AAG K 360 AGC
E GAC	W AAG K	GCC A	G ACA T	R CTG. L	I ACT(D GTA	gac D	S AAA K	330 ATCC	G TCC S	E AGC	ACA T	H .GCC			Q • CAA	D	F AAC	AAG K 360 AGC S 420
GAC	W AAG K	GCC A	G ACA T	R CTG. L	I ACT(D • GTA V	gac D	S AAA K	330 ATCC S	G TCC S	E AGC	ACA T	H GCC A			Q • CAA	D	F AAC	360 CAGC S
E GACI	W AAG K ACA T	GCC A TCT S	G ACA T GAG E	R CTG. L GAC	ACTO	D GTA V GCG A	GAC D GTC V	AAA K TAT	330 ATCC S 390 TAC Y 450	G TCC S TGT C	E AGC S GCT A	ACA T AGA R	H .GCC A	TAC Y	N ATC	Q CAA Q	OTC L	F AAC N	AAG K 360 AGC S 420
GAC	W AAG K ACA T	GCC A TCT S	G ACA T GAG E	R CTG. L GAC	ACTO	D GTA V GCG A	GAC D GTC V	AAA K TAT	330 ATCC S 390 TAC	G TCC S TGT C	E AGC S GCT A	ACA T AGA R	H .GCC A	TAC Y	N ATC	Q CAA Q	OTC L	F AAC N	AAG K 360 AGC S 420

FIGURE 43B

1	D D	I	Q Q	M M	T	Q Q	S S	D, D,	s s	T T		S S			V V		D D		V	T T
21 21	I	T T		R K	A A	S S	Q E	S N	I V	N D	T Ţ	W Y	L V	A S	W	Y Y	Q Q	Q Q	K K	P P
41 41	G G	K K		P P		L L	L L	M <u>I</u>	Y Y	K G	A A	S S	ร ห		E Y		G G	V V	P. P.	s s
61 61		F F			S S	G G	S S	G	T T	E D	F F	T T	L L	T T	I	S S	S S	L L	Q Q	ρ, ρ,
81 81	D D	D D	F F	A A	T T	Y Y	Y Y	C	G	Q Q	Q S	Y Y	N N	S		S P	K F	M T	F	G G
100 100	Q Q	G G		K K		E E		K K												

FIGURE 44A

```
1
21
 S C K A S G G T F S R S A I
S C K A S G Y I F T S S W I
21
 41
41
61
 61
81
 81
 101
101
```

FIGURE 44B

rh10

10	20	30	40	50	60
TTTTTTCTAG	ACCACCATGG	AGACCGATAC	CCTCCTGCTA	TGGGTCCTCC	TGCTATGGGT
70	80	90	100	110	Стест
CCCAGGATCA	ACCGGAGATA	TTCAGATGAC	CCAGTCTCCG	TCGACCCTCT	

rhll

10	20	30	40	50	60
TTTTAAGCTT	GGGAGCTTTG	CCTGGCTTCT	GCTGATACCA	GGATACATAA	GTATCCACAT
70	80	90	100	110	120
TTTCACTGGC	CTTGCAGGTT	ATGGTGACCC	TATCCCCGAC	GCTAGCAGAG	AGGTTCCACG

rh12

10	20	20	40	50	60
TTTTAAGCTT	CTAATTTATG		CCGGTACACT	GGGGTACCTT	CACGCTTCAG
70 TGGCAGTGGA	80 TCTGGGACCG	90 ATTTCACCCT	±00		CAGATGAT

rh13

10	20	30	40	50	60
TTTTTTCTAG	AGCAAAAGTC	TACTTACGTT	TGACCTCCAC	CTTGGTCCCC	TGACCGAACG
70 TGAATGGATA	80 GTTGTAACTC	90 TGTCCGCAGT	100 AATAAGTGGC		

FIGURE 45A

rh20

10 20 30 40 TTTTTCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGTACCGCG 70 80 90 100 GGCGTGCACT CTCAGGTCCA GCTTGTCCAG TCTGGGGCTG AAGTCAAGAA ACCT

rh21

10 20 30 40 50 60 TTTTGAATTC TCGAGACCCT GTCCAGGGGC CTGCCTTACC CAGTTTATCC AGGAGCTAGT 80 90 100 AAAGATGTAG CCAGAAGCTT TGCAGGAGAC CTTCACGGAG CTCCCAGGTT TCTTGACTTC

Α

rh22

20 30 40 50 60 TTTTGAATTC TCGAGTGGAT GGGAAGGATT GATCCTTCCG ATGGTGAAGT TCACTACAAT 70 80 90 100 110 120 CAAGATTICA AGGACCGTGT TACAATTACA GCAGACGAAT CCACCAATAC AGCCTACATG GAACTGAGCA GCCTGAG

rh23

20 30 40 50 60 TTTTTCTAGA GGTTTTAAGG ACTCACCTGA GGAGACTGTG ACCAGGGTTC CTTGGCCCCA 70 80 90 100 110 120 GTCAGCAAAC CAGGGCAGAA ATCCTCTTGC ACAGTAATAG ACTGCAGTGT CCTCTGATCT 130 CAGGCTGCTC AGTT

FIGURE 45B

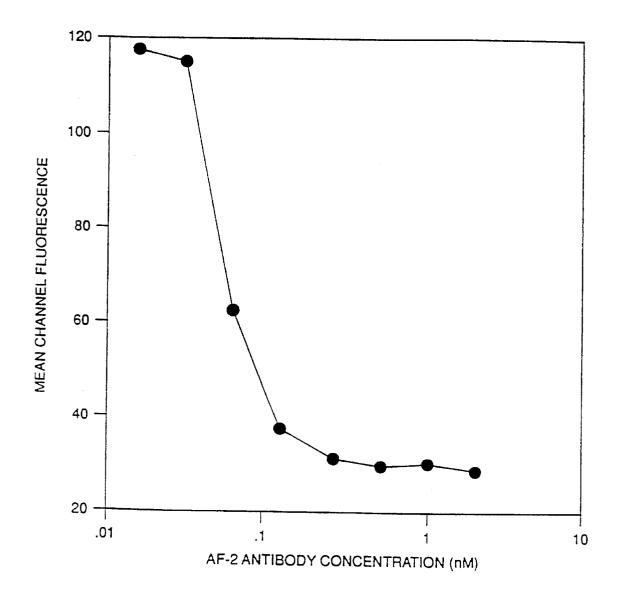


FIGURE 46

HUMANIZED IMMUNOGLOBULINS

CROSS-REFERENCE TO RELATED APPLICATIONS

This is a continuation of application Ser. No. 07/634,278, filed Dec. 19, 1990, U.S. Pat. No. 5,530,101, which is a continuation-in-part application of commonly assigned patent application U.S. Ser. No. 07/590,274, filed Sep. 28, 1990, (now abandoned) and of U.S. Ser. No. 07/310,252, 10 filed Feb. 13, 1989, (now abandoned), which is a contination-in-part of U.S. Ser. No. 07/290,975, filed Dec. 28, 1988, (now abandoned). All of these applications are specifically incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates generally to the combination of recombinant DNA and monoclonal antibody technologies for developing novel therapeutic agents and, more particularly, to the production of non-immunogenic antibodies having strong affinity for a predetermined antigen.

BACKGROUND OF THE INVENTION

The advent of monoclonal antibody technology in the mid 1970's heralded a new age of medicine. For the first time, researchers and clinicians had access to essentially unlimited quantities of uniform antibodies capable of binding to a predetermined antigenic site and having various immunological effector functions. These proteins, known as "monoclonal antibodies" were thought to hold great promise in, e.g., the removal of harmful cells in vivo. Indeed, the clinical value of monoclonal antibodies seemed limitless for this use alone

Unfortunately, the development of appropriate therapeutic products based on these proteins has been severely hampered by a number of drawbacks inherent in monoclonal antibody production. For example, most monoclonal antibodies are mouse derived, and thus do not fix human complement well. They also lack other important immunoglobulin functional characteristics when used in humans.

Perhaps most importantly, non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that after injection of a foreign antibody, the immune response mounted by a patient can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after one or several treatments with any non-human antibodies, subsequent treatments, even for unrelated therapies, can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. Moreover, efforts to 60 immortalize human B-cells or generate human hybridomas capable of producing human immunoglobulins against a desired antigen have been generally unsuccessful, particularly with many important human antigens. Most recently, recombinant DNA technology has been utilized to produce 65 immunoglobulins which have human framework regions combined with complementarity determining regions

2

(CDR's) from a donor mouse or rat immunoglobulin (see, e.g., EPO Publication No. 0239400, which is incorporated herein by reference). These new proteins are called "reshaped" or "humanized" immunoglobulins and the process by which the donor immunoglobulin is converted into a human-like immunoglobulin by combining its CDR's with a human framework is called "humanization". Humanized antibodies are important because they bind to the same antigen as the original antibodies, but are less immunogenic when injected into humans.

However, a major problem with present humanization procedures has been a loss of affinity for the antigen (Jones et al., Nature, 321, 522-525 (1986)), in some instances as much as 10-fold or more, especially when the antigen is a protein (Verhoeyen et al., Science, 239, 1534–1536 (1988)). Loss of any affinity is, of course, highly undesirable. At the least, it means that more of the humanized antibody will have to be injected into the patient, at higher cost and greater risk of adverse effects. Even more critically, an antibody with reduced affinity may have poorer biological functions, such as complement lysis, antibody-dependent cellular cytotoxicity, or virus neutralization. For example, the loss of affinity in the partially humanized antibody HuVHCAMP may have caused it to lose all ability to mediate complement lysis (see, Riechmann et al., Nature, 332, 323-327 (1988); Table 1).

Thus, there is a need for improved means for producing humanized antibodies specifically reactive with strong affinity to a predetermined antigen. These humanized immunoglobulins should remain substantially non-immunogenic in humans, yet be easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

SUMMARY OF THE INVENTION

The present invention provides novel methods for preparing humanized immunoglobulin chains having generally one or more complementarity determining regions (CDR's) from a donor immunoglobulin and a framework region from a human immunoglobulin. The preferred methods comprise first comparing the framework or variable region amino acid sequence of the donor immunoglobulin to corresponding sequences in a collection of human immunoglobulin chains, and selecting as the human immunoglobulin one of the more homologous sequences from the collection. The human immunoglobulin, or acceptor immunoglobulin, sequence is typically selected from a collection of at least 10 to 20 immunoglobulin variable region sequences, and usually will have the highest homology to the donor immunoglobulin sequence of any sequence in the collection. The human immunoglobulin framework sequence will typically have about 65 to 70% homology or more to the donor immunoglobulin framework sequences. The donor immunoglobulin may be either a heavy chain or light chain, and the human collection will contain the same kind of chain. A humanized light and heavy chain can be used to form a complete humanized immunoglobulin or antibody, having two light/ heavy chain pairs, with or without partial or full-length human constant regions.

To form the humanized variable region, amino acids in the human acceptor sequence will be replaced by the corresponding amino acids from the donor sequence if they are in the category

(1) the amino acid is in a CDR.

In another embodiment of the present invention, either in conjunction with the above comparison step or separately,

additional amino acids in the acceptor immunoglobulin chain may be replaced with amino acids from the CDR-donor immunoglobulin chain. More specifically, further optional substitutions of a human framework amino acid of the acceptor immunoglobulin with the corresponding amino acid from a donor immunoglobulin will be made at positions which fall in one or more of the following categories:

- (2) the amino acid in the human framework region of the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is common for that position in human immunoglobulin sequences; or
- (3) the amino acid is immediately adjacent to one of the CDR's; or
- (4) the amino acid is predicted to be within about 3 Å of the CDR's in a three-dimensional immunoglobulin model and capable of interacting with the antigen or with the CDR's of the donor or humanized immunoglobulin.

Moreover, an amino acid in the acceptor sequence may optionally be replaced with an amino acid typical for human sequences at that position if

(5) the amino acid in the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is also rare, relative to other human sequences.

The humanized immunoglobulin chain will typically comprise at least about 3 amino acids from the donor immunoglobulin in addition to the CDR's, usually at least 30 one of which is immediately adjacent to a CDR in the donor immunoglobulin. The heavy and light chains may each be designed by using any one or all three of the position criteria.

When combined into an intact antibody, the humanized light and heavy chains of the present invention will be 35 substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen (such as a protein or other compound containing an epitope). These affinity levels can vary from about 10⁸ M⁻¹ or higher, and may be within about 4 fold, preferably within about 2 fold of the donor immunoglobulin. Ideally, the humanized antibodies will exhibit affinity levels at least about 60 to 90% of the donor immunoglobulin's original affinity to the antigen.

Once designed, the immunoglobulins, including binding fragments and other immunoglobulin forms, of the present invention may be produced readily by a variety of recombinant DNA or other techniques. Preferably, polynucleotides encoding the desired amino acid sequences are produced synthetically and by joining appropriate nucleic acid sequences, with ultimate expression in transfected cells. Notably, the methods of the present invention maximize the likelihood of producing humanized immunoglobulins with optimum binding characteristics without the need for producing intermediate forms that may display stepwise improvements in binding affinity. The humanized immunoglobulins will be particularly useful in treating human disorders susceptible to monoclonal antibody therapy, but find a variety of other uses as well.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1A and FIG. 1B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:1 and 2] and heavy chain (B) [SEQ ID NOS:3 and 4] variable regions of 65 the mouse anti-Tac antibody (upper lines), compared with the human Eu antibody (lower lines), not including signal

4

sequences. The three CDR's in each chain are underlined. Residues in the Eu antibody framework replaced with mouse amino acids in the humanized antibody are double underlined. The number of the first position on each line is given on the left.

FIG. 2A and FIG. 2B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:46 and 47] and heavy chain (B) [SEQ ID NOS:48 AND 49] variable regions of the mouse Fd79 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 3A and FIG. 3B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:50 and 51] and heavy chain (B) [SEQ ID NOS:52 and 53] variable regions of the mouse Fd138-80 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 4A and FIG. 4B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:54 and 55] and heavy chain (B) [SEQ ID NOS:56 and 57] variable regions of the mouse M195 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 5A and FIG. 5B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:58 and 59] and heavy chain (B) [SEQ ID NOS:60 and 61] variable regions of the mouse mik- β 1 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 6A and FIG. 6B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:62 and 63] and heavy chain (B) [SEQ ID NOS:64 and 65] variable regions of the mouse CMV5 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 7A through FIG. 7D. Fluorocytometry of HUT-102 and Jurkat cells stained with anti-Tac antibody or humanized anti-Tac antibody followed respectively by fluorescein-conjugated goat anti-mouse Ig antibody or goat anti-human Ig antibody, as labeled. In each panel, the dotted curve shows the results when the first antibody was omitted, and the solid curve the results when the first and second (conjugated) antibodies were included as described.

FIG. 8A and FIG. 8B. (A) Fluorocytometry of HUT-102 cells stained with 0-40 ng of anti-Tac as indicated, then with biotinylated anti-Tac, and then with phycocrythrin-conju-

gated avidin. (B) Fluorocytometry of HUT-102 cells stained with the indicated antibody, then with biotinylated anti-Tac, and then with phycoerythrin-conjugated avidin.

FIG. 9A and FIG. 9B. Schematic diagram of the plasmids pVg1 (A) and pVk (B). The plasmid pVg1 was constructed 5 from the following fragments: an approximately 4850 base pair BamHI-EcoRI fragment from the plasmid pSV2hph containing the amp and hyg genes; a 630-pb fragment containing the human cytomegalovirus IE1 gene promoter and enhancer flanked at the 5' and 3' by EcoR1 and Xba1 10 linkers respectively; and a 2800 bp XbaI-BamHI fragment containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 15 gene and the gpt replacing the hyg gene.

FIG. 10A and FIG. 10B. Amino acid sequences of the heavy (A) [SEQ ID NOS:5 and 6] and light (B) [SEQ ID NOS:7 and 8] chain variable regions of the PDL and CDR-only humanized anti-Tac antibodies. The PDL ²⁰ sequence is shown on the upper line, the CDR-only sequence below. Amino acid differences are boxed. Complementarity Determining Regions (CDR's) are underlined.

FIG. 11A and FIG. 11B. Double-stranded DNA sequence of fragments encoding the heavy (A) [SEQ ID NO:9] and light (B) [SEQ ID NO:10] chain variable regions of the CDR-only humanized anti-Tac antibody including signal sequences. Oligonucleotides used for gene synthesis are marked by solid lines: above, for oligonucleotides from upper strand, and below, for oligonucleotides from lower strand. Restriction sites used for cloning are underlined.

FIG. 12. FACS analysis of HUT-102 cells stained with PDL and CDR-only humanized anti-Tac antibodies and negative control antibody Fd79.

FIG. 13. Competition by mouse, PDL humanized, and CDR-only humanized anti-Tac antibodies with binding of radioiodinated mouse anti-Tac antibody to HUT-102 cells.

FIG. 14. Scheme for anchored polymerase chain reaction (PCR) cloning of the heavy and light chain variable domain 40 cDNAs. RNA was prepared from about 10⁷ hybridoma cells using the hot phenol extraction method. Briefly, cells were resuspended and vortexed in 1 ml of RNA extraction buffer (50 mM sodium acetate pH 5.2/1% SDS), extracted with 0.5 ml of phenol pH 5.2 at 65° C. for 15 min, followed by 45 another 15 min on ice. The aqueous phase was recovered and precipitated twice with ethanol. cDNA was synthesized from 10 ug of total RNA using reverse transcriptase (BRL, Betheseda, Md.) and oligo-dT₁₂₋₁₈ (pharmacia, Piscatway, N.J.) as primers. A poly(dG) tail was attached to the 3' end 50 of the cDNA using terminal deoxynucleotide transferase (BRL) (E. Y. Loh et al., Science 243, 217 (1989)), the variable domain genes (V) were amplified using AmpliTaq (Perkin Elmer-Cetus) with the primer mc045 (TAATCTA-GAATTCCCCCCCCCCCCCCC [SEQ ID NO:11] that 55 hybridized to the poly(dG) tails and primers that hybridized to the constant region genes (C). For the light chain, the primer used was mc045 (TATAGAGCTCAAGCTTG-GATGGTGGGAAGATGGATACAGTTGGTGC) [SEO ID NO:12]. For the heavy chain, the primer used was mc047 60 (TATAGAGCTCAAGCTTCCAGTGGATA-

GAC(CAT)GATGGGG(GC)TGT(TC)GTTTTGGC) [SEQ ID NO:13]. The sequence in parenthesis indicates a base degeneracy. The degeneracy was introduced so that the primer would be able to hybridize to most gamma chains. 65 The amplified fragments were then digested with EcoRI and HindIII and cloned into pUC18 vector for sequencing.

6

FIG. 15. Comparison of sequences of anti-Tac heavy chain (upper lines) [SEQ ID NO:14] and Eu heavy chain (lower lines) [SEQ ID NO:15]. The 1-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 16. Comparison of sequences of anti-Tac light chain (upper lines) [SEQ ID NO:16] and Eu light chain (lower lines) [SEQ ID NO:17]. The single-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 17. Nucleotide sequence of the gene for the humanized anti-Tac heavy chain variable region gene [SEQ ID NOS:18 and 19]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature heavy chain sequence begins with amino acid #20 Q.

FIG. 18. Nucleotide sequence of the gene for the humanized anti-Tac light chain variable region gene [SEQ ID NOS:20 and 21]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature light chain sequence begins with amino acid #21 D.

FIG. 19A and FIG. 19B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:22, 23, 24, and 25] used to synthesize the humanized anti-Tac heavy chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide.

FIG. **20**A and FIG. **20**B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:26, 27, 28, and 29] used to synthesize the humanized anti-Tac light chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide. The position of a Hind III site in the overlap of JFD2 and JFD3 is shown.

FIG. 21. Schematic diagram of the plasmid pHuGTAC1 used to express the humanized anti-Tac heavy chain. Relevant restriction sites are shown, and coding regions of the heavy chain are displayed as boxes. The direction of transcription from the immunoglobulin (Ig) promoter is shown by an arrow. E_H =heavy chain enhancer, Hyg=hygromycin resistance gene.

FIG. 22. Schematic diagram of the plasmid pHuLTAC used to express the humanized anti-Tac light chain. Relevant restriction sites are shown, and coding regions of the light chain are displayed as boxes. The direction of transcription from the Ig promoter is shown by an arrow.

FIG. 23A and FIG. 23B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:30 and 31] and heavy chain (B) [SEQ ID NOS:32 and 33] variable regions of the antibody mik- β 1. The CDR sequences are underlined. The mature light chain protein begins with amino acid 23 Q and the mature heavy chain protein with amino acid 20 Q, preceded by the respective signal sequences.

FIG. 24A and FIG. 24B. Schematic diagram of the plasmids pVg1-dhfr (A) and pVk (B). The plasmid pVg1-

dhfr contains the following parts: an approximately 4200 base pair BamHI-EcoRI fragment containing the amp and dhfr genes; a 630-bp fragment containing the human cytomegalovirus IE1 gene promoter and enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference) flanked at the 5' and 3' ends by EcoRI and XbaI linkers respectively; and a 2800 bp Xbal-BamHI fragment containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt gene replacing the dhfr gene. The plasmids were constructed from the indicated parts using methods well-known in the art (see, Maniatis et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1989) and commonly assigned PCT Publication No. WO 89/09622, published Oct. 19, 1989. For example, pVg1-dhfr was constructed from the plasmid pVg1 by replacing the Hind III-Bgl II fragment containing the hyg gene with a 660 bp fragment containing the dhfr gene and extending to a Bgl II site (Simonsen et al., Proc. Natl. Acad. Sci. U.S.A. 80, 2495 (1983)).

FIG. 25. Fluorocytometry of YTJB cells stained with (_) Isotype matched control antibody, (---) humanized mik- β 1 antibody, (...) chimeric mik- β 1 antibody. Cells were suspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10⁶/ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with goat antihuman Ig antibody on ice for another 30 min. Then the cells were washed and incubated with FITC labeled rabbit antigoat Ig antibody for 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehyde. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 26A and FIG. 26B. Amino acid sequences of the light chain (A) [SEQ ID NOS:34 and 35] and the heavy chain (B) [SEQ ID NOS:36 and 37] of the humanized mik- β 1 antibody, (lower lines) and human Lay antibody (upper lines), not including signal sequences. The three CDRs in each chain are underlined. Amino acids in the framework that have been replaced with mouse amino acids or consensus human amino acids in the humanized antibody are double underlined.

FIG. 27A and FIG. 27B. Oligonucleotides used in the construction of the humanized mik-β1 heavy chain (B) [SEQ ID NOS:42, 43, 44, and 45] and light chain (A) [SEQ ID NOS:38, 39, 40, and 41]. The following pairs of oligonucleotides were mixed, extended with sequenase and cut with the indicated enzymes before ligation into the pBluescriptII ks (+) vector: wps54 and vc11 with Xba I and Sal I, vc12 and wps57 with Xba I and Sal I, vc16 and vc13 with Xba I and Kpn I, vc14 and vc15 with Xba I and Kpn I. Then the wps54-vc11 and vc12-wps57 fragments were excised with Xba I and Sal I ligated together into the Xba I site of pVg1-dhfr; and the vc16-vc13 fragments and vc14-vc15 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVk.

FIG. 28. Competitive binding of labeled mik- β 1 tracer to 60 YTJB cells. About 10^6 YTJB cells were incubated with 3.0 ng of radio-iodinated mouse mik- β 1 antibody (6 μ Ci/ μ g) and varing amounts of either unlabeled mouse mik- β 1 antibody (\bullet) or humanized mik- β 1 antibody (\circ) in 200 ul of binding buffer (PBS+10% fetal calf serum+0.1% NaN₃+10 μ g/ml 65 mouse monoclonal Ig). After incubation for 2 hr at 0° C. the cells were washed twice with binding buffer without mouse

8

Ig and collected by centrifugation. The radioactivity bound to cells was measured and expressed as the ratio of bound/ free cpm.

FIG. 29. Inhibition of IL-2 stimulated proliferation of human PHA blasts by humanized mik- β 1+humanized anti-Tac antibodies. No antibody added (\square), 2 ug each of humanized mik- β 1 and humanized anti-Tac added (\blacksquare).

FIG. 30A through FIG. 30D. Amino acid sequences of the heavy chain (A) [SEQ ID NOS:48 and 49] and the light chain (B) [SEQ ID NOS:46 and 47] of the murine and humanized Fd79 antibodies, and the heavy chain (C) [SEQ ID NOS:52 and 53] and the light chain (D) [SEQ ID NOS:50 and 51] of the murine and humanized Fd138-80 antibodies. The sequences of the murine antibody as deduced from the cDNA (upper lines) are shown aligned with the humanized antibody sequences (lower lines). The humanized Fd79 and Fd138-80 framework sequences are derived from Pom antibody and Eu antibody, respectively. Residues are numbered according to the Kabat system (E. A. Kabat et al., Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md.) (1987). The three CDRs in each chain are boxed. Residues in the Pom or Eu framework that have been replaced with murine sequences or consensus human sequences are underlined.

FIG. 31A and FIG. 31B. Fluorocytometry of HSV-1 infected Vero cells stained with Fd79 (A) and Fd138-80 (B) antibodies. (. .) Isotype matched control antibody, (...) humanized antibody, (_) chimeric antibody. Vero cells were infected with HSV-1 (Δ305 mutant (F strain)) at 3 pfu/cell overnight. Cells were trypsinized at 0.5 mg/ml for 1 minute, washed extensively with PBS and resuspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10⁶/ ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human antibody (Cappel) on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehyde. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 32A and FIG. 32B. Neutralization of HSV-1 by Fd79 (A) and Fd138-80 (B). Serial dilutions of antibodies were mixed with 100 pfu of virus and incubated at 37° C. for 1 hr. The viruses were then inoculated onto 6-well plates with confluent Vero cells and adsorbed at 37° C. for 1 hr. Cells were overlayed with 1% agarose in medium and incubated for 4 days. Plaques were stained with neutral red.

FIG. 33A and FIG. 33B. Immunostaining of infected Vero cell monolayers to examine protection of cells from viral spread in tissue culture by (A) murine or humanized Fd79, (B) murine or humanized Fd138-80. 24-well plates of confluent Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before adding 200 ul of 10 ug/ml antibodies in medium. At the end of 4 days, culture medium was removed and plates were dried by placing overnight in a 37° C. incubator. To detect viral antigens, each well was incubated with 200 ul of anti-gB antibody at 0.5 ug/ml for 1 hr. at 37° C., washed twice and incubated with 200 ul of peroxidase conjugated goat antimouse IgG (Cappel, 1:300 dilution) for 1 hr. at 37° C. The plates were washed and then developed with the substrate 3-amino-9-ethyl-carbazole (AEC) (Sigma, St. Louis, Mo.) for 15 minutes at room temperature. Reaction was stopped by rinsing with water and air dried.

FIG. 34A and FIG. 34B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ

ID NOS:66 and 67] and heavy chain (B) [SEQ ID NOS:68 and 69] variable regions of the antibody M195. The CDR sequences are underlined. The mature light chain protein begins with amino acid 21 D and the mature heavy chain protein with amino acid 20 E, preceded by the respective signal sequences.

FIG. 35. Fluorocytometry of U937 cells stained with (...) no antibody, (...) humanized M195 antibody, (---) chimeric M195 antibody. Cells were suspended in FACS buffer (PBS+2% FCS+0.1% azide) at approximately 5×10⁶/ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 50 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human Ig antibody on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehyde. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 36A and FIG. 36B. Amino acid sequences of the light chain (A) [SEQ ID NOS:70 and 71] and the heavy chain (B) [SEQ ID NOS:72 and 73] of the humanized M195 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids in the humanized antibody are double underlined.

FIG. 37A and FIG. 37B. Oligonucleotides used in the construction of the humanized M195 heavy chain (A; ma1–4) [SEQ ID NOS:74, 75, 76, and 77] and light chain (B; ma5–8) [SEQ ID NOS:78, 79, 80, and 81]. The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: ma1 and ma2 with Xba I and Kpn I, ma3 and ma4 with Xba I and Kpn I, ma5 and ma6 with Xba I and Hind III, ma7 and ma8 with Xba I and Hind III. Then the ma1–ma2 and ma3–ma4 fragments were excised from pUC18 with Xba I and kpn I and ligated together into the Xba I site of pVg1-dhfr; and the ma5–ma6 and ma7–ma8 fragments were excised with Xba I and Hind III and ligated together into the Xba I site of pVk.

FIG. 38. Competitive binding of labeled M195 tracer to U937 cells. About 4×10⁵ U937 cells were incubated with 4.5 ng of radio-iodinated mouse M195 antibody (6 μci/μg) and varying amounts of either unlabeled mouse M195 antibody (Φ) or humanized M195 antibody (Φ) in 200 ul of binding buffer (PBS+2% fetal calf serum+0.1% sodium azide). After incubation for 2 hr at 0° C., the cells were washed twice with binding buffer and collected by centrifugation. The radioactivity bound to cells was measured and is expressed as the ratio of bound/free cpm.

FIG. 39A and FIG. 39B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:82 and 83] and heavy chain (B) [SEQ ID NOS:84 and 85] variable regions of the antibody CMV5. The CDR sequences are underlined. The start of the mature protein sequences are indicated by arrows, preceded by the respective signal sequences.

FIG. 40A and FIG. 40B. Amino acid sequences of the light chain (A) [SEQ ID NOS:86 and 87] and the heavy chain (B) [SEQ ID NOS:88 and 89] of the humanized CMV5 antibody (lower lines) and human Wol antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 41A and FIG. 41B. Oligonucleotides used in the construction of the humanized CMV5 light chain (A;

10

jb16-jb19) [SEQ ID NOS:90, 91, 92, and 93] and heavy chain (B; jb20-jb22) [SEQ ID NOS:94, 95, 96, and 97]. The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: jb16 and jb17 with Xba I and EcoR I, jb18 and jb19 with Xba I and EcoR I, jb20 and jb21 with Xba I and Kpn I, jb22 and jb23 with Xba I and Kpn I. Then the jb16-jb17 and jb18-jb19 fragments were excised with Xba I and Mlu I and ligated together into the Xba I site of pVk; and the jb20-jb21 and jb22-jb23 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVg1-dhfr.

FIG. 42. Competitive binding of labeled CMV5 tracer to CMV-infected cells. Increasing amounts of mouse (●) or humanized (○) CMV5 antibody was added to CMV-infected HEL cells with tracer radio-iodinated mouse CMV5, and the amount of tracer bound to the cells was determined.

FIG. 43A and FIG. 43B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:98 and 99] and heavy chain (B) [SEQ ID NOS:100 and 101] variable regions of the antibody AF2. The CDR sequences are underlined. The mature light chain protein begins with amino acid 30 N and the mature heavy chain protein with amino acid 36 Q, preceded by the respective signal sequences.

FIG. 44A and FIG. 44B. Amino acid sequences of the light chain (A) [SEQ ID NOS:102 and 103] and the heavy chain (B) [SEQ ID NOS:104 and 105] of the humanized AF2 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 45A and FIG. 45B. Oligonucleotides used in the construction of the humanized AF2 light chain (A; rh10-rh13) [SEQ ID NOS:106, 107, 108, and 109] and heavy chain (B; rh20-23) [SEQ ID NOS:110, 111, 112, and 113]. The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: rh10 and rh11 with Xba I and Hind III, rh12 and rh13 with Xba I and Hind III, rh20 and rh21 with Xba I and EcoR I, rh22 and rh23 with Xba I and EcoR I. Then the rh10-rh11 and rh12-rh13 fragments were excised with Xba I and Hind III and ligated together into then Xba I site of pVk; and the rh20-rh21 and rh22-rh23 fragments were excised with Xba I and Xho I and ligated together into the Xba I site of pVg1-dhfr.

FIG. 46. Fluorescence of HS294T cells incubated with γ -IFN plus varying concentrations of mouse AF2 antibody, and stained with an anti-HLA-D antibody.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, novel means of designing humanized immunoglobulins capable of specifically binding to a predetermined antigen with strong affinity are provided. These improved methods produce immunoglobulins that are substantially non-immunogenic in humans but have binding affinities of at least about 10⁸ M⁻¹, preferably 10⁹ M⁻¹ to 10¹⁰ M⁻¹, or stronger. The humanized immunoglobulins will have a human framework and have one or more complementary determining regions (CDR's), plus a limited number of other amino acids, from a donor immunoglobulin specifically reactive with an antigen. The

immunoglobulins can be produced economically in large quantities and find use, for example, in the treatment of various human disorders by a variety of techniques.

In order that the invention may be more completely understood, several definitions are set forth. As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma (IgG1, IgG2, IgG3, IgG₄), delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Fulllength immunoglobulin "light chains" (about 25 Kd or 214 amino acids) are encoded by a variable region gene at the NH2-terminus (about 110 amino acids) and a kappa or lambda constant region gene at the COOH—terminus. Fulllength immunoglobulin "heavy chains" (about 50 Kd or 446 amino acids), are similarly encoded by a variable region gene (about 116 amino acids) and one of the other aforementioned constant region genes, e.g., gamma (encoding about 330 amino acids).

One form of immunoglobulin constitutes the basic structural unit of an antibody. This form is a tetramer and consists of two identical pairs of immunoglobulin chains, each pair having one light and one heavy chain. In each pair, the light and heavy chain variable regions are together responsible for binding to an antigen, and the constant regions are responsible for the antibody effector functions. In addition to antibodies, immunoglobulins may exist in a variety of other forms including, for example, Fv, Fab, and $(Fab')_2$, as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston et al., Proc. Natl. Acad. Sci. U.S.A., 85, 5879-5883 (1988) and Bird et al., Science, 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood et al., "Immunology", Benjamin, N.Y., 2nd ed. (1984), and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference).

An immunoglobulin light or heavy chain variable region consists of a "framework" region interrupted by three hypervariable regions, also called CDR's. The extent of the $_{40}$ framework region and CDR's have been precisely defined (see, "Sequences of Proteins of Immunological Interest," E. Kabat et al., U.S. Department of Health and Human Services, (1983); which is incorporated herein by reference). The sequences of the framework regions of different light or 45 heavy chains are relatively conserved within a species. As used herein, a "human framework region" is a framework region that is substantially identical (about 85% or more, usually 90-95% or more) to the framework region of a naturally occurring human immunoglobulin. The framework 50 region of an antibody, that is the combined framework regions of the constituent light and heavy chains, serves to position and align the CDR's. The CDR's are primarily responsible for binding to an epitope of an antigen.

Chimeric antibodies are antibodies whose light and heavy 55 chain genes have been constructed, typically by genetic engineering, from immunoglobulin variable and constant region genes belonging to different species. For example, the variable segments of the genes from a mouse monoclonal antibody may be joined to human constant segments, such as 60 gamma 1 and gamma 3. A typical therapeutic chimeric antibody is thus a hybrid protein composed of the variable or antigen-binding domain from a mouse antibody and the constant or effector domain from a human antibody (e.g., A.T.C.C. Accession No. CRL 9688 secretes an anti-Tac 65 chimeric antibody), although other mammalian species may be used.

12

As used herein, the term "humanized" immunoglobulin refers to an immunoglobulin comprising a human framework region and one or more CDR's from a non-human (usually a mouse or rat) immunoglobulin. The non-human immunoglobulin providing the CDR's is called the "donor" and the human immunoglobulin providing the framework is called the "acceptor". Constant regions need not be present, but if they are, they must be substantially identical to human immunoglobulin constant regions, i.e., at least about 85-90%, preferably about 95% or more identical. Hence, all parts of a humanized immunoglobulin, except possibly the CDR's, are substantially identical to corresponding parts of natural human immunoglobulin sequences. A "humanized antibody" is an antibody comprising a humanized light chain and a humanized heavy chain immunoglobulin. For example, a humanized antibody would not encompass a typical chimeric antibody as defined above, e.g., because the entire variable region of a chimeric antibody is non-human. One says that the donor antibody has been "humanized", by the process of "humanization", because the resultant humanized antibody is expected to bind to the same antigen as the donor antibody that provides the CDR's.

It is understood that the humanized antibodies designed by the present method may have additional conservative amino acid substitutions which have substantially no effect on antigen binding or other immunoglobulin functions. By conservative substitutions is intended combinations such as gly, ala; val, ile, leu; asp, glu; asn, gln; ser, thr; lys, arg; and phe, tyr.

Humanized immunoglobulins, including humanized antibodies, have been constructed by means of genetic engineering. Most humanized immunoglobulins that have been previously described (Jones et al., op. cit.; Verhoeyen et al., op. cit.; Riechmann et al., op. cit.) have comprised a framework that is identical to the framework of a particular human immunoglobulin chain, the acceptor, and three CDR's from a non-human donor immunoglobulin chain. In one case (Riechmann et al., op. cit.), two additional amino acids in the framework were changed to be the same as amino acids in other human framework regions. The present invention includes criteria by which a limited number of amino acids in the framework of a humanized immunoglobulin chain are chosen to be the same as the amino acids at those positions in the donor rather than in the acceptor, in order to increase the affinity of an antibody comprising the humanized immunoglobulin chain.

The present invention is based in part on the model that two contributing causes of the loss of affinity in prior means of producing humanized antibodies (using as examples mouse antibodies as the source of CDR's) are:

- (1) When the mouse CDR's are combined with the human framework, the amino acids in the framework close to the CDR's become human instead of mouse. Without intending to be bound by theory, we believe that these changed amino acids may slightly distort the CDR's, because they create different electrostatic or hydrophobic forces than in the donor mouse antibody, and the distorted CDR's may not make as effective contacts with the antigen as the CDR's did in the donor antibody;
- (2) Also, amino acids in the original mouse antibody that are close to, but not part of, the CDR's (i.e., still part of the framework), may make contacts with the antigen that contribute to affinity. These amino acids are lost when the antibody is humanized, because all framework amino acids are made human.

To avoid these problems, and to produce humanized antibodies that have a very strong affinity for a desired antigen, the present invention uses one or more of the following principles for designing humanized immunoglobulins. Also, the criteria may be used singly, or when 5 necessary in combination, to achieve the desired affinity or other characteristics.

A principle is that as acceptor, a framework is used from a particular human immunoglobulin that is unusually homologous to the donor immunoglobulin to be humanized, 10 or use a consensus framework from many human antibodies. For example, comparison of the sequence of a mouse heavy (or light) chain variable region against human heavy (or light) variable regions in a data bank (for example, the National Biomedical Research Foundation Protein Identifi- 15 cation Resource) shows that the extent of homology to different human regions varies greatly, typically from about 40% to about 60-70%. By choosing as the acceptor immunoglobulin one of the human heavy (respectively light) chain variable regions that is most homologous to the heavy 20 (respectively light) chain variable region of the donor immunoglobulin, fewer amino acids will be changed in going from the donor immunoglobulin to the humanized immunoglobulin. Hence, and again without intending to be bound by theory, it is believed that there is a smaller chance of 25 changing an amino acid near the CDR's that distorts their conformation. Moreover, the precise overall shape of a humanized antibody comprising the humanized immunoglobulin chain may more closely resemble the shape of the donor antibody, also reducing the chance of distorting the 30 CDR's.

Typically, one of the 3–5 most homologous heavy chain variable region sequences in a representative collection of at least about 10 to 20 distinct human heavy chains will be chosen as acceptor to provide the heavy chain framework, 35 and similarly for the light chain. Preferably, one of the 1–3 most homologous variable regions will be used. The selected acceptor immunoglobulin chain will most preferably have at least about 65% homology in the framework region to the donor immunoglobulin.

In many cases, it may be considered preferable to use light and heavy chains from the same human antibody as acceptor sequences, to be sure the humanized light and heavy chains will make favorable contacts with each other. In this case, the donor light and heavy chains will be compared only 45 against chains from human antibodies whose complete sequence is known, e.g., the Eu, Lay, Pom, Wol, Sie, Gal, Ou and WEA antibodies (Kabat et al., op. cit.; occasionally, the last few amino acids of a human chain are not known and must be deduced by homology to other human antibodies). 50 The human antibody will be chosen in which the light and heavy chain variable regions sequences, taken together, are overall most homologous to the donor light and heavy chain variable region sequences. Sometimes greater weight will be given to the heavy chain sequence. The chosen human 55 antibody will then provide both light and heavy chain acceptor sequences. In practice, it is often found that the human Eu antibody will serve this role.

Regardless of how the acceptor immunoglobulin is chosen, higher affinity may be achieved by selecting a small 60 number of amino acids in the framework of the humanized immunoglobulin chain to be the same as the amino acids at those positions in the donor rather than in the acceptor. A second principle is that the following categories define what amino acids may be selected from the donor. Preferably, at 65 many or all amino acid positions in one of these categories, the donor amino acid will in fact be selected.

14

Category 1: The amino acid position is in a CDR is defined by Kabat et al., op. cit.

Category 2: If an amino acid in the framework of the human acceptor immunoglobulin is unusual (i.e., "rare", which as used herein indicates an amino acid occurring at that position in less than about 20% but usually less than about 10% of human heavy (respectively light) chain V region sequences in a representative data bank), and if the donor amino acid at that position is typical for human sequences (i.e., "common", which as used herein indicates an amino acid occurring in more than about 25% but usually more than about 50% of sequences in a representative data bank), then the donor amino acid rather than the acceptor may be selected. This criterion helps ensure that an atypical amino acid in the human framework does not disrupt the antibody structure. Moreover, by replacing an unusual amino acid with an amino acid from the donor antibody that happens to be typical for human antibodies, the humanized antibody may be made less immunogenic.

All human light and heavy chain variable region sequences are respectively grouped into "subgroups" of sequences that are especially homologous to each other and have the same amino acids at certain critical positions (Kabat et al., op. cit.). When deciding whether an amino acid in a human acceptor sequence is "rare" or "common" among human sequences, it will often be preferable to consider only those human sequences in the same subgroup as the acceptor sequence.

Category 3: In the positions immediately adjacent to one or more of the 3 CDR's in the primary sequence of the humanized immunoglobulin chain, the donor amino acid(s) rather than acceptor amino acid may be selected. These amino acids are particularly likely to interact with the amino acids in the CDR's and, if chosen from the acceptor, to distort the donor CDR's and reduce affinity. Moreover, the adjacent amino acids may interact directly with the antigen (Amit et al., *Science*, 233, 747–753 (1986), which is incorporated herein by reference) and selecting these amino acids from the donor may be desirable to keep all the antigen contacts that provide affinity in the original antibody.

Category 4: A 3-dimensional model, typically of the original donor antibody, shows that certain amino acids outside of the CDR's are close to the CDR's and have a good probability of interacting with amino acids in the CDR's by hydrogen bonding, Van der Waals forces, hydrophobic interactions, etc. At those amino acid positions, the donor immunoglobulin amino acid rather than the acceptor immunoglobulin amino acid may be selected. Amino acids according to this criterion will generally have a side chain atom within about 3 angstrom units of some atom in the CDR's and must contain an atom that could interact with the CDR atoms according to established chemical forces, such as those listed above.

In the case of atoms that may form a hydrogen bond, the 3 angstroms is measured between their nuclei, but for atoms that do not form a bond, the 3 angstroms is measured between their Van der Waals surfaces. Hence, in the latter case, the nuclei must be within about 6 angstroms (3+sum of the Van der Waals radii) for the atoms to be considered capable of interacting. In many cases the nuclei will be from 4 or 5 to 6 Å apart. In determining whether an amino acid can interact with the CDRs, it is preferred not to consider the last 8 amino acids of heavy chain CDR 2 as part of the CDRs, because from the viewpoint of structure, these 8 amino acids behave more as part of the framework.

Amino acids in the framework that are capable of interacting with amino acids in the CDR's, and which therefore belong to Category 4, may be distinguished in another way. The solvent accessible surface area of each framework amino acid is calculated in two ways: (1) in the intact 5 antibody, and (2) in a hypothetical molecule consisting of the antibody with its CDRs removed. A significant difference between these numbers of about 10 square angstroms or more shows that access of the framework amino acid to solvent is at least partly blocked by the CDRs, and therefore that the amino acid is making contact with the CDRs. Solvent accessible surface area of an amino acid may be calculated based on a 3-dimensional model of an antibody, using algorithms known in the art (e.g., Connolly, J. Appl. Cryst. 16, 548 (1983) and Lee and Richards, J. Mol. Biol. 55, 379 (1971), both of which are incorporated herein by reference). Framework amino acids may also occasionally interact with the CDR's indirectly, by affecting the conformation of another framework amino acid that in turn con-20 tacts the CDR's.

The amino acids at several positions in the framework are known to be capable of interacting with the CDRs in many antibodies (Chothia and Lesk, J. Mol. Biol. 196, 901 (1987), Chothia et al., Nature 342, 877 (1989), and Tramontano et 25 al., J. Mol. Biol. 215, 175 (1990), all of which are incorporated herein by reference), notably at positions 2, 48, 64 and 71 of the light chain and 26-30, 71 and 94 of the heavy chain (numbering according to Kabat, op. cit.), and therefore these amino acids will generally be in Category 4. Typically, 30 humanized immunoglobulins, of the present invention will include donor amino acids (where different) in category 4 in addition to these. The amino acids at positions 35 in the light chain and 93 and 103 in the heavy chain are also likely to interact with the CDRs. At all these numbered positions, 35 choice of the donor amino acid rather than the acceptor amino acid (when they differ) to be in the humanized immunoglobulin is preferred. On the other hand, certain positions that may be in Category 4 such as the first 5 amino acids of the light chain may sometimes be chosen from the 40 acceptor immunoglobulin without loss of affinity in the humanized immunoglobulin.

Chothia and Lesk (op. cit.) define the CDRs differently from Kabat et al. (op. cit.). Notably, CDR1 is defined as including residues 26–32. Accordingly, Riechmann et al., 45 (op. cit.) chose these amino acids from the donor immunoglobulins.

Computer programs to create models of proteins such as antibodies are generally available and well known to those skilled in the art (see, Levy et al., *Biochemistry*, 28, 50 7168–7175 (1989); Bruccoleri et al., *Nature*, 335, 564–568 (1988); Chothia et al., *Science*, 233, 755–758 (1986), all of which are incorporated herein by reference). These do not form part of the invention. Indeed, because all antibodies have similar structures, the known antibody structures, 55 which are available from the Brookhaven Protein Data Bank, can be used if necessary as rough models of other antibodies. Commercially available computer programs can be used to display these models on a computer monitor, to calculate the distance between atoms, and to estimate the 60 likelihood of different amino acids interacting (see, Ferrin et al., *J. Mol. Graphics*, 6, 13–27 (1988)).

In addition to the above categories, which describe when an amino acid in the humanized immunoglobulin may be taken from the donor, certain amino acids in the humanized 65 immunoglobulin may be taken from neither the donor nor acceptor, if then fall in: 16

Category 5: If the amino acid at a given position in the donor immunoglobulin is "rare" for human sequences, and the amino acid at that position in the acceptor immunoglobulin is also "rare" for human sequences, as defined above, then the amino acid at that position in the humanized immunoglobulin may be chosen to be some amino acid "typical" of human sequences. A preferred choice is the amino acid that occurs most often at that position in the known human sequences belonging to the same subgroup as the acceptor sequence.

Humanized antibodies generally have at least three potential advantages over mouse or in some cases chimeric antibodies for use in human therapy:

- Because the effector portion is human, it may interact better with the other parts of the human immune system (e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC)).
- 2) The human immune system should not recognize the framework or constant region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.
- 3) Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of normal antibodies (D. Shaw et al., *J. Immunol.*, 138, 4534–4538 (1987)). Injected humanized antibodies will presumably have a half-life more similar to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

In one aspect, the present invention is directed to designing humanized immunoglobulins that are produced by expressing recombinant DNA segments encoding the heavy and light chain CDR's from a donor immunoglobulin capable of binding to a desired antigen, such as the human IL-2 receptor, attached to DNA segments encoding acceptor human framework regions. Exemplary DNA sequences designed in accordance with the present invention code for the polypeptide chains comprising heavy and light chain CDR's with substantially human framework regions shown in FIG. 1A through FIG. 6B. Due to codon degeneracy and non-critical amino acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. In general, the criteria of the present invention find applicability to designing substantially any humanized immunoglobulin.

The DNA segments will typically further include an expression control DNA sequence operably linked to the humanized immunoglobulin coding sequences, including naturally-associated or heterologous promoter regions. Preferably, the expression control sequences will be eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells, but control sequences for prokaryotic hosts may also be used. Once the vector has been incorporated into the appropriate host, the host is maintained under conditions suitable for high level expression of the nucleotide sequences, and, as desired, the collection and purification of the humanized light chains, heavy chains, light/heavy chain dimers or intact antibodies, binding fragments or other immunoglobulin forms may follow (see, S. Beychok, Cells of Immunoglobulin Synthesis, Academic Press, N.Y., (1979), which is incorporated herein by reference).

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat

op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to the predetermined antigen, such as the human IL-2 receptor, and produced by well known methods in any convenient 5 mammalian source including, mice, rats, rabbits, or other vertebrates, capable of producing antibodies. Suitable source cells for the constant region and framework DNA sequences, and host cells for immunoglobulin expression and secretion, can be obtained from a number of sources, such as the American Type Culture Collection ("Catalogue of Cell Lines and Hybridomas," sixth edition (1988) Rockville, Md., U.S.A., which is incorporated herein by reference).

In addition to the humanized immunoglobulins specifi- 15 cally described herein, other "substantially homologous" modified immunoglobulins to the native sequences can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the framework regions can vary specifi- 20 cally from the sequences in FIG. 1A through FIG. 6B at the primary structure level by several amino acid substitutions, terminal and intermediate additions and deletions, and the like. Moreover, a variety of different human framework regions may be used singly or in combination as a basis for 25 the humanized immunoglobulins of the present invention. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, Gene, 8, 81–97 (1979) and S. Roberts et al., *Nature*, 328, 731–734 30 (1987), both of which are incorporated herein by reference).

Substantially homologous immunoglobulin sequences are those which exhibit at least about 85% homology, usually at least about 90%, and preferably at least about 95% homology with a reference immunoglobulin protein.

Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic 40 cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVg1 (FIGS. 9A and 9B) using sitedirected mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce (Fab')₂ 45 fragments. Single chain antibodies may be produced by joining VL and VH with a DNA linker (see, Huston et al., op. cit., and Bird et al., op. cit.). Also because like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological 50 activities, the genes may be fused to functional regions from other genes (e.g., enzymes, see, commonly assigned U.S. Pat. No. 5,004,692) to produce fusion proteins (e.g., immunotoxins) having novel properties. The nucleic acid sequences of the present invention capable of ultimately 55 expressing the desired humanized antibodies can be formed from a variety of different polynucleotides (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (e.g., V, J, D, and C regions), as well as by a variety of different techniques. Joining appropriate synthetic and 60 genomic sequences is presently the most common method of production, but cDNA sequences may also be utilized (see, European Patent Publication No. 0239400 and L. Reichmann et al., Nature, 332, 323-327 (1988), both of which are incorporated herein by reference).

As stated previously, the DNA sequences will be expressed in hosts after the sequences have been operably

linked to (i.e., positioned to ensure the functioning of) an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection markers, e.g., tetracycline or neomycin, to permit detection of those cells transformed with the desired DNA sequences (see, e.g., U.S. Pat. No. 4,704,362, which is incorporated herein by reference).

E. coli is one prokaryotic host useful particularly for cloning the DNA sequences of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella., Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of wellknown promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a betalactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. Saccharomyces is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the polypeptides of the present invention (see, Winnacker, "From Genes to Clones," VCH Publishers, N.Y., N.Y. (1987), which is incorporated herein by reference). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various COS cell lines, HeLa cells, preferably myeloma cell lines, etc, and transformed B-cells or hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, an enhancer (Queen et al., Immunol. Rev., 89, 49-68 (1986), which is incorporated herein by reference), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, Adenovirus, cytomegalovirus, Bovine Papilloma Virus, and the like.

The vectors containing the DNA segments of interest (e.g., the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (See, generally, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, (1982), which is incorporated herein by reference.)

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention, can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel

electrophoresis and the like (see, generally, R. Scopes, "Protein Purification", Springer-Verlag, N.Y. (1982)). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses. Once purified, partially or to homogeneity as desired, the polypeptides may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings, and the like. (See, generally, *Immunological Methods*, Vols. I and II, Lefkovits and Pernis, eds., Academic Press, New York, N.Y. (1979 and 1981)).

The antibodies of the present invention will typically find use individually in treating substantially any disease susceptible to monoclonal antibody-based therapy. In particular, the immunoglobulins can be used for passive immunization or the removal of unwanted cells or antigens, such as by complement mediated lysis, all without substantial immune reactions (e.g., anaphylactic shock) associated with many prior antibodies. For example, where the cell linked to a disease has been identified as IL-2 receptor bearing, then 20 humanized antibodies that bind to the human IL-2 receptor are suitable (see, U.S. Ser. No. 085,707, entitled "Treating Human Malignancies and Disorders," which is incorporated herein by reference). For such a humanized immunoglobulin, typical disease states suitable for treatment include graft 25 versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia 30 gravis.

The method of producing humanized antibodies of the present invention can be used to humanize a variety of donor antibodies, especially monoclonal antibodies reactive with markers on cells responsible for a disease. For example, 35 suitable antibodies bind to antigens on T-cells, such as those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The antibodies of the present invention can also be used as separately administered compositions given in conjunction with chemotherapeutic or immunosuppressive agents. Possible agents include cyclosporin A or a purine analog 45 (e.g., methotrexate, 6-mercaptopurine, or the like), but numerous additional agents (e.g., cyclophosphamide, prednisone, etc.) well-known to those skilled in the art of medicine may also be utilized.

A preferred pharmaceutical composition of the present 50 invention comprises the use of the subject antibodies in immunotoxins. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. 55 The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells comprising a carcinoma. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For 60 example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can 65 be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Mono-

clonal Antibodies in Clinical Medicine, Academic Press, pp. 168–190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see, Chaudhary et al., *Nature* 339, 394 (1989), which is herein incorporated by reference).

A variety of cytotoxic agents are suitable for use in immunotoxins. Cytotoxic agents can include radionuclides, such as Iodine-131 or other isotopes of iodine, Yttrium-90, Rhenium-188, and Bismuth-212 or other alpha emitters; a number of chemotherapeutic drugs, such as vindesine, methotrexate, adriamycin, and cisplatinum; and cytotoxic proteins such as ribosomal inhibiting proteins like pokeweed antiviral protein, Pseudomonas exotoxin A, ricin, diphtheria toxin, ricin A chain, etc., or an agent active at the cell surface, such as the phospholipase enzymes (e.g., phospholipase C). (See, generally, "Chimeric Toxins," Olsnes and Phil, *Pharmac. Ther.*, 25, 355–381 (1982), and "Monoclonal Antibodies for Cancer Detection and Therapy," eds. Baldwin and Byers, pp. 159–179, 224–266, Academic Press (1985), all of which are incorporated herein by reference.)

The delivery component of the immunotoxin will include the humanized immunoglobulins of the present invention. Intact immunoglobulins or their binding fragments, such as Fab, are preferably used. Typically, the antibodies in the immunotoxins will be of the human IgM or IgG isotype, but other mammalian constant regions may be utilized as desired.

For diagnostic purposes, the antibodies may either be labeled or unlabeled. Unlabeled antibodies can be used in combination with other labeled antibodies (second antibodies) that are reactive with the humanized antibody, such as antibodies specific for human immunoglobulin constant regions. Alternatively, the antibodies can be directly labeled. A wide variety of labels may be employed, such as radionuclides, fluors, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, ligands (particularly haptens), etc. Numerous types of immunoassays are available and are well known to those skilled in the art.

Anti-IL-2 Receptor Antibodies

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), both of which are incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide, being about 55 kD in size (See, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). A gene encoding this protein has been isolated, and predicts a 272 amino acid peptide, including a 21 amino acid signal peptide (see, Leonard, W., et al., Nature 311, 626 (1984)). The 219 NH₂-terminal amino acids of the p55 Tac protein apparently comprise an extracellular domain (see, Leonard, W., et al., Science, 230, 633-639 (1985), which is incorporated herein by reference).

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. In particular, one mouse monoclonal antibody, known as anti-Tac (Uchiyama, et al., *J. Immunol.* 126, 1393 (1981)) has been used to show that IL-2 receptors can be detected on T-cells, but also on cells of the monocyte-macrophage family, Kupffer cells of the liver, Langerhans' cells of the skin and, of course, activated T-cells. Importantly, resting T-cells, B-cells or

circulating machrophages typically do not display the IL-2 receptor (Herrmann, et al., *J. Exp. Med.* 162, 1111 (1985)).

The anti-Tac monoclonal antibody has also been used to define lymphocyte functions that require IL-2 interaction, and has been shown to inhibit various T-cell functions, including the generation of cytotoxic and suppressor T lymphocytes in cell culture. Also, based on studies with anti-Tac and other antibodies, a variety of disorders are now associated with improper IL-2 receptor expression by T-cells, in particular adult T-cell leukemia.

More recently, the IL-2 receptor has been shown to be an ideal target for novel therapeutic approaches to T-cell mediated diseases. It has been proposed that IL-2 receptor specific antibodies, such as the anti-Tac monoclonal antibody, can be used either alone or as an immunoconjugate (e.g., with Ricin A, isotopes and the like) to effectively remove cells bearing the IL-2 receptor. These agents can, for example, theoretically eliminate IL-2 receptor-expressing leukemic cells, certain B-cells, or activated T-cells involved in a disease state, yet allow the retention of mature normal T-cells and their precursors to ensure the capability of mounting a normal T-cell immune response as needed. In general, most other T-cell specific agents can destroy essentially all peripheral T-cells, which limits the agents' therapeutic efficacy. Overall, the use of appropriate monoclonal antibodies specific for the IL-2 receptor may have therapeutic utility in autoimmune diseases, organ transplantation and any unwanted response by activated T-cells. Indeed, clinical trials have been initiated using, e.g., anti-Tac antibodies (see, generally, Waldmann, T., et al., Cancer Res. 45, 625 (1985), Waldmann, T., Science 232, 727-732 (1986) and Kirkman et al., Transplant. Proc. 21, 1766 (1989), all of which are incorporated herein by reference).

Unfortunately, the use of the anti-Tac and other non-human monoclonal antibodies have certain drawbacks, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans

Perhaps more importantly, anti-Tac and other non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that, after injection of a foreign antibody, the immune response elicited by a patient against an antibody can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after the first or several treatments with any different non-human antibodies, subsequent treatments even for unrelated therapies can be ineffective or even dangerous in themselves, because of cross-

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with the human IL-2 receptor, as with many human antigens, has been extremely difficult using typical human monoclonal antibody production techniques. Similarly, utilizing recombinant DNA technology to produce so-called "reshaped" or 65 "humanized" antibodies (see, e.g., Riechmann et al., *Nature* 332, 323 (1988) and EPO Publication No. 0239400), pro-

22

vides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of human-like immunoglobulins specific for the human IL-2 receptor that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of blocking the binding of human IL-2 to its receptor and/or capable of binding to the p55 Tac protein on human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the human IL-2 receptor at affinity levels stronger than about 10⁸ M⁻¹.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for human-like immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the IL-2 receptor on human T-cells are provided. These immunoglobulins, which have binding affinities of at least about 10⁸ M⁻¹, and preferably 10⁹ M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p55 Tac protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the anti-Tac monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Preferred DNA sequences, which on expression code for the

polypeptide chains comprising the anti-Tac heavy and light chain hypervariable regions (with human-like framework regions), are included in FIGS. 15A and and 16A, respectively. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like 10 antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, *Leukocyte Typing*, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The human-like antibodies and pharmaceutical compositions thereof of this invention are particularly useful for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously. The compositions for parenteral administration will commonly comprise a solution of the antibody or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate, human albumin, etc. The concentration of antibody in these formulations can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on fluid volumes, viscosities, etc., in accordance with the particular mode of administration selected.

Thus, a typical pharmaceutical composition for injection could be made up to contain 1 ml sterile buffered water, and 1 to 50 mg of antibody. A typical composition for intravenous infusion could be made up to contain 250 ml of sterile Ringer's solution, and 150 mg of antibody. Actual methods for preparing parenterally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, *Remington's Pharmaceutical Science*, 15th ed., Mack Publishing Company, Easton, Pa. (1980), which is incorporated herein by reference.

The antibodies of this invention can be frozen or lyophilized for storage and reconstituted in a suitable carrier 65 prior to use. This technique has been shown to be effective with conventional immune globulins and art-known lyophilization and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilization and reconstitution can lead to varying degrees of antibody activity loss (e.g., with conventional immune globulins, IgM antibodies tend to have greater activity loss than IgG antibodies) and that use levels may have to be adjusted to compensate.

The compositions containing the present human-like antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from a disease, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially lifethreatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present human-like antibodies of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose, especially 0.5 to 2.5 mg per dose. A preferred prophylactic use is for the prevention of kidney transplant rejection.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Kits can also be supplied for use with the subject antibodies in the protection against or detection of a cellular activity or for the presence of a selected antigen. Thus, the subject antibody composition of the present invention may be provided, usually in a lyophilized form in a container, either alone or in conjunction with additional antibodies specific for the desired cell type. The antibodies, which may be conjugated to a label or toxin, or unconjugated, are included in the kits with buffers, such as Tris, phosphate, carbonate, etc., stabilizers, biocides, inert proteins, e.g., serum albumin, or the like, and a set of instructions for use. Generally, these materials will be present in less than about 5% wt. based on the amount of active antibody, and usually present in total amount of at least about 0.001% wt. based again on the antibody concentration. Frequently, it will be desirable to include an inert extender or excipient to dilute the active ingredients, where the excipient may be present in from about 1 to 99% wt. of the total composition. Where a

second antibody capable of binding to the chimeric antibody is employed in an assay, this will usually be present in a separate vial. The second antibody is typically conjugated to a label and formulated in an analogous manner with the antibody formulations described above.

p75 Chain of IL-2 Receptor

The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55 kD in size (see, Leonard, W., et al., *J. Biol. Chem.* 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., *Proc. Nat. Acad. Sci. U.S.A.*, 83, 9694 (1986) and Sharon et al., *Science* 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of inhibiting the binding of $\ ^{25}$ human IL-2 to its receptor and/or capable of binding to the p75 protein of human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the p75 protein at affinity levels stronger than about 10^7 M^{-1} . These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to p75.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the p75 chain of the human IL-2 receptor are provided. These immunoglobulins, which have binding affinities of at least 10^7 to 10^8 M $^{-1}$, and preferably 10^9 M $^{-1}$ to 10^{10} M $^{-1}$ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p75 protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain 65 CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the

26

mik- $\beta1$ monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the mik- $\beta1$ heavy and light chain CDRs, are included in FIG. 23A and FIG. 23B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft-versus-host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, *Leukocyte Typing*, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference. A preferred use is the simultaneous treatment of a patient with a human-like antibody binding to p55 and a human-like antibody binding to p75 of the IL-2 receptor, i.e., humanized anti-Tac plus humanized mik-β1.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Anti-HSV Antibodies

Herpes Simplex Virus types I and II (HSV-1 and HSV-2), are now estimated to be the second most frequent cause of sexually transmitted diseases in the world. Although completely accurate data are not available, infection estimates range from about 20 to 40% of the U.S. population.

A large number of diseases, from asymptomatic to lifethreatening, are associated with HSV infection. Of particular clinical interest, encephalitis from HSV-1 infection and transmission of HSV-2 from a pregnant mother to her fetus are often fatal. Immunosuppressed patients are also subject to severe complications when infected with the virus.

More than 50 HSV polypeptides have been identified in HSV-infected cells, including at least seven major cell surface glycoproteins (see, Whitley, R., Chapt. 66, and Roizman and Sears, Chapt. 65, Virology, Eds. Fields et al., 2nd ed., Raven Press, N.Y., N.Y. (1990), which are incorporated herein by reference). The specific biologic functions of these glycoproteins are not well defined, although gB and gD have been shown to be associated with cell fusion activity (W. Cai et al., J. Virol. 62, 2596 (1988) and Fuller and Spear, Proc. Natl. Acad. Sci. U.S.A., 5454 (1987)). gB and gD express both type-specific and type-common antigenic determinants. Oakes and Lausch demonstrated that

monoclonal antibodies against gB and gE suppress replication of HSV-1 in trigeminal ganglia (Oakes and Lausch, J. Virol. 51, 656 (1984)). Dix et al. showed that anti-gC and gD antibodies protect mice against acute virus-induced neurological disease (Dix et al., Infect. Immun. 34, 192 (1981)). Whitley and colleagues produced a panel of murine monoclonal antibodies against HSV-1 and showed that several of the antibodies protected mice against encephalitis and death following ocular inoculation with the virus (See, Koga et al., Virology 151, 385 (1986); Metcalf et al., Cur. Eye Res. 6, 10 173 (1987) and Metcalf et al., Intervirology 29, 39 1988), all of which are incorporated herein by reference). Clone Fd79 (anti-gB) prevented encephalitis even when immunization was delayed until 48 hours post-infection. Fd79 and Fd138-80 (anti-gD) significantly reduced the severity of epithelial 15 keratitis and lowered the frequency of persistent viral infection in an outbred mouse model.

Thus, there is a need for improved forms of humanized immunoglobulins specific for HSV antigens that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of HSV mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of HSV to its receptors and/or capable of binding to the HSV specific proteins. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the HSV surface proteins at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR donating mouse monoclonal antibody to HSV.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with an antiviral agent, such as acyclovir or a cytotoxic agent active at viral surfaces. All of these compounds will be particularly useful in treating HSV mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized 50 immunoglobulins specifically reactive with HSV related epitopes either directly on the virus or on infected cells are provided. These immunoglobulins, which have binding affinities to HSV specific antigens of at least about 107 Mand preferably $10^8 \,\mathrm{M}^{-1}$ to $10^{10} \,\mathrm{M}^{-1}$ or stronger, are capable $_{55}$ of, e.g., protecting cells from HSV transmission. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an HSV protein, such as gB and gD proteins. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of HSV mediated disorders in human patients by a variety of techniques.

The HSVs are among the most intensively investigated of all viruses, and the HSV virion structure has been shown to

contain about 33 proteins. Humanized immunoglobulins utilizing CDR's from monoclonal antibodies reactive with these proteins, particularly the eight surface glycoproteins (e.g., gB, gC, gD, gE, gG, gH and gI), represent preferred embodiments of the present invention (see, Spear, P. G., The Herpesviruses, vol. 3, pp. 315–356 (1984) (Roizman, B., ed), Plenum Press, N.Y., N.Y. and Spear, P. G., Immunochemistry of Viruses. The Basis for Serodiagnosis and Vaccines, pp. 425–446 (1985) (Neurath, A. R., eds.), Amsterdam: Elsevier, both of which are incorporated herein by reference).

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of an HSV protein, such as monoclonal antibodies reactive with HSV gB and gD glycoproteins. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate humanized framework regions. Exemplary DNA sequences code for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, shown in FIG. 30A through FIG. 30D. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating an HSV mediated disease state. For example, typical disease states suitable for treatment include any involving HSV infection. Specific diseases include neonatal herpes, herpes encephalitis, ocular herpes, genital herpes and disseminated herpes (see, Corey, L., Chapter 136, Harrison's Principles of Internal Medicine, 11th ed., McGraw-Hill Book Company, N.Y., N.Y. (1987), which is incorporated herein by reference).

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different HSV antigens. For example, suitable HSV antigens to which a cocktail of humanized immunoglobulins may react include gC, gE, gF, gG and gH (see, Rector, J. et al., Infect. Immun. 38, 168 (1982) and Fuller, A. et al., J. Virol. 63, 3435 (1989), both of which are incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with acyclovir or other antiviral agents. Typically, the agents may include idoxuridine or trifluorothymidine, but numerous additional agents (e.g., vidarabine) well-known to those skilled in the art for HSV treatment may also be utilized (see, Corey, L., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill cells infected by HSV. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing an HSV epitope.

The compositions containing the present humanized antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from HSV infection, in an amount sufficient to cure or at least partially arrest the disease and its complications.

An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody 5 per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially life-threatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present humanized immunoglobulins of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose. A preferred prophylactic use is for the prevention of herpes in immunocompromised patients, such as organ transplant recipients.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of HSV antigens, for isolating specific HSV infected cells or fragments of the virus, for vaccine preparation, or the like.

Anti-CD33 Antibodies

There are about 10,000–15,000 new cases of myeloid (also called non-lymphocytic or granulocytic) leukemia in the U.S. per year (Cancer Facts & Figures, American Cancer Society, 1987). There are two major forms of myeloid leukemia: acute myelogenous leukemia (AML) and chronic myelogenous leukemia (CML). Despite treatment with chemotherapy, long-term survival in patients with AML is less than 10–20% (Clarkson et al., CRC Critical Review in Oncology/Hematology 4, 221 (1986)), and survival with CML and related diseases such as chronic myelomonocytic leukemia (CMML), chronic monocytic leukemia (CMMOL) and myelodysplastic syndrome (MDS) is even lower.

The p67 protein or CD33 antigen is found on the surface of progenitors of myeloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or non-hematopoietic cells (see, Leucocyte Typing III, ed. by A. J. 55 McMichael, Oxford University Press, pp. 622–629 (1987), which is incorporated herein by reference). Antibodies that are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are 60 incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference). The reactivity of M195 with a wide 65 variety of cells and tissues was tested. Among normal cells, M195 was reported to bind only to some monocytes and

myeloid progenitor cells. The research also reported that it does not bind to other hematopoietic cells or to non-hematopoietic tissues. M195 bound to cells of most cases of AML and all cases of CML in myeloblastic phase.

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A phase I clinical trial of M195 in AML has been conducted (Scheinberg et al., Proc. ASCO 9, 207 (1990)). M195 radiolabeled with iodine-131 was found to rapidly and specifically target leukemic cells in both the blood and bone marrow

Unfortunately, the use of non-human monoclonal antibodies such as M195 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CD33 antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of myeloid leukemiarelated human disorders, the compositions containing humanized immunoglobulins specifically capable of binding to CD33 antigen. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the CD33 antigen at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CD33.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as cytosine arabinoside or daunorubicin active against leukemia cells, or complexed with a radionuclide such as iodine-131. All of these compounds will be particularly useful in treating leukemia and myeloid cellmediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CD33 related epitopes are provided. These immunoglobulins, which have binding affinities to CD33 of at least about $10^7~M^{-1}$, and preferably $10^8~M^{-1}$ to $10^{10}~M^{-1}$ or stronger, are capable of, e.g., destroying leukemia cells. The humanized immunoglobulins will have a human framework and will have one or

more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with CD33 antigen. In a preferred embodiment, one or more of the CDR's will come from the M195 antibody. Importantly, M195 does not bind to the ultimate hematopoietic stem cells, so M195 used in therapy will minimally interact with and destroy those cells, which are critical for generating all blood cells. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of myeloid cell-mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of CD33 antigen, such as monoclonal antibodies M195, L4B3, L1B2 or MY9. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody M195 are included in FIG. 34A and FIG. 34B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. 25

The antibodies of the present invention will typically find use individually in treating hematologic malignancies. For example, typical disease states suitable for treatment include AML, CML, CMML, CMMOL and MDS (see, generally, Hoffbrand & Pettit, Essential Haematology, Blackwell Scientific Publications, Oxford (1980)). The antibodies may also be used for bone marrow ablation prior to bone marrow transplant.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different myeloid antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include CD13, CD14, CD15, CD16 and CD34 (see, Leukocyte Typing III, op. cit., pp. 576–732).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include cytosine arabinoside and daunorubicin, but numerous additional agents (e.g., 6-thioguanine) well-known to those skilled in the art for leukemia treatment may also be utilized (see, Hoffbrund & Pettit., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill leukemia cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CD33 epitope.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, 60 the antibodies can be utilized for detection of CD33 antigens, for isolating specific myeloid cells, or the like.

It will be understood that although examples pertain to the M195 antibody, producing humanized antibodies with high binding affinity for the CD33 antigen is also contemplated 65 using CDR's from L4B3, L1B2, MY9 or other monoclonal antibodies that bind to an epitope of CD33.

Anti-CMV Antibodies

Cytomegalovirus is a major pathogen of immunocompromised individuals, especially bone marrow transplant recipients, organ tansplant recipients, and AIDS patients (see, generally, Fields et al., Eds., Virology, 2nd ed., Raven Press, New York pp. 1981–2010 (1990), which is incorporated herein by reference). Approximately 15% of bone marrow transplant patients develop CMV pneumonia, with an 85% mortality rate (Meyers, Rev. Inf. Dis. 11 (suppl. 7), S1691 (1989)). About 10% of AIDS patients develop severe CMV disease; and congenitally acquired CMV, often with significant morbidity and mortality, affects 1% of newborns (Fields, op. cit.).

The drug ganciclovir is effective against certain forms of CMV infection, notably chorioretinitis and gastroenteritis, but is not very effective against CMV pneumonia, and it has serious toxicity. Use of pooled human imunoglobulin preparations has shown some beneficial effect for prophylaxis of CMV in bone marrow transplant patients (Meyers, op. cit.), and a combination of high-dose immune globulin and ganciclovir has been reported effective against CMV pneumonia (Emanuel et al., Trans. Proc. XIX (suppl. 7), 132 (1987)). However, the marginal effectiveness, variable potency and high cost of commercial human immune globulin remain serious problems. Hence, there is a great need for new drugs effective against CMV.

CMV is a member of the herpesvirus family of viruses, and as such, has a large double-stranded DNA core, a protein capsid, and an outer lipid envelope with viral glycoproteins on its surface. At least 8 proteins have been detected on the envelope of CMV (Britt et al., J. Virol. 62, 3309 (1988)) and others have been predicted to exist based on the DNA sequence of CMV (Chee et al., Nature 344, 774 (1990)). Murine monoclonal antibodies have been produced against two especially significant CMV glycoproteins: gB, also called p130/55 or gp55-116, and gH, also called p86 (Rasmussen et al., Virology 163, 308 (1988) and Britt et al., op. cit., both of which are incorporated herein by reference) and shown to neutralize infectivity of the virus. Three other neutralizing antibodies to gH are designated CMV5, CMV109 and CMV115. Human monoclonal antibodies to CMV have also been produced (Ehrlich et al., Hybridoma 6, 151 (1987)).

In animal models, murine monclonal antibodies have been shown effective in treating infections caused by various viruses, including members of the herpesvirus family (see, e.g., Metcalf et al., Intervirol. 29, 39 (1988)). Hence, such antibodies may be useful in treatment of CMV infections.

Unfortunately, the use of non-human monoclonal antibodies such as CMV5 and CMV115 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with CMV antigens, as with many antigens, is difficult using typical human monoclonal antibody production techniques. Moreover, the human antibodies produced may lack certain desirable properties, such as high binding affinity and the ability to neutralize all clinical CMV strains. Similarly, utilizing recombinant DNA technology to produce so-called "human-

ized" or "reshaped" antibodies (see, e.g., Riechmann et al., Nature 332, 323 (1988) and EPO Publication No. 0239400, which are incorporated herein by reference), provides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of humanized 5 immunoglobulins specific for CMV antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of CMV-mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of CMV to its receptors and/or capable of binding to CMV 15 antigens. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, 20 with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to CMV at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be ²⁵ capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CMV.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such a acyclovir or ganciclovir active against CMV-infected cells, or complexed with a cytotoxic agent. All of these compounds will be particularly useful in treating CMV-mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CMV and CMVinfected cells are provided. These immunoglobulins, which have binding affinities to CMV specific antigens of at least about 10^7 M⁻¹, and preferably 10^8 M⁻¹ to 10^{10} M⁻¹ or stronger, are capable of, e.g., blocking CMV infection of cells. The humanized immunoglobulins will have a human 55 framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with a CMV antigen. In a preferred embodiment, one or more of the CDR's will come from the CMV5, or CMV109 or CMV115 antibodies. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of CMVmediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain

CDR's from an immunoglobulin capable of binding to a desired epitope of a CMV antigen, such as monoclonal antibodies CMV5 or CMV115. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and non-critical aminoacid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to CMV and produced in any convenient mammalian source, including, mice, rats, rabbits, or other vertebrate capable of producing antibodies by well known methods. Suitable source cells for the DNA sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection (Catalogue of Cell Lines and Hybridomas, Fifth edition (1985) Rockville, Md., U.S.A., which is incorporated herein by reference).

The antibodies of the present invention will typically find use individually in treating CMV-related disorders. For example, typical disease states suitable for treatment include CMV pneumonia, neonatal CMV infection, CMV mononucleosis and CMV-related chorioretinitis and gastroenteritis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different CMV antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include the gB and gH proteins.

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include acyclovir or ganciclovir, but numerous additional agents well-known to those skilled in the art for CMV treatment may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill CMV-infected cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CMV epitope. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see Chaudhary et al., Nature 339, 394 (1989)).

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 1 to 50 mg per dose. A preferred prophylactic use is for the prevention of CMV infection in immunocompromised patients, such as organ or bone marrow transplant recipients.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CMV antigens, for isolating specific CMV-infected cells, or the like.

In particular, the same method may be used to produce a humanized CMV109, CMV115 or other anti-CMV antibody as used to produce humanized CMV5 herein.

Anti-y-IFN Antibodies

In mammals, the immune response is mediated by several types of cells that interact specifically with foreign material, i.e., antigens. One of these cell types, B cells, is responsible for the production of antibodies. Another cell type, T cells, include a wide variety of cellular subsets that destroy virally infected cells or control the in vivo function of both B cells and other hematopoietic cells, including T cells. A third cell type, macrophages, process and present antigens in conjunction with major histocompatibility complex (MHC) proteins to T cells. Communication between these cell types is mediated in a complex manner by lymphokines, such as interleukins 1-6 and γ -IFN (see, generally, Paul, W. E., ed., Fundamental Immunology, 2nd ed., Raven Press, New York (1989), which is incorporated herein by reference.)

One important lymphokine is γ -IFN, which is secreted by some T cells. In addition to its anti-viral activity, γ -IFN stimulates natual killer (NK) cells, activates macrophages, and stimulates the expression of MHC molecules on the surface of cells (Paul, op. cit., pp. 622–624). Hence γ -IFN generally serves to enhance many aspects of immune function, and is a logical candidate for a therapeutic drug in cases where such enhancement is desired, e.g., in treating cancer. Conversely, in disease states where the immune system is over-active, e.g., autoimmune diseases and organ transplant rejection, antagonists of γ -IFN may be used to treat the disease by neutralizing the stimulatory effects of γ -IFN.

One class of effective antagonists of γ -IFN are monoclonal antibodies that bind to and neutralize it (see, e.g., Van der Meide et al., J. Gen. Virol, 67, 1059 (1986)). In in vitro 50 and in vivo mouse models of transplants, anti-y-IFN antibodies have been shown to delay or prevent rejection (Landolfo et al., Science 229, 176 (1985) and Rosenberg et al., J. Immunol. 144, 4648 (1990), both of which are incorporated herein by reference). Treatment of mice prone 55 to develop a syndrome like systemic lupus erythematosus (SLE) with a monoclonal antibody to γ -IFN significantly delayed onset of the disease (Jacob et al., J. Exp. Med. 166, 798 (1987)). Under some conditions, an anti-γ-IFN antibody alleviated adjuvant arthritis in rats (Jacob et al., J. Immunol. 142, 1500 (1989)), suggesting that anti-γ-IFN may be effective against some cases of rheumatoid arthritis in human patients. Multiple sclerosis (MS) in patients is made worse by treatment with γ-IFN (Panitch et al., Neurology 36 (suppl. 1), 285 (1986)), so an anti-γ-IFN antibody may alleviate MS. Thus, an anti-γ-IFN antibody may be effective in treating these and other autoimmune diseases.

For treatment of human patients, a murine monoclonal that binds to and neutralizes human γ -IFN (see, e.g., Yamamoto et al., Microbiol. Immunol. 32, 339 (1988)) may be used. Another murine monoclonal antibody designated AF2 that neutralizes human γ -IFN, and inhibits binding of γ -IFN to its cellular receptor, is disclosed herein. Unfortunately, the use of non-human monoclonal antibodies such as AF2 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, have a relatively short circulating half-life in humans, and lack other important immunoglobulin functional characteristics when used in humans

The present invention provides novel compositions useful, for example, in the treatment of human autoimmune disorders, the compositions containing humanized immunoglobulins specifically capable of binding to γ-IFN. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally ioined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to γ -IFN at affinity levels stronger than about $10^7 \,\mathrm{M}^{-1}$. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to y-IFN.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as a non-steroidal anti-inflammatory drug, a corticosteroid, or an immunosuppressant. All of these compounds will be particularly useful in treating autoimmune disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form; which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with $\gamma\text{-IFN}$ epitopes are provided. These immunoglobulins, which have binding affinities to $\gamma\text{-IFN}$ of at least about $10^7~M^{-1}$, and preferably $10^8~M^{-1}$ to $10^{10}~M^{-1}$ or stronger, are capable of, e.g., neutralizing human $\gamma\text{-IFN}$. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with $\gamma\text{-IFN}$. In a preferred embodiment, one or more of the CDR's will come from the AF2 antibody. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of autoimmune disorders in human patients by a variety of techniques.

The antibodies of the present invention will typically find use individually in treating autoimmune conditions. For

example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid 5 arthritis, systemic lupus erythematosus, and myasthenia gravis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with other lymphokines or lymphokine receptors. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include interleukins 1 through 10 and the p55 and p75 chains of the IL-2 receptor (see, Waldmann, Annu. Rev. Biochem. 58, 875 (1989) and Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), both of which are incorporated herein by reference). Other antigens include those on cells responsible for the disease, e.g., the so-called "Clusters of Differentiation" (Leucocyte Typing III, ed. by A. J. McMichael, Oxford University Press (1987), which is incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include non-steroidal anti-inflammatory agents (e.g., aspirin, ibuprofen), steroids (e.g., prednisone) and immunosuppressants (e.g., cyclosporin A, cytoxan), but numerous additional agents well-known to those skilled in the art may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins, e.g., to kill $\gamma\text{-IFN}$ -secreting cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a $\gamma\text{-IFN}$ epitope.

Humanized antibodies of the present invention can further $_{40}$ find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of $\gamma\textsc{-}\textsc{IFN}$ antigens, or the like.

The following examples are offered by way of illustration, not by limitation.

EXPERIMENTAL

Example 1

Humanized anti-Tac antibody

Design of genes for humanized anti-Tac light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, E. Kabat et al., U.S. Dept. of Health and Human Services, 1983) was used to 55 provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain variable region of anti-Tac is more homologous to the heavy chain of this antibody than to any other complete heavy chain variable region sequence in the National Biomedical Foundation 60 Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence (FIG. 1B, upper lines; see, commonly assigned U.S. Ser. Nos. 07/223,037 filed Sep. 28, 1988, and 07/181,862 filed Apr. 15, 1988, both of which are 65 now abandoned and which are incorporated herein by reference) was aligned with the sequence of the Eu heavy chain

38

(FIG. 1B, lower lines). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of four categories defined above, in which case the anti-Tac amino acid was selected:

- (1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31–35, 50–66, 99–106);
- (2) The Eu amino acid was rare for human heavy chains at that position, whereas the anti-Tac amino acid was common for human heavy chains at that position (amino acids 27, 93, 95, 98, 107–109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67); or
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68). Amino acid #27 is listed in category (2) because the acceptor Eu amino acid Gly is rare, and the donor anti-Tac amino acid Tyr is chemically similar to the amino acid Phe, which is common, but the substitution was actually made because #27 also fell in category (4). Although some amino acids fell in more than one of these categories, they are only listed in one. The amino acids in the humanized heavy and light chains are numbered according to the lower lines of FIG. 1A and FIG. 1B.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 1A, lower lines). The Eu amino acid was selected at each position for the humanized sequence, unless the position again fell into one of the categories (1)–(4):

- (1) CDR's (amino acids 24-34, 50-56, 89-97);
- (2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63);
- (3) Adjacent to CDR's (no amino acids; Eu and anti-Tac were already the same at all these positions); or
- (4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy and light chain genes were selected as follows:

- (1) The nucleotide sequences code for the amino acid sequences chosen as described above;
- (2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies;
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals; and
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes

To synthesize the heavy chain, four oligonucleotides were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing.

Together, the oligonucleotides cover the entire humanized heavy chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide, the following components were added in a final volume of 100 ul:

10	ul	annealed oligonucleotides
0.16	mM each	deoxyribonucleotide
0.5	mM	ATP
0.5	mM	DTT
100	ug/ml	BSA
3.5	ug/ml	T4 g43 protein (DNA polymerase)
25	ug/ml	T4 g44/62 protein (polymerase
	-	accessory protein)
25	ug/ml	45 protein (polymerase accessory
		protein)

The mixture was incubated at 37° C. for 30 min. Then 10 u of T4 DNA ligase was added and incubation at 37° C. resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba I, to the reaction was added 30 50 ul of 2×TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 u of Xba I in 5 ul. The reaction was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods. 35

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence.

To synthesize the light chain, four oligonucleotides JFD1, JFD2, JFD3, JFD4 were synthesized. Two of the oligonucleotides are part of each strand of the light chain, and each 40 oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing. Together, the oligonucleotides cover the entire humanized light chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were combined in 20 ul sequence buffer (40 mM Tris-HCl, pH 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), 50 heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 u of sequenase (US Biochemicals) was added, in a final 55 volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 overlap and therefore in each of the synthesized DNAs). The 60 reactions were run on polyacrylamide gels, and the Xba I—Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then inserted into the Xba I site of the vector $pV\gamma 1$ (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) in the correct orientation by standard methods, to produce the plasmid pHuGTAC1. This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I—Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVx1 (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the circular form: vector—Xba I—fragment 1—Hind III—fragment 2—Xba I—vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC, therefore contains the complete humanized light chain and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTAC1 and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with fluoresceinconjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (FIG. 7A), clearly show that the humanized antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells, giving similar results.

For the next experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment was performed. About 5×105 HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess.

Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycoerythrin- conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10–40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrin-conjugated avidin that bound in the last step, thus decreasing 5 fluorescence (FIG. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater 10 affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more.

Example 2

A second humanized anti-Tac antibody Higher level expression of the humanized anti-Tac antibody

Three new plasmid vectors were prepared for expression of the humanized antibodies. The plasmid pVg1 (FIG. 9A) contains a human cytomegalovirus IE1 promoter and 20 enhancer (Boshart et al., Cell 41, 521 (1985), which is

incorporated herein by reference), the human genomic Cyl segment including part of the preceding intron, and the hygomycin gene (Blochlinger et al., *Mol. Cell. Biol.* 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 9B) is similar to pVg1 but contains the human genomic Cx segment and the gpt gene. The plasmid pVg1-dhfr was constructed similarly to pVg1 but contains a dihydrofolate reductase (dhfr) gene (Simonsen et al., *Proc. Natl. Acad. Sci. U.S.A.* 80, 2495 (1984), 30 which is incorporated herein by reference) in place of the

hygomycin gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and 55 cloned into the Xba I sites of the plasmid vectors pVk and pVg1. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were 40 then transfected with the plasmid encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture 45 supernatant of transfected cells by protein A sepharose chromatography.

Construction of the second humanized anti-Tac antibody

To determine whether it was actually necessary to use the mouse anti-Tac amino acids in categories (2)–(4) in the 50 humanized anti-Tac antibody to retain binding affinity, a second humanized anti-Tac antibody was constructed. In the second antibody, only mouse anti-Tac amino acids in Category (1), i.e., in the CDR's themselves, were used, with all other amino acids coming from the human Eu framework. 55 For purposes of this discussion, the original humanized anti-Tac antibody will be called the "PDL humanized antibody," and the second humanized antibody will be called the "CDR-only humanized antibody." The amino acid sequences of the PDL and CDR-only humanized antibody 60 (variable regions) are compared in FIG. 10A and FIG. 10B.

The CDR-only humanized anti-Tac heavy and light chain variable (V) region gene segments were constructed in essentially the same manner as the light chain of the PDL humanized anti-Tac immunoglobulin, as described above. 65 Specifically, each V region gene segment was synthesized in two halves. For each half, two overlapping, opposite-strand

oligonucleotides, approximately 110 to 130 bases in length (FIG. 11A and FIG. 11B), were annealed and extended with sequenase (U.S. Biochemicals). The resulting double strand fragments were digested with either Xba I and Hind III (light chain) or Xba I and Sal I (heavy chain) and inserted into plasmid pUC19. Clones with the correct sequence were identified by DNA sequencing. Complete heavy and light chain genes were generated by inserting the V region halves into the Xba I sites of pVg 1 and pVk respectively by three-fragment ligation.

The CDR-only humanized antibody was expressed in the same manner as the PDL humanized antibody, by transfecting first the light chain containing plasmid and then the heavy chain containing plasmid into SP2/0 cells. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody, which was purified by protein A sepharose chromatography. Antibody concentration was determined by ELISA using purified PDL humanized antibody as a standard. That the purified CDR-only humanized antibody is assembled into H₂L₂ tetramers as expected was shown by analysis using reducing and non-reducing polyacrylamide gel electrophoresis.

The ability of the CDR-only humanized immunoglobulin to bind to the IL-2 receptor was assessed by fluorescence staining. Approximately 3.4×10⁵ HUT-102 cells, which are known to highly express the IL-2 receptor on their surface, were incubated with 200 ng of either the PDL or CDR-only humanized antibody, washed, and then incubated with fluorescein-conjugated goat anti-human IgG antisera. Cell fluorescence was measured by flow cytometry with a FACScan (Becton Dickinson). As shown in FIG. 12, the PDL humanized antibody strongly stained the cells. However, staining by the CDR-only antibody was indistinguishable from staining by the negative control antibody humanized Fd79, which binds the gB glycoprotein of herpes simplex virus and not HUT-102 cells. Hence, by this assay, the CDR-only humanized antibody does not detectably bind the IL-2 receptor.

Binding of the PDL and CDR-only humanized anti-Tac antibodies to the IL-2 receptor were also compared in a competitive binding assay. Approximately 4×10⁵ HUT-102 cells were incubated with 1.5 ng of radioiodinated mouse anti-Tac antibody (7×106 cpm/ug) and varying amounts of each humanized antibody (4 to 512 ng) in 200 ul total volume of binding buffer (RPMI 1040 medium, 10% fetal calf serum, 10 ug/ml murine IgG2a, 0.1% sodium azide). After incubation for 2 hours at 0° C., 800 ul of binding buffer was added, cells were collected by centrifugation and radioactivity was measured. The relative binding by the two humanized antibodies and by mouse anti-Tac is shown in a plot of bound/free labelled antibody versus competitor concentration (FIG. 13). The PDL humanized anti-Tac antibody affinity for IL-2 receptor is essentially equal to that of the mouse anti-Tac antibody, because it competes about equally well. But competition by the CDR-only humanized anti-Tac antibody to IL-2 receptor was undetectable at the antibody concentrations used, indicating a binding affinity reduction of at least 100-fold as compared to the PDL humanized anti-Tac antibody. Because the sequences of the PDL and CDR humanized anti-Tac antibodies differ only at positions where mouse framework residues in categories (2)–(4) were used in the PDL molecule, we conclude that at least one of these mouse framework residues are essential for high affinity binding.

Example 3

Construction of 5 other humanized antibodies Cloning of heavy and light chain cDNAs

Five other humanized antibodies were designed and produced using the principles and categories disclosed herein. The antibodies are Fd79 and Fd138-80 which respectively bind to the gB and gD glycoproteins of herpes simplex virus (Metcalf et al., *Intervirology* 29, 39 (1988)), M195 (Tansimoto et al., *Leukemia* 3, 339 (1989)) which binds to the CD33 antigen, mik-β1 (Tusdo et al., *Proc. Natl. Acad. Sci. U.S.A.* 86, 1982 (1989)) which binds to the p75 chain of the IL-2 receptor, and CMV5 which binds to the gH glycoprotein of cytomegalovirus.

cDNAs for the heavy chain and light chain variable domain genes of each antibody were cloned using anchored polymerase chain reactions (Loh et al., *Science* 243, 219 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that 15 hybridized to the dG tails and contained EcoRI sites (Scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For each antibody, at least two heavy chain and two kappa clones were sequenced and 20 found to have the same sequence. The deduced amino acid sequences of the mature light and heavy chain variable regions are shown in FIGS. 2A–6B, upper lines. Design of humanized antibodies

In order to retain high binding affinity of the humanized 25 antibodies, the principles and categories described above were utilized when designing the antibodies. Based on high sequence homology, human antibodies were selected to provide both the acceptor light and heavy chain human frameworks for the mouse antibodies, as follows: human 30 Pom for Fd79, human Eu for Fd138-80, human Eu for M195, human Lay for mik- β 1, and human Wol for CMV5.

The computer programs ABMOD and ENCAD (Levitt, *J. Mol. Biol.*, 168, 595 (1983) and Zilber et al., *Biochemistry* 29, 10032 (1990), both of which are incorporated herein by 35 reference) was used to construct a model of the variable region of each mouse antibody. The model was used to determine the amino acids in each framework that were close enough to the CDR's to potentially interact with them (category 4 above). For each antibody, the positions found to 40 fall in the categories (1)–(5) defined above are given in Table 1, numbered as in FIGS. 2A–6B.

TABLE 1

Category	Light Chain	Heavy Chain
	Fd79 Antibod	l <u>y</u>
1	24 20 54 50 02 100	21 25 50 66 00 111
2	24–38, 54–50, 93–100 9, 45, 46, 83	31–35, 50–66, 99–111
3	53	82, 112
4	53	112 97
5	81	97
3		a.d.,
	Fd138-80 Antib	ouy
1	24-34, 50-56, 89-97	31-35, 50-66, 99-110
2	48. 63	93, 98, 111, 112,
-	-10, 05	113, 115
3		30, 67, 98, 111
4	36, 48, 87	
-	30, 40, 07	27, 30, 37, 48, 67, 68, 98
	M195 Antiboo	
	MIT95 Allubot	iy
1	24-38, 54-60, 93-101	31-35, 50-66, 95-105
2	10, 52, 67, 110	93, 95, 98, 106, 107
-	10, 52, 07, 110	108, 110
3		30, 67, 98, 106
4	40, 52, 74	27, 30, 48, 68, 98
	mik-β1 Antibo	
	nik-pi Aliubo	<u>uy</u>
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108

TABLE 1-continued

Category	Light Chain	Heavy Chain
2	13	84, 89, 90
3		30, 49
4	70	29, 30, 72, 73
5	41	,,,
	CMV5 Antibo	ody
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108
2	<u> </u>	69, 80
3	49	30
4	49	24, 27, 28, 30, 97
5	_	5

In designing each humanized antibody, at each position the amino acid was selected to be the same as in the human acceptor sequence, unless the position fell in categories (1)–(4), in which case the amino acid from the mouse donor sequence was used, or in category (5), in which case an amino acid typical for human sequences at that position was used.

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including signal peptides typically from the mouse antibody chains, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included splice donor signals typical for immunoglobulin genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 base long with a 15-20 base overlap. Double stranded DNA fragments were synthesized with Klenow or Taq polymerase or sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of pVg1 (heavy chains of Fd79 and Fd138-80) or pVg1-dhfr (heavy chains of M195, mik-\(\beta\)1, CMV5) or pVk (all light chains) 45 expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.).

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2M Glycine-HCl, pH 3.0 and neutralized with 1M Tris pH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of the humanized antibodies

The binding of the humanized antibodies to cell types expressing the corresponding antigens was tested: HSV-infected cells for Fd79 and Fd138-80, U937 cells for M195, YTJB cells for mik-β1 and CMV-infected cells for CMV5. By fluorocytometry, the humanized antibodies bind approximately as well as the original mouse antibodies and the

corresponding chimeric antibodies. Moreover, the humanized antibodies compete approximately as well as the corresponding mouse antibodies against the radiolabeled mouse antibodies for binding to the cells, so the humanized antibodies have approximately the same binding affinity as the 5 mouse antibodies, typically within about 2 fold or better, see, e.g., Table 2.

TABLE 2

Binding affinities of	murine and humanized	antibodies.	10
Mouse $K_a (M^{-1})$	Humanized $K_a (M^{-1})$		
Fd79 (anti-gB) Fd138-80 (anti-gD)	1.1×10^8 5.2×10^7	5.3×10^{7} 4.8×10^{7}	15

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other antibodies. In comparison to other monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 4

Design of genes for anti-Tac human-like light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, Kabat, E., et al., U.S. Dept. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain of anti-Tac is 35 more homologous to the heavy chain of this antibody than to any other heavy chain sequence in the National Biomedical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence was aligned with the 40 sequence of the Eu heavy chain (FIG. 15). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of the following categories, in which case the anti-Tac amino acid was selected.

- (1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31–35, 50–66, 99–106);
- (2) The Eu amino acid was unusual for human heavy chains at that position, whereas the anti-Tac amino acid was typical for human heavy chains at that position (amino acids 27, 93, 95, 98, 107–109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67).
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68).

Some amino acids fell in more than one of these categories but are only listed in one.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 16). The Eu amino acid was selected at each position, unless the position again fell into one of the categories (1)–(4), (with light chain replacing 65 heavy chain in the category definitions):

(1) CDRs (amino acids 24-34, 50-56, 89-97).

46

- (2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63).
- (3) Adjacent to CDRs (no amino acids; Eu and anti-Tac were already the same at all these positions).
- (4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy (FIG. 17) and light chain (FIG. 18) genes were selected as follows:

- (1) the nucleotide sequences code for the amino acid sequences chosen as described above.
- (2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies.
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals.
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes

To synthesize the heavy chain, four oligonucleotides HES12, HES13, HES14, HES15 (FIG. 19A) were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. 19B). Together, the oligonucleotides cover the entire humanized heavy chain (FIG. 17) with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide (FIG. 19B), the following components were added in a final volume of 100 ul:

_	10	ul mM each	annealed oligonucleotides deoxyribonucleotide	
0		mM	ATP	
	0.5	mM	DTT	
	100	ug/ml	BSA	
	3.5	ug/ml	T4 g43 protein (DNA polymerase)	
	25	ug/ml	T4 g44/62 protein (polymerase accessory protein)	
5	25	ug/ml	45 protein (polymerase accessory protein)	
			protess,	

The mixture was incubated at 37° C. for 30 min. Then 10 U of T4 DNA ligase was added and incubation at 37° C. resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba I, to the reaction was added 50 ul of 2×TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 U of Xba I in 5 ul. The reaction was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods.

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence (FIG. 17).

To synthesize the light chain, four oligonucleotides JFD1, JFD2, JFD3, JFD4 (FIG. **20**A) were synthesized. Two of the 5 oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. **20**B). Together, the oligonucleotides cover the entire humanized light chain (FIG. **18**) with a few extra nucleotides at each end to allow 10 cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these olignucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were combined in 20 ul sequenase buffer (40 mM Tris-HCl, pH 15 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal, JFD3 and JFD4 were treated in the same way. Each reaction was made $10\,\mbox{mM}$ in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 U of sequenase (US Biochemicals) was added, in a final volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 25 overlap and therefore in each of the synthesized DNAs; FIG. 20B). The reactions were run on polyacrylamide gels, and the Xba I-Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and 30 correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then 35 inserted into the Xba I site of the vector pVγI in the correct orientation by standard methods, to produce the plasmid pHuGTAC1 (FIG. 21). This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I—Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVk1 was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the 45 circular form: vector—Xba I—fragment 1—Hind III—fragment 2—Xba I—vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC (FIG. 22), therefore contains the complete humanized light chain (FIG. 18) 50 and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTAC1 and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the 55 plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids (FIGS. 21, 22) by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was 60 incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with fluorescein-conjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (FIG. 7A), clearly show that the humanized 65 antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the

original mouse anti-Tac antibody was also used to stain these cells (FIG. 7B and FIG. 7C), giving similar results.

For further experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment was performed. About 5×10⁵ HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess. Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycoerythrin-conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10–40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrin-conjugated avidin that bound in the last step, thus decreasing fluorescence (FIG. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more. Biological properties of the humanized antibody

For optimal use in treatment of human disease, the humanized antibody should be able to destroy T-cells in the body that express the IL-2 receptor. One mechanism by which antibodies may destroy target cells is antibody-dependent cell-mediated cytotoxicity, abbreviated ADCC (Fundamental Immunology, Paul, W., Ed., Raven Press, New York (1984), at pg. 681), in which the antibody forms a bridge between the target cell and an effector cell such as a macrophage that can lyse the target. To determine whether the humanized antibody and the original mouse anti-Tac antibody can mediate ADCC, a chromium release assay was performed by standard methods. Specifically, human leukemia HUT-102 cells, which express the IL-2 receptor, were incubated with 51Cr to allow them to absorb this radionuclide. The HUT-102 cells were then incubated with an excess of either anti-Tac or humanized anti-Tac antibody. The HUT-102 cells were next incubated for 4 hrs with either a 30:1 or 100:1 ratio of effector cells, which were normal purified human peripheral blood mononuclear cells that had been activated by incubation for about 20 hrs with human recombinant IL-2. Release of 51Cr, which indicated lysis of the target HUT-102 cells, was measured and the background subtracted (Table 3). The results show that at either ratio of effector cells, anti-Tac did not lyse a significant number of the target cells (less than 5%), while the humanized antibody did (more than 20%). Hence, the humanized antibody is likely to be more efficacious than the original mouse antibody in treating T-cell leukemia or other T-cell mediated diseases.

TABLE 3

Percent 51Cr rele	Percent 51Cr release after ADCC		
	Effect	or: Target ratio	
Antibody	30:1	100:1	
Anti-Tac Humanized anti-Tac	4% 24%	<1% 23%	

Higher level expression of the humanized anti-Tac antibody
Two new plasmid vectors were prepared for expression of
the humanized antibody. The plasmid pVg1 (FIG. 9A)
contains a human cytomegalovirus IE1 promoter and
enhancer (Boshart et al., Cell 41, 521 (1985)), the human 15
genomic Cγ1 segment including part of the preceding intron,
and the hygomycin gene (Blochlinger et al., Mol. Cell. Biol.
4, 2929 (1984), which is incorporated herein by reference)
for selection. The plasmid pVk (FIG. 9B) is similar to pVg1
but contains the human genomic Cκ segment and the gpt 20
gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and cloned into the Xba I sites of the plasmid vectors pVk and 25 pVG1. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were then transfected with the plasmid encoding the heavy chain 30 followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture supernatant of transfected cells by protein A sepharose 35 chromatography.

From the foregoing, it will be appreciated that the human-like immunoglobulins of the present invention offer numerous advantages of other human IL-2 receptor-specific antibodies. In comparison to anti-Tac mouse monoclonal 40 antibodies, the present human-like immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 5

Design of genes for mikβ1 humanized light and heavy chains

To exert its biological effects, IL-2 interacts with a 50 specific high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), which is incorporated herein by reference). The human IL-2 receptor is a complex 55 multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55 kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. 60 Acad. Sci. U.S.A., 83, 9694 (1986) and Sharon et al., Science 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 65 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

50

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. The antibody, mik- β 1, binds to the p75 chain (Tsudo et al., *Proc. Nat. Acad. Sci. U.S.A.* 86, 1982 (1989), which is incorporated herein by reference).

Cloning of heavy chain and light chain cDNA

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., *Science* 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC19 vector for sequencing. For mik-β1, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 23A and FIG. 23B. Construction and expression of chimeric antibody

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cyl segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. U.S.A. 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVg1-dhfr but contains the human genomic Ck segment and the gpt gene. Derivatives of the mik-β1 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, C. Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric mik- β 1 antibody was shown to bind to YTJB cells, which express the p75 antigen, by flow cytometry (FIG. 25).

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., *Proc. Natl. Acad. Sci. U.S.A.* 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence database (performed with the MicrorGenie Sequence Analysis Software (Beckman)), the antibody Lay was chosen to provide the framework sequences for humanization of mik-β1.

15

The computer program ENCAD (M. Levitt, *J. Mol. Biol.* 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the mik-β1 variable region. The model was used to determine the amino acids in the mik-β1 framework that were close enough to the CDRs to potentially interact with them (category 4 below). To design the humanized light and heavy chain mik-β1 variable regions, at each position the amino acid was chosen to be the same as in the Lay antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Lay amino acid was unusual for human antibodies at that position, whereas the mik-β1 amino acid was typical for human antibodies at that position.
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDRs).

For positions in these categories, the amino acid from the (mouse) mik- $\beta 1$ antibody was used. In addition, a position was in the fifth category if

(5) The Lay amino acid was highly unusual for human antibodies at that position, and the mik-β1 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 4. Some amino acids may be in more than one category. The final sequences of the humanized mik- β 1 light and heavy chain variable domains are shown in FIG. 26A and FIG. 26B, compared with the Lay sequences.

TABLE 4

Category	Light Chain	Heavy Chain
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108
2	13	84, 89, 90
3	30, 49	
4	70	29, 30, 72, 73
5	41	1

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse mik-\(\beta\)1 45 chains (FIG. 23A and FIG. 23B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an 50 XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the 55 splice donor signal (FIG. 27A and FIG. 27B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 base long with about a 20 base overlap. Double stranded DNA fragments were synthesized with sequenase from each pair 60 of oligonucleotides, digested with restriction enzymes, ligated to pBluescriptII KS (+) (Stratagene) vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors. In vitro mutagenesis 65 was used to change an Ala amino acid originally encoded by oligonucleotide wps54 to the Glu (E) at position 1 of the

humanized heavy chain (FIG. 26B) by changing the nucleotides CT to AG. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by EiISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was cluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies

The humanized mik-β1 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to YTJB cells, which express p75 chain at a high level, in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 25), showing that it recognizes the same p75 protein.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse mik- β 1 antibody (FIG. 28). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in *Fundamental Immunology* (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The binding affinity of the humanized mik- β 1 antibody was within about 2-fold of the affinity of the mouse mik- β 1 antibody.

The ability of humanized mik-β1 plus humanized anti-Tac antibody to inhibit IL-2 stimulated proliferation of human lymphocytes was determined. Human mononuclear cells, collected from human blood by centrifugation on Ficoll-Paque (Pharmacia), were diluted to 2×106 cells/ml in RPMI medium+10% fetal calf serum (FCS). A 1/200 volume of phytohemagglutinin P (Difco) was added and the cells were incubated for 4 days. The cells were incubated an additional days in RPMI+10% FCS+10 u/ml IL-2. 10⁵ of these PHA activated blasts were then incubated with or without 2 ug each of humanized mik- $\beta1$ and humanized anti-Tac in $15\overline{0}$ μl of RPMI+10% FCS in wells of a 96-well plate for 1 hr, to which various dilutions of IL-2 (Amgen) were then added in 50 μ l medium. The cells were incubated 48 hr, 0.5 μ Ci methyl-3H-thymidine (Amersham, 82 Ci/mmol) was added, and the cells were incubated 24 hr. Cells were harvested with a cell harvester and radioactivity determined. The combination of the antibodies greatly inhibited proliferation of the cells in response to IL-2 (FIG. 29), suggesting a combination of the antibodies will have strong immunosuppressive properties. Humanized mik-β1 plus humanized anti-Tac inhibited proliferation much more strongly than did either anti-

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other p75 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 6

Design of genes for Fd79 and Fd138-80 humanized light and heavy chains

Exemplary DNA sequences coding for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, are shown in FIG. **30**A through FIG. **30**D.

Cloning of heavy chain and light chain cDNA

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain regions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). This method yields clones with authentic variable domain sequences, in contrast to other methods using mixed primers designed to anneal to the variable domain sequence (J. W. 15 Larrick et al., Bio/Technology 7, 934 (1989) and Y. L. Chiang et al., BioTech. 7, 360 (1989)). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For Fd79, two gamma-1 specific and 5 kappa specific clones were 20 sequenced. The two gamma-1 specific clones are identical in sequence. This heavy chain cDNA fragment encodes a signal peptide of 19 amino acids, a V region in mouse heavy chain subgroup IIIB, a D segment, and a J_H 1 segment with 4 alterations compared to the genomic $J_H 1$ sequence. The 25 deduced amino acid sequence is shown in FIG. 30A.

The five kappa specific clones belong to two groups. Two clones are identical and encode a kappa chain in which the conserved amino acid 23 cysteine has been substituted by a tyrosine, probably representing the non-productive allele. 30 The other three clones have an identical sequence encoding a signal peptide sequence of 20 amino acids, a V region in mouse kappa chain subgroup III, and a $J_k 2$ segment with a single alteration compared to the genomic $J_k 2$ sequence (FIG. 30B). The validity of the heavy chain and the kappa 35 chain sequences was subsequently confirmed by the construction and expression of a chimeric antibody as discussed below.

The heavy chain and the kappa chain of Fd138-80 were cloned similarly. Three clones each of the heavy chain and 40 the kappa chain were sequenced. All three heavy chain clones have an identical sequence encoding a signal peptide sequence of 19 amino acids, a V region in mouse heavy chain subgroup II, a D segment and the J_H3 segment (FIG. 30C). The three kappa clones are also identical in sequence. This light chain fragment encodes a signal peptide sequence of 20 amino acids, a V region gene in mouse kappa chain subgroup V and the J_L5 segment (FIG. 30D). Both chains shown no irregularities in coding sequence; their validity was subsequently confirmed by construction and expression 50 of a chimeric antibody.

Construction and expression of chimeric antibodies

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1 (FIG. 9A) contains a human cytomegalovirus IE1 55 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγl segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 60 9B) is similar to pVg1 but contains the human genomic Cκ segment and the gpt gene. Derivatives of the Fd79 and Fd138-80 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the 65 ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained

splice donor signals and XbaI sites (See, C. Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibodies, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric Fd79 and Fd138-80 antibodies were shown to bind to HSV-1 infected vero cells by flow cytometry. Viral neutralization assays also indicated that the chimeric antibodies retain the neutralization activities of the murine antibodies (data not shown, but see below for similar results with humanized antibodies).

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Pom was chosen to provide the framework sequences for humanization of Fd79.

The computer program ENCAD (Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the Fd79 variable region. Inspection of the refined model of murine Fd79 revealed two amino acid residues in the framework that are close enough to have significant contacts with the CDR residues (Table 5). Lys in light chain position 49 has contacts with 3 amino acids in CDR2 of the light chain (L50 Tyr, L53 Asn, L55 Glu) and 2 amino acids in CDR3 of the heavy chain (H99 Asp, H100 Tyr). Leu in heavy chain position 93 also shows interactions with 2 amino acids in CDR2 of the heavy chain (H35 Ser, H37 Val) and an amino acid in CDR3 of the heavy chain (H100C Phe). Hence, L49 Lys and H93 Leu were retained in the construction of humanized Fd79, as their replacement with human Pom framework residues would be likely to introduce distortions into the CDRs. Also, 7 other residues in the Pom framework (5 in the light chain and 2 in the heavy chain) were substituted with common human residues (identical to the murine Fd79 sequence in 6 of the choices) because of their rare occurrence in other human antibodies. The elimination of unusual amino acids in the framework may further reduce immunogenicity. The murine sequences and the corresponding humanized sequences are shown in FIG. 30A and FIG. 30B. Substituted residues in the Pom framework are underlined.

TABLE 5

Residues in the framework sequence showing contacts with residues in the hypervariable regions.		
Residue No. I Amino Acid Contacting CDR residues ²		Contacting CDR residues ²
Fd79		
L49 H93 Fd138-80	Lys Leu	L50Y, L53N, L55E, H99D, H100Y H35S, H37V, H100CF
L36 H27 H30 H48 H66 H67	His Tyr Tyr Phe Lys Ala	L34V, L89Q H32H, H34I H32H, H53R H63F H63F H63F

¹The amino acid residues are numbered according to the Kabat system (E.A Kabat et al., Sequences of Proteins of Immunological Interest, National Institutes of Health, Bethesda, MD (1987)): the first letter (H or L) stands for the heavy chain or light chain. The following number is the residue number, The last letter is the amino acid one letter code.

²The hypervariable regions are defined according to Kabat: Light chain

²The hypervariable regions are defined according to Kabat: Light chain CDR1: residue 24-34; CDR2: 50-56; CDR3: 89-97. Heavy chain CDR1: 31-35; CDR2: 50-65; CDR3: 95-102.

Similarly, the murine heavy chain and light chain 25 sequences of Fd138-80 were subjected to sequence homology search against the NBRF protein sequence database. The sequences of the human antibody Eu were selected to provide the framework sequences for humanized Fd138-80. Inspection of a computer-generated model of Fd138-80 30 revealed 6 amino acid residues in the framework that are close enough to have important contacts with CDR residues. The residues and their contacting counterparts are listed in Table 5; these murine residues were retained in the construction of humanized Fd138-80. Two other residues (L87) 35 Phe and H37 Met) show significant contacts with L98 Phe, which is immediately adjacent to CDR3, so these two mouse residues were also retained. Eight amino acids in the Eu framework (2 in the light chain and 6 in the heavy chain) were substituted with the murine residues (which are also consistent with the human consensus residues) because of their rare occurrence in other human antibodies. The murine Fd138-80 sequences and the corresponding humanized sequences are shown in FIG. 30C and FIG. 30D. Substituted residues in the Eu framework are underlined.

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the signal peptides, generally utilizing codons 50 found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four 55 overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized 60 on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase, digested with restriction enzymes, ligated to pUC18 vector and sequenced. The two fragments 65 with the correct sequences were then ligated into the XbaI sites of pVg1 or pVk expression vectors.

The synthetic genes were then cloned into the pVg1 and pVk expression vectors. For each humanized antibody constructed, the heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibodies were purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies

The humanized Fd79 and Fd138-80 antibodies were characterized in comparison to the murine and chimeric antibodies. Both humanized antibodies bind to Vero cells infected with HSV-1 in a fluorocytometric analysis in a manner similar to the chimeric antibodies (FIG. 31A and FIG. 31B), suggesting that they recognize their respective viral antigens. To more quantitatively assess the binding activity, radioiodinated murine antibodies were bound to virally infected cells and Scatchard analysis performed.

The affinities of the humanized antibodies were determined by competition with the iodinated antibodies. Vero cells infected with HSV-1 were used as source of gB and gD antigens. Increasing amounts of competitor antibody (mouse or humanized) were added to 1.5 ng of radioiodinated tracer mouse antibody (2 uCi/ug) and incubated with 4×10⁵ infected Vero cells in 0.2 ml of binding buffer (PBS+2% FCS+0.1% azide) for 1 hr. at 4° C. Cells were washed and pelleted, and their radioactivities were measured. The concentrations of bound and free tracer antibody were calculated. The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in *Fundamental Immunology* (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference).

The measurements indicate that there is no significant loss of binding affinities in the humanized antibodies (Table 6). Specifically, there is an approximately 2-fold decrease in affinity in humanized Fd79 compared to the murine Fd79 (Ka of $5.3\times10^7~M^{-1}$ vs. $1.1\times10^8~M^{-1}$). The affinity of humanized Fd138-80 is comparable to that of the murine antibody (Ka of $4.8\times10^7~M^{-1}$ vs $5.2\times10^7~M^{-1}$).

TABLE 6

Binding affinities of murine and humanized antibodies.		
Mouse K _a (M ⁻¹)	$\begin{array}{c} \text{Humanized} \\ \text{K}_{\text{a}} \ (\text{M}^{-1}) \end{array}$	
Fd79 (anti-gB) Fd138-80 (anti-gD)	1.1×10^{8} 5.2×10^{7}	5.3×10^{7} 4.8×10^{7}

Murine Fd79 and Fd138-80 have been shown to neutralize HSV-1 in vitro without complement (J. Koga et al., Virology 151, 385 (1986)), so the neutralizing activities of the humanized antibodies were compared with the mouse antibodies. Serial dilutions of equal quantities of murine and humanized antibodies were incubated with virus for 1 hr. before inoculation onto Vero cells. After 4 days, cells were stained with neutral red to visualize plaques. Results from these plaque reduction assays indicated that both humanized Fd79 and Fd138-80 neutralize virus as efficiently as their murine counterparts (FIGS. 32A and B). Both humanized and murine Fd79 cause a 90% reduction of plaques at an

antibody concentration of 10 nM (1.5 ug/ml). Similarly, humanized and murine Fd138-80 were able to cause a 90% plaque reduction at equivalent levels.

The antibodies were also investigated for their ability to protect cells from viral spread in tissue culture. Vero cells 5 were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before addition of 10 ug/ml antibody. After four days, cells were stained with an anti-gB antibody for detection of viral antigens on infected cells. Results indicated that both murine and humanized Fd79 at 10 10 ug/ml protected culture cells from infection (FIG. 33A). However, neither murine nor humanized Fd138-80 were able to protect cells against viral spread (FIG. 33B), despite their ability to neutralize virus before inoculation. Both gB and gD are thought to be associated with cell fusion and 15 virus infectivity (W. Cai et al., J. Virol. 62, 2596 (1988) and A. O. Fuller and P. G. Spear, Proc. Natl. Acad. Sci. U.S.A. 84, 5454 (1987)). However, it is possible that Fd79 blocks both the infectivity and cell fusion functions of gB, while Fd138-80 blocks only the infectivity function of gD, so virus 20 can still spread cell-to-cell.

The binding, neutralization and protection results all indicate that the humanized Fd79 and Fd138-80 antibodies have retained the binding activities and the biological properties of the murine monoclonal antibodies. The availability 25 of humanized antibodies with specificity for HSV gB and gD, inter alia, provides an opportunity for studies of the in vivo potency and immunogenicity of humanized antibodies in treating viral diseases. The recognition by Fd79 and Fd138-80 of type-common epitopes of gB and gD (J. Koga 30 et al., Virology 151, 385 (1986)) expands the therapeutic potential to herpes simplex virus type 2 as well as type 1.

The use of a combination of two or more humanized antibodies in therapy is important for reducing the development of antibody resistant strains. Combination therapy of 35 humanized antibodies with other antiviral agents such as acyclovir provides further opportunities to combat diseases when chemotherapeutic agents alone have not been effective. As Fd79 and Fd138-80 reduce the frequency of viral persistence in a murine ocular model (J. F. Metcalf et al., 40 Cur. Eye Res. 6, 173 (1987)), the humanized antibodies, typically together with other antiviral agents, are capable of reducing episodes of recurrent genital infection, an area where traditional anti-viral agents have not been effective (L. Corey et al., N. Engl. J. Med. 306, 1313 (1982)). 45 Incorporation of the human constant domains can also enhance effector functions, such as antibody-dependent cellular cytotoxicity, leading to greater therapeutic efficiency in human patients.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other HSV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid 55 sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 7

Design of genes for M195 humanized light and heavy chains
The p67 protein or CD33 antigen is found on the surface
of progenitors of myeloid cells and of the leukemic cells of
most cases of AML, but not on lymphoid cells or nonhematopoietic cells (see, Leucocyte Typing III, ed. by A. J. 65
McMichael, Oxford University Press, pp. 622–629 (1987),
which is incorporated herein by reference). Antibodies that

are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

58

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference).

Cloning of heavy chain and light chain cDNA

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For M195, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 34A and FIG. 34B. Construction and expression of chimeric antibody

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cyl segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl Acad. Sci. U.S.A. 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVg1-dhfr but contains the human genomic Ck segment and the gpt gene. Derivatives of the M195 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and Xbal sites (see, Oueen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric M195 antibody was shown to bind to U937 cells, which express the CD33 antigen, by flow cytometry (FIG. 35).

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of M195.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the M195 variable region. The model was used to determine the amino acids in the M195 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain M195 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of four categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the M195 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse M195 antibody was used: The amino acids in each category are shown in Table 7. Some amino acids may be in more than one category. The final sequences of the humanized M195 light and heavy chain variable domains are shown in FIG. 36A and FIG. 36B, compared with the Eu sequences.

TABLE 7

Category	Light Chain	Heavy Chain
1	24–38, 54–60, 93–101	31–35, 50–66, 99–105
2	10, 52, 67, 110	93, 95, 98, 106, 107, 108, 110
3 .		30, 67, 98, 106
4	40, 52, 74	27, 30, 48, 68, 98

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse M195 chains (FIG. 34A and FIG. 34B), generally utilizing codons 45 found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four 50 overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 37A and FIG. 37B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction 60 enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were 65 carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies

The humanized M195 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to U937 cells in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 35), showing that it recognizes the same CD33 antigen.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse M195 antibody (FIG. 38). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in *Fundamental Immunology* (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The mouse M195 had an affinity comparable to the published value (Tanimoto et al., op. cit.) and the humanized M195 antibody had an affinity the same as the mouse M195 to within experimental error.

Humanized M195 is useful to mediate antibody-dependent cellular cytotoxicity when human effector cells and human CD33-expressing cells are used. This is analogous to other humanized antibodies, such as reported by Junghans et al., Cancer Research 50, 1495 (1990), which is incorporated herein by reference.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CD33 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 8

Design of genes for CMV5 humanized light and heavy chains

Three neutralizing antibodies to the gH glycoprotein of human cytomegalovirus (CMV) are designated CMV5, CMV109 and CMV115.

Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVg1-dhfr (FIGS. 24A and 24B) using site-directed mutagenesis, such as after CH1 to produce Fab

fragments or after the hinge region to produce $(Fab')_2$ fragments.

Cloning of heavy chain and light chain cDNA

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain 5 reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoR 10 I and HindIII and cloned into the pUC18 vector for sequencing. For CMV5, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced 15 amino acid sequences are shown in FIG. 39A and FIG. 39B. Similarly, by using techniques, which are well-known in the art, cDNAs for the CMV109 and CMV115 antibodies may be obtained and their sequence determined.

Construction and expression of chimeric antibody

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cyl segment including part of 25 the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. U.S.A. 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVg1-dhfr but contains the human genomic Ck segment and the gpt 30 gene. Derivatives of the CMV5 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete 45 antibody were detected by ELISA. Purified chimeric CMV5 antibody was shown to bind to CMV-infected cells, which express the gH antigen, by immunostaining of CMV-infected human embryonic lung fibroblasts.

respective plasmid vectors between the cytomegalovirus 40 promoter and the partial introns of the constant regions.

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human anti- 55 body is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework 60 sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Wol was chosen to 65 provide the framework sequences for humanization of CMV5.

62

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the CMV5 variable region. The model was used to determine the amino acids in the CMV5 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain CMV5 variable regions, at each position the amino acid was chosen to be the same as in the Wol antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Wol amino acid was unusual for human antibodies at that position, whereas the CMV5 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Wol light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse CMV5 antibody was used. In addition, a position was in the fifth category if the Wol amino acid was highly unusual for human antibodies at that position, and the CMV5 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 8. Some amino acids may be in more than one category. The final sequences of the humanized CMV5 light and heavy chain variable domains are shown in FIG. 40A and FIG. 40B, compared with the Wol sequences.

TABLE 8

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31–35, 50–66, 99–108 69, 80
2		69, 80
3	49	30
4 5		24, 27, 28, 30, 97 5

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse CMV5 chains (FIG. 39A and FIG. 39B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 41A and FIG. 41B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction

enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells are selected for gpt expression. Clones are screened by 10 assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is 15 eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Humanized antibody was also produced by transient transfection. The heavy chain and light chain plasmids were 20 transfected into S194 cells (ATCC TIB 19) by the DEAE-dextran method (Queen et al., Mol. Cell. Biol. 4, 1043 (1984), which is incorporated herein by reference), and humanized CMV5 antibody was purified from the media supernatant as above. Antibody was quantitated by ELISA 25 assay for human Ig.

Properties of humanized antibodies

The humanized CMV5 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized CMV5 antibody was shown to bind about as well 30 as the mouse and chimeric antibodies to CMV antigen, by immunostaining of CMV-infected human embryonic lung (HEL) cells (ATCC CCL 137). HEL cells monolayers in 96-well plates were infected with CMV at 0.01 pfu/cell, incubated for 4 days, dried at 37° C. and stored wrapped at 35 4° C. 100 μl blotto (5% Carnation Instant Milk in PBS at pH 7.4) was added to each well and incubated at 37° C. for 30 min. The blotto was poured off and 75 µl of a series of 2-fold dilutions of mouse, chimeric and humanized CMV5 antibody was added to the wells. The plate was incubated 1 hr 40 at 37° C. and washed twice with blotto (each wash was left on for 10 min). Then 75 µl of diluted peroxidase (HRP) conjugated goat anti-mouse or anti-human IgG (Tago) was added to each well and incubated for 1 hr at 37° C. The plate was washed 2× with PBS and 150 µl of HRP substrate 45 solution was added to each well. Color was allowed to develop at room temperature. The plates were washed with water and air dried. The wells were examined under a microscope to determine the highest dilution of the antibodies that formed a colored precipitate on the CMV-infected 50 cells. For all three antibodies, 63 ng/ml was the least amount of antibody that produced a detectable precipitate, indicating that humanized CMV5 binds about as well as the mouse and chimeric antibodies.

To compare the affinities of mouse and humanized CMV5 in another way, a competition experiment was performed. Plates of CMV-infected HEL cells as above were incubated with blotto for 30 min at 37° C. The blotto was poured off and dilutions of mouse or humanized CMV5 were added to each well in 75 µl of PBS. Then 125 µl of radio-iodinated 60 mouse CMV5 (1 µCi/µg) in PBS, containing 28,000 cpm was added to each well and incubated at 37° C. for 2.5 hr. The plate was washed 5 times with PBS, and the contents of each well were solubilized with 200 µl of 2% SDS and counted. Increasing concentrations of mouse and humanized 65 CMV5 inhibited binding of the radiolabeled CMV5 about equally well (FIG. 42), so humanized CMV5 has approxi-

64

matley the same binding affinity as mouse CV5. An irrelevant antibody did not compete in this assay.

The ability of humanized CMV5 to neutralize CMV is compared to that of mouse CMV5. Mouse and humanized CMV5 are successively diluted by 2-fold in 100 µl of DME medium+2% FCS in wells of a 96-well plate. 100 µl of CMV, which has been diluted to contain 100 tissue culture infectious dose-50% (TCID50) units, are added to each well and incubated for 60 min at 37° C. Each well of antibody-virus mixture is added to a well of subconfluent HEL cells in a 96-well plate from which the medium has been removed. The cells are incubated for 5 days and cytopathic effect (CPE) is examined in each well under a microscope. The highest dilution of antibody that inhibits CPE by 90% is a measure of the neutralizing ability of the antibody. The humanized CMV5 antibody will neutralize CMV antibody approximately as well as the mouse CMV5 antibody.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CMV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 9

Design of genes for AF2 human-like light and heavy chains This example is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of γ-IFN, such as monoclonal antibody AF2. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody AF2 are included in FIG. 43A and FIG. 43B. Due to codon degeneracy and non-critical aminoacid substitutions, other DNA sequences can be readily substituted for those sequences.

Cloning of heavy chain and light chain cDNA

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For AF2, two gamma-2b specific and two kappa specific clones were sequenced. The two gamma-2b clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 43A and FIG. 43B.

Construction and expression of chimeric antibody

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. U.S.A. 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVg1-dhfr but contains the human genomic Cκ segment and the gpt gene. Derivatives of the AF2 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain

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reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, Queen et al., Proc. Natl. Acad. Sci. U.S.A. 36, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

65

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Chimeric AF2 antibody was shown to bind to human γ -IFN by ELISA.

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. U.S.A. 86, 20 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of AF2.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the AF2 variable region. The model was used to determine the amino acids in the AF2 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain AF2 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the AF2 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding 50 region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is 55 interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse AF2 antibody was used. In addition, a position was in the fifth category if the 60 Eu amino acid was highly unusual for human antibodies at that position, and the AF2 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 9. Some amino acids may be in more than one category. The final sequences of the humanized AF2 light and heavy chain

66

variable domains are shown in FIG. 44A and FIG. 44B, compared with the Eu sequences.

TABLE 9

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-106
2	48	93, 95, 98, 107, 108, 109, 111
3		30, 98, 107
4 5	48, 70 63	27, 28, 30, 98, 107

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, plus typical immunoglobulin signal sequences, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 45A and FIG. 45B) The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences are then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions are carried out under conditions wellknown in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones are screened by assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies

The humanized AF2 antibody is characterized in comparison to the murine and chimeric antibodies. The humanized antibody will bind to γ -IFN in an ELISA assay in a manner similar to the mouse and chimeric antibodies, showing that it recognizes γ -IFN.

To compare the binding affinities of mouse AF2 antibody and humanized AF2 antibody, a competitive ELISA assay is performed. An ELISA plate is coated with human recombinant γ -IFN by adding 100 μ l of a 500 ng/ml solution of γ -IFN in PBS to each well and incubating overnight at 4° C. Subsequent steps are carried out at room temperature. The γ -IFN solution is removed and 200 μ l of ELISA buffer (0.1% Tween-20, 1% Bovine serum albumin in PBS) is added to each well and incubated for 1 hr. After removing the solution, varying amounts of competitor antibody (mouse AF2 or humanized AF2) in 100 μ l PBS is added to each well, along with an amount of biotinylated AF2 predetermined to give a good ELISA response. The plate is incubated for 1 hr

and then washed 3 times with ELISA buffer. An amount of horseradish peroxidase (HRP)-conjugated strepavidin predetermined to be in excess is added in 100 μl PBS to each well and incubated for 30 min. The plate is washed 3 times in ELISA buffer, and 100 μl of substrate solution for HRP is 5 added to each well. The plate is incubated for 10–30 min, and the optical density of each well is determined with an ELISA reader (BioRad). The decrease in optical density with increasing concentrations of competitor antibodies mouse AF2 and humanized AF2 are plotted. Mouse AF2 and 10 humanized AF2 will compete similarly, showing that their binding affinities for γ -IFN are approximately the same. The procedures used are well known in the art (e.g., Harlow and Lane, op. cit.).

An important biological activity of γ -IFN is the induction 15 of expression of class II HLA antigens on cells. To determine the ability of mouse and humanized AF2 to neutralize this activity, about 5×10^4 HS294T cells (Basham et al., J. Immunol. 130, 1492 (1983), which is incorporated herein by reference) are plated in 1.0 ml DMEM medium+10% FCS 20 in each well of a 24-well plate. After overnight incubation, 0.1 nM interferon and varying amounts of mouse or humanized AF2 are added to the cells, and the plate is incubated for 72 hr. The cells are removed from the plate with 0.05M EDTA, stained with monoclonal antibody L243 from the 25 American Type Culture Collection (ATCC) against HLA-D

antigen, washed, stained with FITC conjugated goat antimouse Ig and analyzed with a FACScan (Becton-Dickinson). Increasing concentrations of mouse AF2 reduce fluorescence of the cells (FIG. 46), indicating the antibody is preventing induction of HLA-D by γ -IFN. The humanized AF2 will act similarly to mouse AF2 in this assay, showing that it neutralizes the biological activity of γ -IFN.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other $\gamma\textsc{-}\textsc{IFN}$ specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

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( 1 ) GENERAL INFORMATION:
     ( i i i ) NUMBER OF SEQUENCES: 113
(2) INFORMATION FOR SEO ID NO:1:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 106 amino acids
                (B) TYPE: amino acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: unknown
      ( i i ) MOLECULE TYPE: protein
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: Protein
                ( B ) LOCATION: 1..106
                ( D ) OTHER INFORMATION: /note= "Variable region of the mouse
                        anti-Tac antibody light chain."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:1:
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85 90
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Phe Gly Ser Gly Thr Lys Leu Glu Leu Lys

-continued

1 0 0

(2) INFORMATION FOR SEQ ID NO:2:

- ($\,^{\mathrm{i}}$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER INFORMATION: /note= "Variable region of the human Eu antibody light chain."

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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 Ile
 Gin
 Met
 Thr
 Gln
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 Pro
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 Thr
 Ile
 Thr
 Cys
 Arg
 Ala
 Ser
 Ile
 Asn
 Thr
 Trp

 Leu
 Ala
 Tyr
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 Ile
 Ile

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..116
 - (D) OTHER INFORMATION: /note= "Variable region of the mouse anti-Tac antibody heavy chain."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Gln 1	Val	Gln	Lcu	G 1 n 5	Gln	Scr	G 1 y	Ala	G l u 10	Lcu	Ala	Lys	Pro	G l y 15	Ala
Scr	V a l	Lys			C y s				Gly		Thr	Phc	Thr 30	Ser	Тут
Arg	Мει			V a l	Lys		A r g 4 0		Gly	Gln	G 1 y	L c u 4 5	Glu	Тгр	I l e
Gly	Туг 50				Ser						T y r 6 0	A s n	Gln	L y s	Phe
L y s 6 5	A s p	L y s	Ala	Thr	L c u 70	Thr	Ala	A s p	Lys	S c r	Scr	Ser	Thr	Ala	Туг 80

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-continued
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90 95
                              Scr Leu Thr
                        Gly
                              Gly Val Phc
                                                     Tyr Trp Gly Gln Gly Thr Thr Lcu
                         100
             Val
                  Scr
                        Scr
                   1 1 5
( 2 ) INFORMATION FOR SEQ ID NO:4:
       ( i ) SEQUENCE CHARACTERISTICS:
              ( A ) LENGTH: 117 amino acids
              ( B ) TYPE: amino acid
              ( C ) STRANDEDNESS: single
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- (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..117
 - (D) OTHER INFORMATION: /note= "Variable region of the human Eu antibody heavy chain."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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Ser	V a l	Lys	V a 1 2 0	Scr	Суs	Lys	Ala	S c r 2 5	Gly	G 1 y	Thr	Phc	S c r 3 0	Агд	Scr
Ala	I 1 c	I 1 c 3 5	Тгр	Val	Arg	Gln	A 1 a 4 0	Pro	G 1 y	Gln	G 1 y	L c u 4 5	G1 u	Тrр	Mct
Gly	G 1 y 5 0	Ilc	Val	Pro	Mct	Phc 55	G 1 y	Pro	Pro	Аsп	T y r 6 0	Ala	G l n	Lys	Phe
G 1 n 6 5	G 1 y	Arg	V a 1	Thr	I 1 c 7 0	Thr	Ala	A s p	Glu	S c r 7 5	Thr	A s n	Thr	Ala	T y r 8 0
Met	G 1 u	Lcu	Scr	S c r 8 5	Lcu	Arg	Ser	Glu	A s p 9 0	Thr	Ala	Phe	Туг	P h c 9 5	Суs
Ala	Gly	Gly	T y r 1 0 0	Gly	I 1 c	Туг	Ser	Pro 105	G 1 u	Glu	Туг	A s n	G 1 y 1 1 0	G 1 y	Leu
Val	Thr	V a 1 1 1 5	Ѕсг	Scr											

(2) INFORMATION FOR SEQ ID NO:5:

- ($\,i\,$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..116
 - (D) OTHER INFORMATION: /note= "Variable region of the PDL humanized anti-Tac antibody heavy chain."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:5:
 - Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ser

-continued

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      Arg
      Met
      His
      Trp
      Val
      Arg
      Gln
      Ala
      Pro
      Gly
      Gln
      Gly
      Leu
      Glu
      Trp
      Ile

      Gly
      Tyr
      Ile
      Asn
      Pro
      Ser
      Thr
      Gly
      Tyr
      Thr
      Glu
      Tyr
      Asn
      Gln
      Lys
      Phe

      Lys
      Asp
      Lys
      Ala
      Thr
      Ile
      Thr
      Ala
      Asp
      Glu
      Ser
      Thr
      Asn
      Thr
      Ala
      Tyr

      Met
      Glu
      Leu
      Ser
      Leu
      Arg
      Ser
      Glu
      Asp
      Thr
      Ala
      Val
      Tyr
      Tyr
      Tyr
      Tyr
      Tyr
      Tyr
      Tyr
      Tyr
      Cys

      Ala
      Arg
      Gly
      Gly
      Val
      Phe
      Asp
      Tyr
      Trp
      Gly
      Gln
      Gly
      Thr
      Leu
      Val

      Thr
      Val
      Ser
      Ser
      Leu
      Asp
      Tyr
      Trp
      Gly
      Gln
      Gly
      Thr
      Leu
      Val
      Leu
      Val
      Leu
      V
```

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..116
 - (D) OTHER INFORMATION: /note= "Variable region of the CDR-only humanized anti-Tac antibody heavy chain."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:6:

G 1 n 1	V a l	Gln	Leu	Val 5	Gln	Scr	Gly	Ala	Glu 10	V a 1	Lys	Lys	Pго	G 1 y 1 5	Ser
Sег	Val	Lys	V a 1 2 0	Ser	C y s	Lys	Ala	S e r 2 5	Gly	Gly	Thr	Phe	S c r 3 0	Scr	Туг
Агд	Met	H i s	Тrр	Val	Arg	Gln	A 1 a 4 0	Pro	Gly	Gln	Gly	L e u 4 5	Glu	Тгр	Мсі
Gly	Туг 50	Ilc	Asn	Pro	Scr	T h r 5 5	Gly	Туr	Thr	Glu	T y r 6 0	Asn	Gln	L y s	Phe
L y s 6 5	A s p	Arg	V a l	Thr	I 1 c 7 0	Thr	Ala	Asp	Giu	S с г 7 5	Thr	Аsп	Thr	Ala	Туг 80
Мει	Glu	Leu	Ser	S с т 8 5	Leu	Агд	Scr	Glu	A s p 9 0	Thr	Ala	Phc	Туг	P h c 9 5	C y s
Ala	Gly	Gly	G 1 y 1 0 0	Gly	Val	Phc	Asp	T y r 1 0 5	Glu	Туг	A s n	Gly	G 1 y 1 1 0	Leu	Val
Thr	V a l	S c r 1 1 5	Scr												

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..106

(D) OTHER INFORMATION: /notc= "Variable region of the PDL humanized anti-Tac antibody light chain."

(\times i) SEQUENCE DESCRIPTION: SEQ ID NO:7:

 Asp 11c
 G1n
 Mc1 5 5
 G1n
 Scr Pro Scr Thr 10
 Lcu Scr Scr Ala Scr Val Scr Val 15
 Gly 15

 Asp Arg Val Thr 20
 Ile Thr Cys Scr Ala Scr Scr Ala Scr Scr Scr Ile Scr Tyr Mct 25

 His Trp Tyr Gln Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr 35

 Thr 50
 Scr Asn Leu Ala Scr Gly Val Pro Ala Arg Phe 65

 Gly Scr Gly Thr Glu Phe 70
 Thr Leu Thr Ile Scr Scr Scr Leu Gln Pro Asp 80

 Asp Phe Ala Thr Tyr Tyr Cys His Gln Arg 90
 Scr Thr Tyr Pro Leu Thr Phe 90

 Gly Gly Gln Gly Thr Lys Val Glu Val Lys
 Lys

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..106
 - (D) OTHER INFORMATION: /notc= "Variable region of the CDR-only humanized anti-Tac antibody light chain."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:8:

 Asp 11e
 Gln Met Thr Gln Ser Pro Ser Thr Leu Ser Ala Ser Val Gly

 Asp Arg Val Thr 20
 Thr Cys Ser Ala Ser Ser Ser Ser Ile Ser Tyr Met 30

 His Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Met Tyr 35

 Thr Thr Ser Asn Leu Ala Ser Gly Val Pro Ser Arg Phe Ile Gly Ser 50

 Gly Ser Gly Thr Glu Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Asp 65

 Asp Phe Ala Thr Tyr Tyr Cys His Gln Arg Ser Thr Tyr Pro Leu Thr 105

 Phe Gly Gln Gly Thr Lys Val Glu Val Lys 106

($\,2\,$) Information for SEQ id no:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 443 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA
- (i i i) HYPOTHETICAL: NO

(ix)FEATURE:

(A) NAME/KEY: misc_feature

(B) LOCATION: 1..443

(D) OTHER INFORMATION: /note= "Sequence encoding heavy chain variable region of CDR-only humanized anti-Tac antibody including signal sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGCTTCTAGA TGGGATGGAG CTGGATCTTT CTCTTCCTCC TGTCAGGTAC CGCGGGCGTG 6.0 CACTCTCAGG TCCAGCTTGT CCAGTCTGGG GCTGAAGTCA AGAAACCTGG CTCGAGCGTG 120 AAGGTCTCCT GCAAGGCTTC TGGCGGGACC TTTTCTAGCT ACAGGATGCA CTGGGTAAGG 180 CAGGCCCCTG GACAGGGTCT GGAATGGATG GGATATATTA ATCCGTCGAC TGGGTATACT 2 4 0 GAATACAATC AGAAGTTCAA GGACAGGGTC ACAATTACTG CAGACGAATC CACCAATACA 3 0 0 GCCTACATGG AACTGAGCAG CCTGAGATCT GAGGACACCG CATTCTATTT CTGTGCAGGG 3 6 0 GGTGGGGGAG TCTTTGACTA CGAATACAAT GGAGGGCTGG TCACAGTCTC CTCAGGTGAG 4 2 0 TCCTTAAAAC CTCTAGACGA TAT 4 4 3

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 411 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..411
- (D) OTHER INFORMATION: /notc= "Sequence encoding light chain variable region of the CDR-only humanized anti-Tac antibody including signal sequence."

($\,x\,$ i) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CAAATCTAGA TGGAGACCGA TACCCTCCTG CTATGGGTCC TCCTGCTATG GGTCCCAGGA 60
TCAACCGGAG ATATTCAGAT GACCCAGTCT CCATCTACCC TCTCTGCTAG CGTCGGGGAT 120
AGGGTCACCA TAACCTGCTC TGCCAGCTCA AGTATAAGTT ACATGCACTG GTACCAGCAG 180
AAGCCAGGCA AAGCTCCCAA GCTTCTAATG TATACCACAT CCAACCTGGC TTCTGGAGTC 240
CCTTCTCGCT TCATTGGCAG TGGATCTGGG ACCGAGTTCA CCCTCACAAT CAGCTCTCTG 300
CAGCCAGATG ATTTCGCCAC TTATTACTGC CATCAAAGGA GTACTTACCC ACTCACGTTC 360
GGTCAGGGGA CCAAGGTGGA GGTCAAACGT AAGTACACTT TTCTAGATAT A 411

($\,2\,$) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1,.29
- (D) OTHER INFORMATION: /standard_namc= "Primer mc045"

```
( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:11:
TAATCTAGAA TTCCCCCCC CCCCCCCC
                                                                                                                29
( 2 ) INFORMATION FOR SEQ ID NO:12:
        ( \,\mathrm{i}\, ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 46 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_feature
                ( B ) LOCATION: 1..46
                ( D ) OTHER INFORMATION: /standard_name= "Primer mc045"
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:12:
TATAGAGCTC AAGCTTGGAT GGTGGGAAGA TGGATACAGT TGGTGC
( 2 ) INFORMATION FOR SEQ ID NO:13:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 50 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_feature
                ( B ) LOCATION: 1..50
                ( D ) OTHER INFORMATION: /standard_namc= "Primer mc047"
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:13:
TATAGAGCTC AAGCTTCCAG TGGATAGACH GATGGGGSTG TYGTTTTGGC
                                                                                                                50
( 2 ) INFORMATION FOR SEQ ID NO:14:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 116 amino acids
                ( B ) TYPE: amino acid
                (C) STRANDEDNESS: single
                ( D ) TOPOLOGY: unknown
      ( i i ) MOLECULE TYPE: protein
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: Protein
                ( B ) LOCATION: 1..116
                ( D ) OTHER INFORMATION: /notc= "Anti-Tac heavy chain amino
                        acid sequence."
      ( x i ) SEQUENCE DESCRIPTION: SEO ID NO:14:
       Gln Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Ala Lys Pro Gly Ala
       Ser Val Lys Met Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr
       Arg Met His Trp Val Lys Gln Arg Pro Gly Gln Gly
                                                                                   Leu Glu Trp Ile
                                                    4 0
            Tyr Ile Asn Pro Ser Thr Gly Tyr Thr Glu Tyr Asn Gln Lys Phe
50 60
```

-continued

Lys Asp Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr 70

Met Gln Leu Ser Ser Leu Thr Phe Glu Asp Ser Ala Val Tyr Tyr Cys 95

Ala Arg Gly Gly Gly Val Phe Asp Tyr Trp Gly Gln Gly Thr Thr Leu 115

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..117
 - (D) OTHER INFORMATION: /note= "Eu heavy chain amino acid sequence."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Gin Val Gin Leu Val Gin Ceu Val Gin Ser Giy Ala Giu Val Lys Lys Pro Giy Ser Ser Val Lys Val Lys Val Ser Cys Lys Ala Ser Giy Giy Giy Thr Phe Ser Arg Ser Ala Ile Ile Trp Val Arg Gin Ala Pro Giy Giy Gin Giy Leu Giu Trp Met 50 Asa Sor Arg Ser Giy Giy Gin Giy Leu Giu Trp Met 50 Asa Sor Arg Ser Arg Ser Giy Giy Gir Ala Gir Lys Phe 65 Asa Sor Arg Gir Arg Met Asa Sor Gir Arg Gir Ala Thr Ala Tyr Asa Thr Ala Tyr Asa Gir Arg Ser Arg Ser Giu Asp Thr Ala Phe Tyr Phe Cys Sor Ala Gir Trr Nala Tyr Asa Gir Trr Nala Tyr Ala Thr Val Ser Ser Ser

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS; single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..106
 - (D) OTHER INFORMATION: /note= "Anti-Tac light chain amino acid sequence."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:16:
- Gln Ilc Val Lcu Thr Gln Scr Pro Ala Ilc Mct Scr Ala Scr Pro Gly

1				5					1 0					1 5	
Glu	Lys	Val	Thr 20	Ilc	Thr	Суs	Ser	A l a 2 5	Scr	Scr	S c r	I 1 c	S c r 3 0	Туг	Меι
H i s	Trp	P h c 3 5	Gln	Gln	Lys	Pro	G 1 y 4 0	Thr	Scr	Pro	Lys	L c u 4 5	Trp	I 1 c	Туг
Thr	Thr 50	Scr	Asn	Leu	Ala	S c r 5 5	G 1 y	Val	Pro	Ala	A r g	Phe	S e r	G 1 y	Scr
G 1 y 6 5	S c r	Gly	Thr	Scr	Туг 70	Scr	Lcu	Thr	Ilc	S c r 7 5	Arg	Мει	Glu	Ala	Glu 80
A s p	Ala	Ala	Thr	T y r 8 5	Туг	C y s	H i s	Gln	Arg 90	Scr	Thr	Туг	Pro	L c u 9 5	Thr
Phe	Gly	S c r	G 1 y 1 0 0	Thr	Lys	Lcu	G 1 u	L c u 1 0 5	Lys						

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER INFORMATION: /notc= "Eu light chain amino acid sequence."

($\,\mathbf{x}\,$ i $\,$) SEQUENCE DESCRIPTION: SEQ ID NO:17:

```
      Asp 11c
      G1n
      Mci Thr 5
      G1n
      Scr Pro Scr Thr 10
      Leu Scr A1a Scr Val G1y 15
      G1y Scr Val G1y 15

      Asp Arg Val Thr 20
      Thr G1n
      Cys Arg A1a Scr G1n Scr G1n Scr G1n Scr I1c Asn Thr Trp 30
      Trp Trp 30
      Leu A1a Trp Tyr G1n G1n Lys Pro G1y Lys A1a Pro Lys Leu Leu Mci 40

      Tyr Lys Sor Scr G1y Thr G1n G1n Scr Scr Leu G1u Scr G1y Val Pro G6
      Scr Arg Phc I1c G1y 55

      Scr G1y Scr G1y Thr G1u Phc Thr Leu Thr I1c Scr Scr Scr Leu G1n Pro 65

      Asp Asp Phc A1a Thr Tyr Tyr Cys G1n G1n Tyr Asn Scr Asp Scr Lys 95

      Mci Phc G1y G1n G1y Thr Lys Val G1u Val Lys
```

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 433 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA
- (i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 6..410
 - (D) OTHER INFORMATION: /product= "Humanized anti-Tac heavy chain variable region, Seq ID. 19"

85 86

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:18: TCTAG ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGT ACC Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Ser Gly Thr 4 7 GCG GGC GTG CAC TCT CAG GTC CAG CTT GTC CAG TCT GGG GCT GAA GTC Ala Gly Val His Scr Gln Val Gln Lcu Val Gln Scr Gly Ala Glu Val 9 5 AAG AAA CCT GGC TCG AGC GTG AAG GTC TCC TGC AAG GCT TCT GGC TAC Lys Lys Pro Gly Scr Scr Val Lys Val Scr Cys Lys Ala Scr Gly Tyr 1 4 3 ACC TTT ACT AGC TAC AGG ATG CAC TGG GTA AGG CAG GCC CCT GGA CAG
Thr Phc Thr Scr Tyr Arg Mei His Trp Val Arg Gln Ala Pro Gly Gln
50 55 191 GGT CTG GAA TGG ATT GGA TAT ATT AAT CCG TCG ACT GGG TAT ACT GAA Gly Leu Glu Trp Ile Gly Tyr Ile Asn Pro Ser Thr Gly Tyr Thr Glu 2 3 9 TAC AAT CAG AAG TTC AAG GAC AAG GCA ACA ATT ACT GCA GAC GAA TCC
Tyr Asn Gln Lys Phc Lys Asp Lys Ala Thr Ilc Thr Ala Asp Glu Scr 287 ACC AAT ACA GCC TAC ATG GAA CTG AGC AGC CTG AGA TCT GAG GAC ACC 3 3 5 Thr Asn Thr Ala Tyr Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr GCA GTC TAT TAC TGT GCA AGA GGG GGG GGG GTC TTT GAC TAC TGG GGC Ala Val Tyr Tyr Cys Ala Arg Gly Gly Gly Val Phc Asp Tyr Trp Gly 125 3 8 3 CAA GGA ACC CTG GTC ACA GTC TCC TCA GGTGAGTCCT TAAAACCTCT Gln Gly Thr Leu Val Thr Val Ser Ser 4 3 0

4 3 3

(2) INFORMATION FOR SEO ID NO:19:

AGA

(i) SEOUENCE CHARACTERISTICS:

1 3 0

- (A) LENGTH: 135 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Met Gly Trp Ser Trp Ite Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly Val His Scr Gln Val Gln Leu Val Gln Scr Gly Ala Glu Val Lys Lys Pro Gly Scr Scr Val Lys Val Scr Cys Lys Ala Scr Gly Tyr Thr Phc Thr Scr Tyr Arg Met His Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp Ile Gly Tyr Ile Asn Pro Ser Thr Gly Tyr Thr Glu Tyr Asn Gln Lys Phc Lys Asp Lys Ala Thr Ilc Thr Ala Asp Glu Scr Thr Asn Thr Ala Tyr Mei Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val 100 105 1 2 0 Thr Leu Val Thr Val Ser Ser 1 3 0 1 3 5

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 403 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: CDS (B) LOCATION: 6..383
- (D) OTHER INFORMATION: /product= "Humanized anti-Tac light chain variable region: Seq ID. 21"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:20:

		TA TGG GTC CTC CTG C cu Trp Val Leu Leu L 10	
		ATG ACC CAG TCT CCA Met Thr Gln Ser Pro 25	
		ACC ATA ACC TGC TCT Thr llc Thr Cys Scr 40	
		CAG CAG AAG CCA GGC Gln Gln Lys Pro Gly 60	Lys Ala
		AAC CTG GCT TCT GGA Asn Leu Ala Ser Gly 75	
		ACC GAG TTC ACC CTC Thr Glu Phe Thr Leu 90	
		ACT TAT TAC TGC CAT Thr Tyr Tyr Cys His 105	
		GGG ACC AAG GTG GAG Gly Thr Lys Val Glu 120	
CGTAAGTACA CTTTTCTA	G A		4 0 3

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 126 amino acids (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Μ e ι	Glu	Thr	A s p	T h r 5	Lcu	Leu	Leu	Тrр	V a 1 1 0	L e u	L c u	Leu	Trp	V a 1 1 5	Pro
Gly	Scr	Thr	G 1 y 2 0	Asp	Ile	Gln	Μcι	T h r 2 5	Gln	Ser	Pro	Scr	T h r 3 0	Leu	Ser
Ala	Scr	V a 1 3 5	Gly	A s p	Arg	Val	T h r 4 0	Ilc	Thr	C y s	Ser	A l a 4 5	Ser	Scr	Scr
Ile	S c r 5 0	Туг	Мει	H i s	Тгр	T y r 5 5	Gln	Gln	Lys	Pro	G 1 y 6 0	L y s	Ala	Pro	L y s
Lcu	Leu	Ile	Туг	Thr	Thr	Scr	A s n	Lcu	Ala	Ser	Gly	V a l	Pro	Ala	Arg

-continued 6.5 7 0 7 5 8 0 Scr Gly Gly Ser Gly The Glu Phc Thr Lcu Thr Scr Туг H i s P r oThr Рhс Gly Gln Leu Gly Thr Lys Val Glu Val Lys 120 (2) INFORMATION FOR SEQ ID NO:22: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 126 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..126 (D) OTHER INFORMATION: /standard_name= "Oligo HES12" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac heavy chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:22: AGCTTCTAGA TGGGATGGAG CTGGATCTTT CTCTTCCTCC TGTCAGGTAC CGCGGGCGTG 6 0 CACTCTCAGG TCCAGCTTGT CCAGTCTGGG GCTGAAGTCA AGAAACCTGG CTCGAGCGTG 120 AAGGTC 1 2 6 (2) INFORMATION FOR SEQ ID NO:23: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 129 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..129 (D) OTHER INFORMATION: /standard_name= "Oligo HES13" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac heavy chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:23: CCCAGTCGAC GGATTAATAT ATCCAATCCA TTCCAGACCC TGTCCAGGGG CCTGCCTTAC 60 CCAGTGCATC CTGTAGCTAG TAAAGGTGTA GCCAGAAGCC TTGCAGGAGA CCTTCACGCT 120 CGAGCCAGG 129 (2) INFORMATION FOR SEQ ID NO:24: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 124 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO	
(i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1124 (D) OTHER INFORMATION: /standard_namc= "Oligo HES14" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac heavy chain gene."	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
TATATTAATC CGTCGACTGG GTATACTGAA TACAATCAGA AGTTCAAGGA CAAGGCAACA	6 0
ATTACTGCAG ACGAATCCAC CAATACAGCC TACATGGAAC TGAGCAGCCT GAGATCTGAG	120
GACA	120
	124
(2) INFORMATION FOR SEQ ID NO:25:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(i i) MOLECULE TYPE: DNA	
(i i i) HYPOTHETICAL: NO	
(i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1128 (D) OTHER INFORMATION: /standard_name= "Oligo HES15" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac heavy chain gene."	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
ATATCGTCTA GAGGTTTTAA GGACTCACCT GAGGAGACTG TGACCAGGGT TCCTTGGCCC	6 0
CAGTAGTCAA AGACCCCCCC CCCTCTTGCA CAGTAATAGA CTGCGGTGTC CTCAGATCTC	1 2 0
AGGCTGCT	1 2 8
(2) INFORMATION FOR SEQ ID NO:26:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(i i) MOLECULE TYPE: DNA	
(i i i) HYPOTHETICAL: NO	
(i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1120 (D) OTHER INFORMATION: /standard_name= "Oligo JFD1" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac light chain gene."	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
CAAATCTAGA TGGAGACCGA TACCCTCCTG CTATGGGTCC TCCTGCTATG GGTCCCAGGA	6 0
TCAACCGGAG ATATTCAGAT GACCCAGTCT CCATCTACCC TCTCTGCTAG CGTCGGGGAT	1 2 0
(2) INFORMATION FOR SEQ ID NO:27:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 base pairs (B) TYPE: nucleic acid	

-continued (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..114 (D) OTHER INFORMATION: /standard_name= "Oligo JFD2" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac light chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:27: ATAAATTAGA AGCTTGGGAG CTTTGCCTGG CTTCTGCTGG TACCAGTGCA TGTAACTTAT 6 0 ACTTGAGCTG GCAGAGCAGG TTATGGTGAC CCTATCCCCG ACGCTAGCAG AGAG 1 1 4

(2) INFORMATION FOR SEQ ID NO:28:					
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 123 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear					
(i i) MOLECULE TYPE: DNA					
(i i i) HYPOTHETICAL: NO					
(i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1123 (D) OTHER INFORMATION: / / now="One of four of synthesize the humaning gene."	standard_namc= "O	i to			
(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID	NO:28:				
GCTCCCAAGC TTCTAATTTA TA	CCACATCC	AACCTGGCTT	CTGGAGTCCC	TGCTCGCTTC	6 0
AGTGGCAGTG GATCTGGGAC CG.	AGTTCACC	CTCACAATCA	GCTCTCTGCA	GCCAGATGAT	1 2 0
TTC					1 2 3
(2) INFORMATION FOR SEQ ID NO:29:					
(i) SEQUENCE CHARACTERISTICS:					
(i i) MOLECULE TYPE: DNA					
(i i i) HYPOTHETICAL: NO					
(i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1122 (D) OTHER INFORMATION: / note="One of four synthesize the humaningene."	/standard_namc= "Coligonucleotides use	d to			
(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID	NO:29:				
TATATCTAGA AAAGTGTACT TA	CGTTTGAC	CTCCACCTTG	GTCCCCTGAC	CGAACGTGAG	6 0
TGGGTAAGTA CTCCTTTGAT GG	САСТААТА	AGTGGCGAAA	TCATCTGGCT	GCAGAGAGCT	1 2 0
G A					1 2 2

								-cc	ntinue	d						
(2)	NFORM	ATION FO	OR SEQ I	D NO:30:		•										
	(i	(NCE CHA A) LENG B) TYPE C) STRA D) TOPG	GTH: 384 E: nucleic ANDEDN	basc pair: acid ESS: singl											
	(ii) MOLEC	CULE TY	PE: cDNA	.											
	(iii) НҮРОТ	HETICAL	L: NO												
	(i x	(A) NAM B) LOC D) OTHI	ATION: 1. ER INFOI	.384	-	_	chain varia	able							
	(x i) SEQUE	NCE DES	CRIPTIO	N: SEQ I	D NO:30:										
ATG Mci	GAT Asp	TTT Phc	C A A G l n	GTG Val 5	CAG Gln	ATT	TTC Phc	AGC Scr	TTC Phc 10	CTG Leu	CTA Leu	ATC llc	AGT Ser	G C C A 1 a 1 5	TCA Ser	4 8
GTC Val	ATA	C T G L c u	T C C S c r 2 0		GGA Gly									G C A A 1 a		9 6
ATG Μει	TCT Scr	GCG Ala 35	TCT Scr	C C A P r o	GGG Gly	GAG Glu	AAG Lys 40	GTC Val	ACC	ATG Met	ACC Thr	T G C C y s 4 5		GGC Gly		1 4 4
T C A S c r	AGT Scr 50	GTA Val	AGT Scr	TTC Phc	ATG Mct	T A C T y r 5 5								T C C S c r		192
CCC Pro 65	AGA Агg	CTC Lcu	C T G L e u	ATT	TAT Tyr 70		A C A T h r							GTC Val	C C T P r o 8 0	2 4 0
GTT Val		TTC Phe				GGG G1y								A C A T h r 9 5		288
AGC Scr	C G A A r g	ATG Met	G A G G 1 u 1 0 0	GCT Ala	GAA Glu	GAT Asp	G C T A 1 a	G C C A 1 a 1 0 5	ACT Thr	ТАТ	TAC Tyr	T G C C y s	C A G G 1 n 1 1 0	CAG Gln	TGG Trp	3 3 6
AGT Scr	ACT	T A C T y r 1 1 5	C C G P r o											CTG Lcu		3 8 4
(2)I	NFORM/	TION FO	R SEQ II	NO:31:												
	(i	()	NCE CHA A) LENC B) TYPE D) TOPC	FTH: 128 : amino a	amino aci cid	ds										
	(i i) MOLEC	ULE TYP	E: proteii	ı											
	(x i) SEQUE	NCE DES	CRIPTIO	N: SEQ II	O NO:31:										
M e t		Phc		5					1 0					1 5		
Val Mei		Leu Ala	2 0					2 5					3 0			
		3 5			- 1 3	5.4	40	, 4 1		171 U L	4 41 1	4 5	JUI	СГУ	o c r	

WIC I	3 6 1				Gly		L y s 4 0					C y s 4 5	Ser	Gly	Scr
Scr	S c r 5 0	V a 1	Ser	Phc	Met	T y r 5 5	Trp	Туr	Gln	Gln	Arg 60	Pro	G 1 y	Scr	Ser
Pro 65	Arg	Lcu	Leu	Ιlε	Туг . 70	A s p	Thr	Scr	A s n	L c u 7 5	Ala	Scr	Gly	V a 1	Pro 80
Val	Агд	Phc	Scr	Gly	S c r	G 1 y	S c r	Gly	Thr	Ser	Туг	Scr	Lcu	Thr	Ilc

		 	-
			-
		_	

Scr Arg Mcı Glu Ala Glu Asp Ala Ala Thr Tyr Tyr Cys Gln Gln Trp 100 Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu Lys 120

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 414 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..414
 - (D) OTHER INFORMATION: /product="Heavy chain var. region of the antibody mik-beta1: SeqID 33"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:32:

		CTG Lcu						4 8
		CAG Gln						9 6
		T C C S c r						1 4 4
		CAC His						1 9 2
		ATA I1c 70		 	 _			2 4 0
		CTG Lcu						2 8 8
		AAC Asn						3 3 6
		GGG Gly						3 8 4
		GTC Val						4 1 4

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH; 138 amino acids

 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION; SEQ ID NO:33:

Met Ala Val Leu Gly Leu Leu Phe Cys Leu Val Thr Phe Pro Ser Cys 1 0

```
        Val
        Leu
        Ser
        Gln
        Val
        Gln
        Leu
        Lys
        Gln
        Ser
        Gly
        Pro
        Gly
        Leu
        Val
        Gln

        Pro
        Ser
        Gln
        Ser
        Leu
        Ser
        Ile
        Thr
        Cys
        Thr
        Val
        Ser
        Gly
        Phe
        Ser
        Val

        Thr
        Ser
        Tyr
        Gly
        Val
        His
        Trp
        Ile
        Arg
        Gln
        Ser
        Pro
        Gly
        Lys
        Gly
        Leu

        Glu
        Trp
        Leu
        Gly
        Val
        Ile
        Trp
        Ser
        Gly
        Gly
        Ser
        Thr
        Asp
        Tyr
        Asp
        Tyr
        Asp
        Tyr
        Asp
        Asp
        Tyr
        Asp
        Thr
        Asp
        Thr
        Ile
        Tyr
        Asp
        Gly
        Asp
        Tyr
        Asp
        Tyr
        Asp
        Gly
        Phe
        Asp
        Tyr
        Tyr
        Asp
        Gly
        Phe
        Asp
        Ala
        Tyr
        Asp
```

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER INFORMATION: /note= "Amino acid sequence of the light chain for humane Lay antibody."

($\,x\,$ i $\,$) SEQUENCE DESCRIPTION: SEQ ID NO:34:

 Asp 11c
 G1n
 Mct
 Thr 5
 G1n
 Scr Pro Scr Scr Leu Scr Leu Scr Val Scr Val Gly 15
 Gly 15

 Asp Arg Val Thr 20
 11c Thr Cys G1n Ala Scr G1n Ala Scr G1n Asn Val Asn Ala Tyr 20
 Asn Trp 30
 Ala Tyr 30
 Ala T

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:

-continued

- (A) NAME/KEY: Protein
- (B) LOCATION: 1..106
- (D) OTHER INFORMATION: /note= "Amino acid sequence of the light chain of the humanized mik-beta1 antibody."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Tyr Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu 11c Tyr 35 Gly Scr Gly Thr Asp Tyr Thr Phc Thr Ilc Scr Scr Leu Gln Pro Glu 65 75 80 Asp Ilc Ala Thr Tyr Tyr Cys Gln Gln Trp Scr Thr Tyr Pro Leu Thr 85 90 95

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1 122
 - (D) OTHER INFORMATION: /note= "Amino acid sequence of the heavy chain of the human Lay antibody."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Ala Met Ser Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 35 Asn Gly Arg Phe Thr Ile Scr Arg Asn Asp Scr Lys Asn Thr Leu Tyr 65 70 80 Leu Gln Mei Asn Gly Leu Gln Ala Glx Val Ser Ala Ile Tyr Tyr Cys 85Ala Arg Asp Ala Gly Pro Tyr Val Scr Pro Thr Phc Phc Ala His Trp 100

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single

(A) NAME/KEY: misc_fcature (B) LOCATION: 1..136

(D) OTHER INFORMATION: /standard_name= "Oligo vc14"

-continued

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( D ) TOPOLOGY: unknown
     ( i i ) MOLECULE TYPE: protein
    ( i i i ) HYPOTHETICAL: NO
     ( i x ) FEATURE:
              ( A ) NAME/KEY: Protein
              ( B ) LOCATION: 1..119
              ( D ) OTHER INFORMATION: /notc= "Amino acid sequence of the
                    heavy chain of the humanized mik-beta1 antibody."
     ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:37:
      Gly Val Ilc Trp Scr Gly Gly Scr Thr Asp Tyr Asn Ala Ala Phc Ilc
50 55
                      Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu
70 75 80
      Gln Met Asn Ser Leu Gln Ala Glu Asp Thr Ala Ile Tyr Tyr Cys Ala
85 90 95
       Arg Ala Gly Asp Tyr Asn Tyr Asp Gly Phc Ala Tyr Trp Gly Gln Gly 100
       Thr Leu Val Thr Val Scr Scr
                 115
( 2 ) INFORMATION FOR SEQ ID NO:38:
       ( i ) SEQUENCE CHARACTERISTICS:
              ( A ) LENGTH: 107 base pairs
              ( B ) TYPE: nucleic acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: linear
     ( i i ) MOLECULE TYPE: DNA
    ( i i i ) HYPOTHETICAL: NO
     ( i x ) FEATURE:
              ( A ) NAME/KEY: misc_feature
              ( B ) LOCATION: 1..107
              ( D ) OTHER INFORMATION: /standard_namc= "Oligo vc13"
     ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:38:
TTCTGCTGGT ACCAGTACAT GAAACTTACA CTTGAGCTGC CACTGCAGGT GATGGTGACG
                                                                                                60
CGGTCACCCA CTGAGGCACT GAGGCTAGAT GGAGACTGGG TCATTTG
                                                                                               107
( 2 ) INFORMATION FOR SEQ ID NO:39:
       ( i ) SEQUENCE CHARACTERISTICS:
              ( A ) LENGTH: 136 base pairs
              ( B ) TYPE: nucleic acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: linear
     ( i i ) MOLECULE TYPE: DNA
    ( i i i ) HYPOTHETICAL: NO
     ( i x ) FEATURE:
```

-continued (x i) SEQUENCE DESCRIPTION: SEQ ID NO:39: CATGTACTGG TACCAGCAGA AGCCAGGAAA AGCTCCGAAA CTTCTGATTT ATGACACATC 60 CAACCTGGCT TCTGGAGTCC CTTCCCGCTT CAGTGGCAGT GGGTCTGGGA CCGATTACAC 120 CTTTACAATC TCTTCA 1 3 6 (2) INFORMATION FOR SEQ ID NO:40: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 137 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..137 (D) OTHER INFORMATION: /standard_name= "Oligo vc15" (x i) SEQUENCE DESCRIPTION: SEQ ID NO:40: TGTGTCTAGA AAAGTGTACT TACGTTTTAC CTCGACCTTG GTCCCTTGAC CGAACGTGAG CGGGTAAGTA CTCCACTGCT GGCAGTAATA AGTGGCTATA TCTTCCGGCT GAAGTGAAGA 1 2 0 GATTGTAAAG GTGTAAT 1 3 7 (2) INFORMATION FOR SEQ ID NO:41: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 108 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..108 (D) OTHER INFORMATION: /standard_name= "Oligo vc16" (x i) SEQUENCE DESCRIPTION: SEQ ID NO:41: CACATCTAGA CCACCATGGA TTTTCAAGTG CAGATCTTCA GCTTCCTGCT AATCAGTGCC 6.0 TCAGTCATAC TGTCCAGAGG AGATATTCAA ATGACCCAGT CTCCATCT 108 (2) INFORMATION FOR SEQ ID NO:42: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 138 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..138 (D) OTHER INFORMATION: /standard_name= "Oligo vc11" (x i) SEQUENCE DESCRIPTION: SEQ ID NO:42: TAGTCTGTCG ACCCACCACT CCATATCACT CCCACCCACT CGAGTCCCTT TCCAGGAGCC 6 0

TGGCGGACCC	AGTGTACACC	ATAACTTGTT	ACGGTGAAAC	CACTGGCGGC	ACAAGACAGT	1 2 0
C T C A G A G A T C	CTCCTGGC					1 3 8
(2) INFORMATION	FOR SEQ ID NO:43:					
	JENCE CHARACTERIST (A) LENGTH: 126 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs single				
(ii) MOL	ECULE TYPE: DNA					
(iii) HYP	OTHETICAL: NO					
	TURE: (A) NAME/KEY: misc_ (B) LOCATION: 1126 (D) OTHER INFORMA		Oligo vc12"			
(xi)SEQ	JENCE DESCRIPTION: S	EQ ID NO:43:				
тостосстсс	ACAGACTATA	ATGCAGCTTT	C A T A T C C A G A	T T T A C C A T C A	GCAGAGACAA	6 0
C A G C A A G A A C	ACACTGTATC	T C C A A A T G A A	TAGCCTGCAA	GCCGAGGACA	C A G C C A T A T A	1 2 0
ттаттб						126
(2) INFORMATION	FOR SEQ ID NO:44:					
	JENCE CHARACTERISTI (A) LENGTH: 130 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA					
(iii)HYP0	OTHETICAL: NO					
	URE: (A) NAME/KEY: misc_ (B) LOCATION: 1130 (D) OTHER INFORMA		Oligo wps54"			
(xi)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:44:				
A C A C T C T A G A	CCACCATGGC	TGTCTTGGGG	СТССТСТТСТ	GCCTGGTGAC	ATTCCCAAGC	6 0
ТСТСТАТ	CCGCTGTCCA	GCTGCTAGAG	AGTGGTGGCG	GTCTGGTGCA	GCCAGGAGGA	1 2 0
TCTCTGAGAC						1 3 0
(2) INFORMATION	FOR SEQ ID NO:45:					
	JENCE CHARACTERISTI (A) LENGTH: 118 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA					
(iii)HYPO	OTHETICAL: NO				•	
	URE: (A) NAME/KEY: misc_ (B) LOCATION: 1118 (D) OTHER INFORMAT		Oligo wps57" _.			
(xi)SEQL	ENCE DESCRIPTION: SI	EQ ID NO:45:				
A C A C T C T A G A	AGTTAGGACT	CACCTGAAGA	GACAGTGACC	AGAGTCCCTT	GGCCCCAGTA	6 0
AGCAAAACCG	T C G T A A T T A T	AGTCCCCAGC	TCTGGCACAA	T A A T A T A T G G	СТСТСТСС	1 1 8

(2) INFORMATION FOR SEQ ID NO:46:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (\mathbf{x} i) SEQUENCE DESCRIPTION; SEQ ID NO:46:

Asp 1	11с	Val	Leu	Thr 5	Gln	Scr	Pro	A 1 a	S c r 1 0	Leu	Ala	Val	Ѕег	L c u 1 5	Gly
Gln	Arg	Ala	Thr 20	Ile	Ser	C y s	Arg	A 1 a 2 5	Scr	Gln	Scr	V a l	S c r 3 0	Thr	Scr
Thr	Туг	A s n 3 5	Туг	Меι	H i s	Trp	T y r 4 0	Gin	G 1 n	Lys	Pro	G 1 y 4 5	Gln	Pro	Pro
Lys	L e u 5 0	Leu	11с	Lys	Туг			Asn			S c r 6 0	G 1 y	Val	Pro	Ala
Arg 65	Phc	Ser	G 1 y	Scr				Thr		P h c 7 5	Thr	Lcu	A s n	I 1 c	H i s
Pro	V a l	Glu	Glu	G l u 8 5	A s p	Thr	V a l	Thr	Туг 90	Туг	C y s	Gln	H i s	S c r 9 5	Тrр
Glu	Ile	Pro	T y r 1 0 0	Thr	P h c	G 1 y	Gly	G 1 y 1 0 5	Тһг	L y s	Lси	Glu	I 1 c 1 1 0	Lys	

(2) INFORMATION FOR SEQ ID NO:47;

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:47:

G 1 u 1	Ile	Val	Меі	Thr 5	Gln	Sсг	Рго	Ala		Leu			Scr	Pro 15	G .1 y
Glu	Arg	Ala	Thr 20	Leu	Ser	C y s	Агд			Gln		V a l	S c r 3 0	Thr	Scr
Thr	Туr	A s n 3 5	Туг	Met	H i s	Trp	T y r 4 0	Gln	Gln	Lys	Pro	G 1 y 4 5	Gln	Scr	Pro
Arg			I 1 c	Lys	Туг		Scr		Lcu	Glu	S c r 6 0	G 1 y	I 1 с	Pro	Ala
Arg 65	Phe	Scr	G 1 y	Ser		Scr				Phc 75	Тһт	Leu	Thr	I 1 e	S с т 8 0
Arg	Leu	Glu	Ser	G 1 u 8 5	A s p	Phc	Ala	V a l	Туг 90	Туг	C y s	Gln	H i s	S c r 9 5	Тгр
Glu	I l e		-	Thr		-		-		-				Lys	

(2) INFORMATION FOR SEQ ID NO:48:

- ($\,i\,$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:48:

```
Giu Met Iic Leu Val Giu Ser Giy Giy Giy Leu Val Lys Pro Giy Ala
Ser Leu Lys Leu Ser Cys Ala Ala Ser Giy Phe Thr Phe Ser Asn Tyr
20 Ser Leu Ser Trp Val Arg Giy Giy Giy Arg Iic Tyr Ser Asp Arg Ciu Trp Val
Ala Ser Iic Ser Arg Giy Giy Giy Arg Iic Tyr Ser Pro Asp Asn Leu
So Giy Arg Phe Thr Iic Ser Arg Giy Giy Giy Arg Iic Tyr Ser Pro Asp Asn Leu
Gin Mei Ser Ser Iic Ser Arg Giy Giy Giy Giy Arg Iic Tyr Ser Pro Asp Asn Leu
So Gin Mei Ser Ser Leu Lys Ser Giu Asp Aia Lys Asn Thr Leu Tyr
So Gin Mei Ser Ser Leu Lys Ser Giu Asp Thr Ala Leu Tyr Tyr Cys
So Giy Thr Giy Thr Thr Val Iic Val Ser Ser
```

(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:49:

```
Glu Val Gln Leu Leu Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Gly

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Asn Tyr

Gly Leu Ser Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val

Ala Ser Ile Ser Arg Gly Gly Gly Arg Ile Tyr Ser Pro Asp Asn Leu

Gly Arg Phe Thr Ile Ser Arg Asn Asn Ser Leu Gln Ala Glu Asp Ser Tyr Gly Asn Thr Leu Tyr

Gly Arg Glu Gly Ile Tyr Tyr Ala Asp Tyr Gly Phe Phe Asp Val Trp

Gly Gln Gln Gly Thr Leu Val Thr Val Ser Ser
```

(2) INFORMATION FOR SEQ ID NO:50:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (\times i) SEQUENCE DESCRIPTION: SEQ ID NO:50:

Asp 1	Ilc	V a l	Mct	Thr 5	Gln	Scr	H i s	Lys	Phc 10	Мсι	Ser	Thr	Scr	V a l 1 5	Gly
A s p	Arg									Gln				S c r	Ala

-continued

 Val
 Trp
 His
 Gln
 Gln
 Lys
 Ser
 Gly
 Gln
 Ser
 Pro
 Lys
 Leu
 Leu
 Ile

 Tyr
 Trp
 Ala
 Ser
 Thr
 Arg
 His
 Thr
 Gly
 Val
 Pro
 Asp
 Arg
 Phc
 Thr
 Gly

 Ser
 Gly
 Ser
 Gly
 Thr
 Asp
 Phc
 Thr
 Leu
 Thr
 Ile
 Thr
 Asn
 Val
 Gln
 Ser

 Glu
 Asp
 Leu
 Ala
 Asp
 Tyr
 Phc
 Cys
 Gln
 Gn
 Tyr
 Ser
 Ile
 Phc
 Pro
 Leu

 Thr
 Phc
 Gly
 Ala
 Gly
 Thr
 Arg
 Leu
 Glu
 Leu
 Lys

(2) INFORMATION FOR SEQ ID NO:51:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:51:

 Asp
 Ile
 Gln
 Met
 Thr
 Gln
 Ser
 Pro
 Ser
 Thr
 Leu
 Ser
 Ala
 Ser
 Val
 Gly

 Asp
 Arg
 Val
 Thr
 Ile
 Thr
 Cys
 Lys
 Ala
 Ser
 Gln
 Asp
 Val
 Gly
 Ser
 Ala

 Val
 Trp
 His
 Gln
 Lys
 Pro
 Gly
 Lys
 Leu
 Leu
 Ile

 Tyr
 Trp
 Ala
 Ser
 Thr
 Arg
 His
 Thr
 Gly
 Val
 Pro
 Ser
 Arg
 Phe
 Thr
 Gly

 Ser
 Gly
 Ser
 Gly
 Thr
 Gly
 Phe
 Thr
 Leu
 Thr
 Leu
 Thr
 Gly
 Pro
 Ser
 Ser
 Leu
 Gln
 Pro
 Ro

 Ser
 Gly
 Ser
 Gly
 Thr
 Thr
 Cys
 Gln
 Thr
 Tyr
 Ser
 Ie
 Pro
 Ser
 Leu
 Gln
 Pro
 Ro

 Ser
 Asp
 Phe
 Ala<

(2) INFORMATION FOR SEQ ID NO:52:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 121 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:52:
- Gin Val Gin Leu Gin Gin Ser Asp Ala Giu Leu Val Lys Pro Giy Ala 1 10
 Ser Val Lys Ile Ser Cys Lys Val Ser Giy Tyr Thr Phe Thr Asp His 30
 Thr Ile His Trp Met Lys Gin Arg Asp Gly His Thr Arg Tyr Ser Glu Lys Phe Giy Phe 50
 Lys Giy Lys Ala Thr Leu Thr Ala Asp Lys Ser Ala Ser Thr Aia Tyr 70
 Met His Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys

```
Ala Arg Gly Arg Asp Ser Arg Glu Arg Asn Gly Phe Ala Tyr Trp Gly 105

Gln Gly Thr Leu Val Thr Val Ser Ala 120
```

(2) INFORMATION FOR SEQ ID NO:53:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 121 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:53:

```
        Gin
        Val
        Gin
        Lcu
        Val
        Gin
        Scr
        Gly
        Ala
        Glu
        Val
        Lys
        Lys
        Pro
        Gly
        Scr

        Scr
        Val
        Lys
        Val
        Scr
        Cys
        Lys
        Ala
        Scr
        Gly
        Tyr
        Thr
        Phc
        Thr
        Asp
        His

        Thr
        Ilc
        His
        Trp
        Mci
        Arg
        Gln
        Ala
        Pro
        Gly
        Gly
        Gly
        Lcu
        Glu
        Trp
        Phc

        Gly
        Tyr
        Ilc
        Tyr
        Pro
        Arg
        Asp
        Gly
        His
        Thr
        Arg
        Gly
        Fro
        Gly
        Fro
        Glu
        Lys
        Glu
```

(2) INFORMATION FOR SEQ ID NO:54:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:54:

Asp 1	Ile	Val	Leu	Thr 5	Gln	Ѕсг	Pro	Ala	S c r 1 0	Leu	Ala	Val	Ser	L c u 1 5	G 1 y
Gln	Агд	Ala	Thr 20	Ilc	Ser	C y s	Arg	A 1 a 2 5	Sсг		Sсг	V a 1	A s p 3 0	A s n	Туr
G 1 y	Ilc	S c r 3 5	Phe	Mct	Аѕп	Trp	P h c 4 0	Gln	G 1 n	Lys	Рго	G 1 y 4 5	Gln	Рго	Рго
Lys	L c u 5 0	Lcu	I 1 c	Туг	Ala	A 1 a 5 5 .	Ser	Asn	Gln	Gly	S c r 6 0	Gly	V a l	Pro	Ala
Arg 65	Phc	Scr	Gly	Ser	G l y 70	Scr	Gly	Thr	A s p	P h c 7 5	Ser	Leu	Asn	Ile	H i s
Pro	Mct	Glu	Glu	A s p 8 5	A s p	Thr	Ala	Met	T y r 9 0	Phe	C y s	Gln	G 1 n	S c r 9 5	Lys
Glu	V a l	Pro	Trp					G 1 y 1 0 5						L y s	

-continued

```
( i ) SEQUENCE CHARACTERISTICS:
```

- (A) LENGTH: 111 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:55:

 Asp I le le Gln
 Met Scr Scr Scr Leu Scr Ala Scr Val Gly

 Asp Arg
 Val Thr 20
 Thr Cys Arg 25
 Scr Glu Scr Val Asp Asn Tyr 25

 Gly I le Scr Phe Met Asn Trp Phe 40
 Gln Gln Lys Pro Gly Lys Ala Pro 45

 Lys Leu Leu Leu I le Tyr Ala Ala Scr Asn Gln Gly Scr Gly Val Pro Scr 50

 Arg Phe Scr Gly Scr Gly Scr Gly Thr Asp Phe 75
 Thr Leu Thr I le Scr 80

 Scr Leu Cu Thr I le Scr 70
 Scr Gly Thr Asp Phe Thr Cys Gln Gln Gln Scr Tys 90

(2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:56:

(2) INFORMATION FOR SEQ ID NO:57:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide

```
      Gln Val Gln Lcu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ser 1

      Scr Val Lys Val Ser Cys Lys Ala Ser Gly Gly Tyr Thr Phc Thr Asp Tyr 25

      Asn Mct His Trp Val Arg Gln Ala Pro Gly Gly Gln Gly Lcu Glu Trp Ilc 45

      Gly Tyr Ilc Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn Gln Lys Phc 50

      Lys Scr Lys Ala Thr Ilc Thr Ala Asp Glu Ser Thr Asn Thr Ala Tyr 65

      Ala Arg Gly Arg 100

      Thr Val Ser Ser
```

(2) INFORMATION FOR SEQ ID NO:58:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:58:

(2) INFORMATION FOR SEQ ID NO:59:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: pcptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:59:
- Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly

 1

 Asp Arg Val Thr Ile Thr Cys Ser Gly Ser Ser Ser Val Ser Phe Met 25

 Tyr Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr

-contin	ıu

Asp Thr Scr Asn Lcu Ala Scr Gly Val Pro Scr Arg Phc Scr Gly Scr 50 Scr Arg Phc Scr Gly Gln Gly Thr Tyr Tyr Cys Gln Gln Trp Scr Thr Tyr Pro Lcu Thr 90 Scr Thr Tyr Pro Lcu Thr 90 Scr Gly Gln Gly Thr Lys Val Glu Val Lys 105

(2) INFORMATION FOR SEQ ID NO:60:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(\times i) SEQUENCE DESCRIPTION: SEQ ID NO:60:

 Gln
 Val
 Gln
 Lcu
 Lys
 Gln
 Ser
 Gly
 Pro
 Gly
 Lcu
 Val
 Gln
 Pro
 Ser
 Gly
 Lcu
 Val
 Gln
 Pro
 Ser
 Gly
 Pro
 Ser
 Val
 Gln
 Ser
 Gly
 Pro
 Gly
 Phe
 Ser
 Val
 Thr
 Ser
 Tyr

 Gly
 Val
 His
 Trp
 Ile
 Arg
 Gln
 Ser
 Pro
 Gly
 Lys
 Gly
 Leu
 Glu
 Trp
 Leu

 Gly
 Val
 Ile
 Trp
 Ser
 Gly
 Ser
 Thr
 Asp
 Tyr
 Asp
 Tyr
 Asp
 Tyr
 Asp
 Tyr
 Asp
 Ser
 Lys
 Ser
 Gln
 Phe
 Phe
 Ala
 Ile
 Tyr
 Asp
 Ile
 Asp
 Thr
 Ala
 Ile
 Tyr
 Tyr
 Asp
 Ile
 Asp
 Thr
 Ala
 Ile
 Tyr
 Tyr
 Tyr

($\,2\,$) Information for SEQ ID NO:61:

- ($\,$ i $\,$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(\times i) SEQUENCE DESCRIPTION: SEQ ID NO:61:

G 1 u 1	V a l	Gln	Leu	L c u 5	Glu	Ser	Gly	Gly	G 1 y 1 0	Leu	Val	Gln	Pro	G l y 1 5	G 1 y
Sет	Lcu	Arg			C y s		Ala		Gly			V a 1	T h r 3 0	Scr	Туг
Gly	Val	H i s 3 5		Val		Gln			Gly		Gly	L c u 4 5	Glu	Тгр	V a l
Gly	V a 1 5 0	Ile	Тгр	Scr			Scr				A s n 6 0	Ala	Ala	Phe	Ile
S c r 6 5	Arg	P h c	Thr	Ile	S c r 7 0	Arg	A s p	A s n	Scr	L y s 7 5	Asn	Thr	Lcu	Туг	L c u 80
Gln	Mct	Asn	Ser	Leu	Gln	Ala	Glu	Asp	Thr	Ala	Ilc	Туг	Туг	Суs	Ala

Arg Ala Gly Asp Tyr Asn Tyr Asp Gly Phc Ala Tyr Trp Gly Gln Gly 100

Thr Lcu Val Thr Val Scr Scr 115

(2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:62:

 Asp Ile Val Leu Thr Gln Ser Pro Ala Thr Leu Ser Val Thr Pro Gly

 1
 5

 Asp Ser Val Ser Leu Ser Cys Arg Ala Ser Gln Ser Ile Ser Asn Asn 20

 Leu His Trp Tyr Gln Gln Lys Ser His Glu Ser Pro Arg Leu Leu Ile 35

 Lys Tyr Ala Ser Gln Ser Ille Ser Gly Ile Pro Ser Arg Phe Ser Gly 55

 Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser Val Asn Gly Val Glu Thr 65

 Glu Asp Phe Gly Met Tyr Phe Cys Gln Gln Ser Asn Ser Trp Pro His 85

 Thr Phe Gly Gly Gly Gly Thr Lys Leu Glu Ile Lys

(2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:63:

Glu Ile Val Leu Thr Gln Ser Pro Gly Thr Leu Ser Leu Ser Pro Gly

Gly

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Ile Ser Asn Asn
20

Leu His Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg Leu Leu Ile

Ser Gly

Ser Arg Leu Glu

Ser Gly

Ser Gly

Ser Gly

Ser Asn Ser Trp

Ser His

Ser Pro Gly

Ser Asn Ser Trp

Ser His

Ser Pro Gly

Ser Asn Ser Trp

Ser His

(2) INFORMATION FOR SEQ ID NO:64:

(i) SEQUENCE CHARACTERISTICS:

-continued

- (A) LENGTH: 119 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:64:

Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala 1 10 15 Thr Met Asn Trp Val Lys Gln Ser His Gly Gln Asn Leu Glu Trp 11e 35 Gly Leu Ilc Asn Pro Tyr Asn Gly Gly Thr Ser Tyr Asn Gin Lys Phe 50 Lys Gly Lys Ala Thr Lou Thr Val Asp Lys Scr Scr Asn Thr Ala Tyr 65 75 80 Arg Arg Gly Phc Arg Asp Tyr Scr Mcı Asp Tyr Trp Gly Gln Gly

Thr Scr Val Thr Val Scr Scr

(2) INFORMATION FOR SEQ ID NO:65:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 119 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Thr Mei Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 35 Gly Leu Ile Asn Pro Tyr Asn Gly Gly Thr Ser Tyr Asn Gln Lys Phe 50 55 Met Glu Leu Ser Ser Leu Phe Ser Glu Asp Thr Ala Val Tyr Tyr Cys 85 90

Thr Lcu Val Thr Val Scr Scr

(2) INFORMATION FOR SEQ ID NO:66:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 393 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA

-continued

	(i x		RE: A) NAM B) LOC														
	(x i) SEQUE	NCE DES	CRIPTIO	N: SEQ II	D NO:66:											
ATG Mc1	GAG Glu	AAA Lys	GAC Asp	ACA Thr	CTC Leu	CTG Lcu	CTA Lcu	T G G T r p	GTC Val 10	CTG Lcu	CTT Lcu	CTC Lcu	TGG Trp	GTT Val 15	C C A P r o		4 8
GGT Gly	T C C S c r	A C A T h r	G G T G 1 y 2 0	GAC Asp	ATT	GTG Val	CTG Leu	A C C T h r 2 5	CAA Gln	TCT Scr	C C A P r o	GCT Ala	T C T S c r 3 0	TTG Lcu	GCT Ala		9 6
GTG Val	T C T S c r	CTA Lcu 35	GGG Gly	CAG Gln	AGG Arg	GCC Ala	A C C T h r 4 0	ATC	TCC Scr	TGC Cys	AGA Arg	GCC Ala 45	AGC Scr	GAA Glu	AGT Scr	1	. 4 4
GTT Val	GAT Asp 50	AAT Asn	ТАТ	GGC Gly	ATT	AGT Scr 55	TTTPhc	ATG Mcl	AAC Asn	T G G T r p	TTC Phc 60	CAA Gln	CAG Gln	AAA Lys	CCA Pro	1	92
G G A G 1 y 6 5	CAG Gln	C C A P r o	C C C	AAA Lys	C T C L c u 7 0	CTC Lcu	ATC	TAT Tyr	G C T A 1 a	G C A A 1 a 7 5	TCC Scr	AAC Asn	CAA Gln	GGA Gly	T C C S c r 8 0		4 0
G G G G l y	GTC Val	CCT Pro	GCC Ala	A G G A r g 8 5	TTTPhc	AGT Scr	GGC Gly	AGT Ser	G G G G 1 y 9 0	TCT Ser	G G G G l y	ACA Thr	GAC Asp	TTC Phc 95	AGC Scr	2	8 8 8
CTC Lcu	AAC Asn	ATC	CAT His 100	C C T P r o	ΑTG	GAG Glu	GAG Glu	G A T A s p 1 0 5	G A T A s p	ACT Thr	G C A A 1 a	ΑTG Mcι	TAT Tyr 110	TTC Phc	TGT Cys	3	3 6
CAG Gln	C A A G 1 n	A G T S c r 1 1 5	AAG Lys	GAG Glu	GTT Val	C C G P r o	T G G T r p 1 2 0	ACG Thr	TTC Phe	GGT Gly	GGA Gly	G G C G 1 y 1 2 5	ACC Thr	AAG Lys	CTG Leu	3	8 4
	ATC IIc 130															3	9 3

(2) INFORMATION FOR SEQ ID NO:67:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 131 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:67:

M c t 1	Glu	Lys	A s p	Thr 5	Lcu	Lcu	Lcu	Trp	V a 1 1 0	Leu	Lcu	Lcu	Тrp	V a l 1 5	Pro	
Gly	Scr	Тһг	G l y 2 0	A s p	I 1 c	Val	Leu	Thr 25	Gln	Ser	Pro	Ala	S c r 3 0	Lcu	Ala	
Val	Ser	L c u 3 5	Gly	Gln	Агд	Ala	Thr 40	I 1 c	S c r	C y s	Arg	A 1 a 4 5	Scr	Glu	S c r	
Val	A s p 5 0	Asn	Туг	G 1 y	Ilc	S c r 5 5	Phc	Μcι	A s n	Тгр	P h c 6 0	G 1 n	Gln	L y s	Pro	
G 1 y 6 5	Gln	Рго	Pro	Lys	L c u 70	Lcu	I 1 c	Туг	Ala	A 1 a 7 5	Ser	A s n	Gln	G 1 y	S c r 8 0	
Gly	V a 1	Pro	Ala	A r g 8 5	Phc	Ser	Gly	Scr	G 1 y 9 0	Scr	Gly	Thr	A s p	P h c 9 5	Ser	
Leu	A s n	Ilc	H i s 100	Pro	Μcι	Glu	Glu		A s p	Thr	Ala	Met	T y r 1 1 0	Phc	Суs	
Gln	Gln	S c r 1 1 5	L y s	Glu	V a 1	Pro	Trp 120	Thr	P h c	G 1 y	G 1 y	G 1 y 1 2 5	T h r	Lys	Leu	
Glu	Ile	Lvs														

Glu Ilc Lys

1 3 0

(2) INFORMATION FOR SEQ ID NO:68:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 405 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: cDNA

- (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..405

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:68:

	GGA Gly								4 8
	CAC His								9 6
	GGG Gly								1 4 4
	G A C A s p 5 0								1 9 2
	Т G G Т г р								2 4 0
	AAG Lys								288
	GCC Ala							G C A A 1 a	3 3 6
	ТАСТуг								3 8 4
ACC Thr	T C A S c r 1 3 0	ACC Thr							4 0 5

($\,2\,$) INFORMATION FOR SEQ ID NO:69:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 135 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Ме t 1	Gly	Тгр	Scr	Trp - 5	Ile	Phc	Leu	Phc	L c u 1 0	Leu	Scr	Gly	Thr	A 1 a 1 5	Gly
Val	His	Scr										Glu			Lys
Pro	G 1 y	A 1 a 3 5	Scr	V a 1	Lys	Ilc	S c r 4 0	Суs	L y s	Ala	Sсг	G 1 y 4 5	Туг	Thr	Phc
Thr	A s p 5 0	Туг	Asn	Μcι	H i s	T r p 5 5	V a 1	Lys	Gln	Scr	H i s 60	Gly	Lys	Ser	Lcu
Glu	Trn	Ile	Glv	Тут	Ilc	Туг	Рто	Tvr	Asn	Glv	Glv	Thr	Glv	Туг	Asn

(2) INFORMATION FOR SEQ ID NO:70:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:70:

Asp 1	Ilc	Gln	Μcι	Thr 5	Gln	Scr	Рго	Ser	Thr 10	Lcu	Scr	Ala	Ser	V a 1 1 5	Gly
Asp	Arg	Val	T h r 2 0	11с	Thr	Суs	Arg	A 1 a 2 5		Gln			A s n 3 0	Thr	Тгр
Leu	Ala	T r p 3 5	Туг	Gln	Gln	Lys	P r o 4 0	Gly	L y s	Ala	Pro	L y s 4 5	Lcu	Leu	Mct
Туг	L y s 5 0	Ala	Scr	Scr	Leu	G l u 5 5	Ser	Gly	V a 1	Pro	S c r 6 0	Arg	Phe	Ilc	Gly
S c r 6 5	Gly	Ser	Gly	Thr	G 1 u 7 0	Phc	Thr	Lcu	Thr	I I c 7 5	Scr	Ser	Leu	Gln	Pro 80
Asp	A s p	P _, h c	Ala	Thr 85	Туг	Туг	C y s	Gln	G l n 9 0	Туг	A s n	Ser	A s p	S c r 9 5	L y s
Меι			G l n 100												

(2) INFORMATION FOR SEQ ID NO:71:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(\star i) SEQUENCE DESCRIPTION: SEQ ID NO:71:

Asp 1	Ilc	Gln	Mct	Thr 5	Gln	Scr	Pro	Ser	S c r 1 0	Lcu	Scr	Ala	Ser	V a 1 1 5	Gly
Asp	Arg	Val	Thr 20	Ile	Thr	Суs	Arg	A 1 a 2 5		Glu	Scr	V a 1	A s p 3 0	A s n	Туг
Gly	I 1 c	S c r 3 5	Phe	Меι	A s n	Trp	P h c 4 0	Gln	Gln	Lys	Pro	G 1 y 4 5	Lys	A 1 a	Рго
Lys	L c u 5 0	Lcu	Ilc	Туг	Ala	A 1 a 5 5	Scr	A s n	GIn	Gly	S c r 6 0	Gly	V a 1	Рго	Scr
Arg 65	Phe	Ser	Gly	Ser	G 1 y 7 0	Scr.	Gly	Thr	Asp	Phc 75	Thr	Leu	Thr	I 1 c	S c r 8 0
Ser	Lcu	Gln	Pro	A s p 8 5	A s p	Phc	Ala	Thr	T y r 9 0	Туг	C y s	Gln	Gln	S c r 9 5	Lys
Glu	V a 1	Pro	Trp	Thr	Phc	Gly	Gln	Gly	Thr	Lys	V a l	G 1 u	Ilc	Lys	

133 134

(2) INFORMATION FOR SEQ ID NO:72:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:72:

G 1 n	V a l	Gln	Leu	Val 5	Gln	Ser	G 1 y	Ala	G 1 u 1 0	V a l	L y s	Lys	Рго	G l y 1 5	Scr
Scr	Val	Lys	V a 1 2 0	Ser	Суs	Lys	Ala	S c r 2 5	Gly	Gly	Thr	Phc	S с г 3 О	Arg	Sег
Ala	Ilc	I 1 c 3 5	Тrр	V a l	Агд	Gln	A 1 a 4 0	Pro	Gly	Gln	Gly	L c u 4 5	Glu	Тгр	Мсι
Gly	G 1 y 5 0	Ile	V a l	Pro	Меι	P h c 5 5	Gly	Pro	Pro	A s n	Туг 60	Ala	Gln	Lys	P h c
G 1 n 6 5	Gly	Arg	Val	Thr	I 1 e 7 0	Thr	Ala	A s p	Glu	S c r 7 5	Thr	Asn	Thr	Ala	Туг 80
Меι	Glu	Lси	Ser	S c r 8 5	Lcu	Arg	Ser	Glu	A s p 9 0	Thr	Ala	Phe	Туг	P h c 9 5	C y s
Ala	Gly	Gly	T y r 1 0 0	Gly	Ile	Туг	Ser	Pro 105	Glu	Glu	Туг	Asn	G l y 1 1 0	Gly	Leu
V a l	Thr	V a l	Scr	Ser											

(2) INFORMATION FOR SEQ ID NO:73:

(i) SEQUENCE CHARACTERISTICS:

1 1 5

- (A) LENGTH: 116 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:73:

G 1 n	V a l	Gln	Leu	V a 1 5	Gln	Ser	G 1 y	Ala	G 1 u 1 0	V a l	Lys	L y s	Pro	G l y 1 5	Ser
Ser	V a 1	Lys	V a 1 2 0	Scr	C y s	L y s	Ala	S c r 2 5	Gly	Туг	Thr	P h c	T h r 3 0	A s p	Туг
Αsπ	Met	H i s 3 5	Тгр	V a 1	Arg	Gln	A 1 a 4 0	Pro	G 1 y	Gln	Gly	Lси 45	Glu	Тгр	Ile
Gly	T y r 5 0	Ilc	Туr	Pro	Туг	A s n 5 5	G 1 y	Gly	Thr	Gly	Туг 60	Asn	Gln	Lys	Phe
L y s 6 5	Ser	Lys	Ala	Thr	I 1 c 7 0	Thr	Ala	A s p	Glu	S c r 7 5	Thr	A s n	Thr	Ala	Туг 80
Mct	Glu	Lcu	Ser	S c r 8 5	Leu	Arg	Scr	Glu	A s p 9 0	Thr	Ala	V a l	Туг	Туг 95	C y s
Ala	Arg	Gly	Arg 100	Pro	Ala	Mct	A s p	T y r 1 0 5	Тгр	Gly	Gln	Gly	Thr 110	Leu	V a 1
Thr	V a l	Ser	Ser												

1 1 5

(2) INFORMATION FOR SEQ ID NO:74:

(i) SEQUENCE CHARACTERISTICS:

	(A) LENGTH: 132 base (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear					
(ii) MOL	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQ	UENCE DESCRIPTION: SE	Q ID NO:74:				
ТАТАТСТАСА	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	тсстсствтс	AGGAACTGCT	6 0
GGCGTCCACT	CTCAGGTTCA	GCTGGTGCAG	TCTGGAGCTG	AGGTGAAGAA	GCCTGGGAGC	1 2 0
T C A G T G A A G G	тт					1 3 2
(2) INFORMATION	FOR SEO ID NO:75:					
	UENCE CHARACTERISTIC (A) LENGTH: 133 base p (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SE	Q ID NO:75:				
AGCCGGTACC	ACCATTGTAA	G G A T A A A T A T	A T C C A A T C C A	TTCCAGGCCT	TGGCCAGGAG	6 0
CCTGCCTCAC	CCAGTGCATG	TTGTAGTCAG	TGAAGGTGTA	GCCAGAAGCT	TTGCAGGAAA	1 2 0
CCTTCACTGA	GCT					1 3 3
(2) INFORMATION	FOR SEQ ID NO:76:					
	JENCE CHARACTERISTIC (A) LENGTH: 112 base p (B) TYPE: nucleic acid (C) STRANDEDNESS: s: (D) TOPOLOGY: linear ECULE TYPE: DNA (oligon	pairs ingle				
(xi)SEQU	JENCE DESCRIPTION: SE	Q ID NO:76:				
Т G G T G G T A C C	GGCTACAACC	AGAAGTTCAA	GAGCAAGGCC	ACAATTACAG	C A G A C G A G A G	6 0
T A C T A A C A C A	GCCTACATGG	AACTCTCCAG	CCTGAGGTCT	GAGGACACTG	C A	1 1 2
(2) INFORMATION	FOR SEQ ID NO:77:					
	JENCE CHARACTERISTIC (A) LENGTH: 111 base p (B) TYPE: nucleic acid (C) STRANDEDNESS: si (D) TOPOLOGY: linear	airs				
(ii) MOL	ECULE TYPE: DNA (oligor	nucleotide)				
(xi)SEQL	JENCE DESCRIPTION: SE	Q ID NO:77:				
T A T A T C T A G A	GGCCATTCTT	ACCTGAAGAG	ACAGTGACCA	GAGTCCCTTG	GCCCCAGTAG	6 0
TCCATAGCGG	GGCGCCCTCT	TGCGCAGTAA	TAGACTGCAG	TGTCCTCAGA	C .	1 1 1
(2) INFORMATION I	FOR SEQ ID NO:78:					
	JENCE CHARACTERISTIC (A) LENGTH: 122 base p (B) TYPE: nucleic acid (C) STRANDEDNESS: si (D) TOPOLOGY: linear	airs				
(ii) MOLI	ECULE TYPE: DNA (oligon	nucleotide)				
(xi)SEQU	ENCE DESCRIPTION: SE	Q ID NO:78:				

-continued TATATCTAGA CCACCATGGA GAAAGACACA CTCCTGCTAT GGGTCCTGCT TCTCTGGGTT 60 CCAGGTTCCA CAGGTGACAT TCAGATGACC CAGTCTCCGA GCTCTCTGTC CGCATCAGTA 120 1 2 2 (2) INFORMATION FOR SEQ ID NO:79: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 122 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:79: TCAGAAGCTT AGGAGCCTTC CCGGGTTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 60 AATTGTCGAC ACTTTCGCTG GCTCTGCATG TGATGGTGAC CCTGTCTCCT ACTGATGCGG A C 1 2 2 (2) INFORMATION FOR SEQ ID NO:80: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 119 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:80: TCCTAAGCTT CTGATTTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 60 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 119 (2) INFORMATION FOR SEQ ID NO:81: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:81: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 60 TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 118 (2) INFORMATION FOR SEQ ID NO:82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 381 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..381 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA 48 Met Val Phe Thr Pro Gln Ile Leu Gly Leu Met Leu Phe Trp Ile Ser 1.0

G C C A 1 a	T C C S c r	AGA Arg	G G T G 1 y 2 0	GAT Asp	ATT	GTG Val	CTA Leu	A C T T h r 2 5	CAG Gln	TCT Scr	C C A P r o	GCC Ala	A C C T h r 3 0	CTG Lcu	TCT Scr	9 6
GTG Val	ACT Thr	C C G P r o 3 5	GGA Gly	GAT Asp	AGC Scr	GTC Val	A G T S c r 4 0	CTT Lcu	TCC Scr	TGC Cys	AGG Arg	G C C A 1 a 4 5	AGC Scr	CAA Gln	AGT Scr	1 4 4
ATT	AGC Scr 50	AAC Asn	AAC Asn	CTA Leu	CAC His	T G G T r p 5 5	TAT Tyr	CAA Gln	CAA Gln	AAA Lys	T C A S c r 6 0	CAT His	GAG Glu	TCT Scr	C C A P r o	192
AGG Arg 65	CTT Lcu	CTC Lcu	ATC	AAG Lys	T A T T y r 7 0	GCT Ala	TCC Scr	CAG Gln	T C C S c r	ATC Ilc 75	TCT Scr	GGG Gly	ATC Ilc	C C C P r o	T C C S c r 8 0	2 4 0
AGG Arg	TTC Phc	AGT Scr	GGC Gly	AGT Scr 85	G G A G I y	T C A S c r	GGG Gly	ACA Thr	G A T A s p 9 0	TTC Phe	ACT Thr	CTC Lcu	AGT Scr	GTC Val 95	AAC Asn	2 8 8
GGT Gly	GTG Val	GAG Glu	A C T T h r 1 0 0	GAA Glu	GAT Asp	TTTPhc	G G A G I y	A T G M c t 1 0 5	TAT Tyr	TTC Phe	TGT Cys	CAA Gln	C A G G 1 n 1 1 0	AGT Scr	AAC Asn	3 3 6
AGT Scr	TGG Trp	C C T P r o 1 1 5	CAT His	ACG Thr	TTC Phe	GGA Gly	G G G G l y 1 2 0	GGG Gly	ACC Thr	AAG Lys	CTG Lcu	G A A G 1 u 1 2 5	ATA Ilc	AAA Lys		3 8 1

(2) INFORMATION FOR SEQ ID NO:83:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 127 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:83:

Mc t	V a 1	Phe	Thr	P r o 5	Gln	Ilc	Leu	Gly	L c u 1 0	Met	Leu	Phe	Тrр	I 1 e 1 5	Scr
Ala	Scr	Arg	G 1 y 2 0	Asp	I 1 c	Val	Lcu	Thr 25	Gln	Scr	Pro	Ala	T h r 3 0	Lcu	S c r
Val	Thr	Pro 35	Gly	A s p	Ser	V a 1	S c r 4 0	Lcu	Scr	C y s	Arg	A 1 a 4 5	Scr	Gln	Scr
I 1 c	S c r 5 0	Asn	Asn	Lcu	His	T r p 5 5	Туг	Gln	Gln	L y s	S c r 6 0	H i s	Glu	Ser	Pro
Arg 65	Lcu	Lcu	I 1 c	Lys	T y r 7 0	Ala	Scr	Gln	Scr	I 1 c - 7 5	Scr	Gly	I 1 c	Pro	S c r 8 0
Arg	P h c	Scr	Gly	S c r 8 5	G 1 y	Scr	G 1 y	Thr	A s p 9 0	Phe	Thr	Leu	Ser	V a l 9 5	Asn
Gly	Val	Glu	Thr 100	Glu	A s p	Phc	Gly	M c t 1 0 5	Туг	Phc	Суs	Gln	G l n 1 1 0	Scr	Asn
S c r	Тгр	Pro 115	H i s	Thr	Phe	Gly	G 1 y 1 2 0	G 1 y	Thr	Lys	Leu	Glu 125	Ilc	Lys	

(2) INFORMATION FOR SEQ ID NO:84:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 414 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA
- (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..414

-continued

(x i)	SEQUE	ICE DES	CRIPTIO	N: SEQ II	NO:84:					
									GCA Ala 15	4 8
									GTG Val	9 6
									TCA Scr	1 4 4
									AAC Asn	192
									TAC Tyr	2 4 0
									T C C S c r 9 5	288
									G C A A 1 a	3 3 6
									T A C T y r	3 8 4
				GTC Val						4 1 4

(2) INFORMATION FOR SEQ ID NO:85:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 138 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:85:

M c 1	Gly	Trp	Ser	T r p	Ilc	Phc	Leu	Phc	L c u 1 0	Lcu	Scr	Gly	Thr	A 1 a	Gly
Val	His	Scr	G 1 u 2 0	V a 1	Gln	Leu	G 1 n	G 1 n 2 5	Scr	G 1 y	Pro	Glu	L c u 3 0	Val	Lys
Рго	G 1 y	A 1 a 3 5	Ser	Mct	L y s	I 1 c	S e r 4 0	Суs	Lys	Ala	Scr	V a 1 4 5	Туr	Scr	Phc
Thr	G 1 y 5 0	Туг	Thr	Мει	A s n	T r p 5 5	V a l	Lys	Gln	Scr	H i s 6 0	G 1 y	G 1 n	Asn	Lcu
G 1 u 6 5	Тгр	I 1 e	Gly	Leu	I I c 7 0	Asn	Pro	Туг	A s n	G 1 y	Gly	Thr	Ser	Туг	A s n 8 0
Gln	L y s	Phc	L y s	G l y 8 5	Lys	AIa	Thr	Leu	Thr 90	V a 1	A s p	L y s	Scr	S c r 9 5	Asn
Thr	Ala	Туг	Met 100	Glu	Leu	Leu	Ser	L c u 1 0 5	Thr	Scr	Ala	A s p	S c r 1 1 0	Ala	V a l
Туг	Туr	C y s	Thr	Arg	Arg	G l y	P h c 1 2 0	Arg	A s p	Туr	Ser	M c t 1 2 5	A s p	Туr	Тгр
Gly	G 1 n 1 3 0	Gly	Thr	Ser	V a 1	T h r 1 3 5	V a l	S e r	Ser						

```
( i ) SEQUENCE CHARACTERISTICS:
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- (A) LENGTH: 108 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear $\,$

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:86:

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Gln Ser Gln Ser Gln Ser Leu Ser Pro Gly
Thr Leu Ser Gln Ser Val Ser Gly
Tyr Leu Gly Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg Leu Cys
Ileu Tyr Gly Ala Ser Ser Arg Ala Thr Gly Ile Pro Asp Arg Leu Cu
Gly Ser Gly Ser Gly Trp Tyr Tyr Asp Phe Thr Leu Thr Ile Ser Arg Leu Glu
Fro Glu Asp Phe Ala Val Tyr Tyr Tyr Cys Gln Gln Tyr Gly Ser Leu Ser Gly
Thr Phe Gly Gln Gln Gly Thr Lys Val Ile Lys

(2) INFORMATION FOR SEQ ID NO:87:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

($\,x\,$ i $\,$) SEQUENCE DESCRIPTION: SEQ ID NO:87:

G 1 u 1	Ilc	Val	Leu	T h r 5	Gln	Scr	Pro	G 1 y	Thr 10	Leu	Scr	Lcu	Scr	Pro 15	G 1 y
Glu	Arg	Ala	Thr 20	Lcu	Ser	C y s	Arg			Gln			S e r 3 0	A s n	A s n
Leu	His	T r p	Туг	Gln	Gln	L y s	Pro 40	Gly	Gln	Ala	Pro	Arg 45	Leu	Leu	11е
Lys	Туг 50	Ala	Ser	Gln		I 1 e 5 5	S c r	G 1 y	I 1 c	Pro	A s p 6 0	Агд	Phc	Ser	Gly
S e r 6 5	Gly	Ser		Thr	A s p 7 0	P h c	Thr	Leu	Thr			Arg		Glu	Pro 80
Glu	A s p	Phc	Ala	V a 1 8 5	Туг	Туг	Суs			Scr		Ser	Тгр	Pro 95	His
Thr				G 1 y						L y s					

(2) INFORMATION FOR SEQ ID NO:88:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:88:

-continued

 Ser
 Val
 Arg
 Val
 Ser
 Cys
 Lys
 Thr
 Ser
 Gly
 Gly
 Thr
 Phe
 Val
 Asp
 Tyr

 Lys
 Gly
 Leu
 Trp
 Val
 Arg
 Gln
 Ala
 Pro
 Gly
 Lys
 Gly
 Leu
 Glu
 Trp
 Val

 Gly
 Gln
 Ilc
 Pro
 Leu
 Arg
 Phe
 Asn
 Gly
 Glu
 Val
 Lys
 Pro
 Ser
 Phe
 Asn
 Pro
 Gly
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(2) INFORMATION FOR SEQ ID NO:89:

- ($\,\,$ i $\,$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: pcptide
- ($\,x\,$ i $\,$) SEQUENCE DESCRIPTION: SEQ ID NO:89:

(2) INFORMATION FOR SEQ ID NO:90:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 129 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA (oligonucleotide)
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:90:

TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTCA GACTCATGCT CTTCTGGATT 60

TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG 120

GGAGAAAGG

	UENCE CHARACTERISTI (A) LENGTH: 128 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	onucleotide)				
(xi)SEQU	UENCE DESCRIPTION: SI	EQ ID NO:91:				
T A G A C A G A A T	TCACGCGTAC	TTGATAAGTA	GACGTGGAGC	TTGTCCAGGT	ттттсттсст	6 0
ACCAGTGTAG	GTTGTTGCTA	ATACTTTGGC	TGGCCCTGCA	GGAAAGTGTA	GCCCTTTCTC	1 2 0
CCGGTGAT						1 2 8
(2) INFORMATION	FOR SEQ ID NO:92:					
	JENCE CHARACTERISTI (A) LENGTH: 113 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	mucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SE	EQ ID NO:92:				
AAGAGAATTC	ACGCGTCCCA	GTCCATCTCT	GGAATACCCG	ATAGGTTCAG	TGGCAGTGGA	6 0
TCAGGGACAG	ATTTCACTCT	C A C A A T A A G T	AGGCTCGAGC	CGGAAGATTT	TGC	1 1 3
(2) INFORMATION	FOR SEQ ID NO:93:					
	JENCE CHARACTERISTII (A) LENGTH: 116 base (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQL	JENCE DESCRIPTION: SE	EQ ID NO:93:				
ГАСАТСТАСА	GTTGAGAAGA	CTACTTACGT	T T T A T T T C T A	CCTTGGTCCC	TTGTCCGAAC	6 0
GTATGAGGCC	AACTGTTACT	CTGTTGACAA	TAATACACAG	C A A A A T C T T C	CGGCTC	1 1 6
(2) INFORMATION I	FOR SEQ ID NO:94:					
	DENCE CHARACTERISTIC (A) LENGTH: 134 base (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii) MOLI	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SE	Q ID NO:94:				
ГАТАТСТАСА	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	TCCTCCTGTC	AGGAACTGCA	6 0
GGTGTCCACT	CTCAAGTCCA	ACTGGTACAG	TCTGGAGCTG	AGGTTAAAAA	.GCCTGGAAGT	1 2 0
T C A G T A A G A G	тттс					1 3 4
(2) INFORMATION I	FOR SEQ ID NO:95:					
(JENCE CHARACTERISTIC (A) LENGTH: 134 base; (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii) MOLI	ECULE TYPE: DNA (oligo	nucleotide)				

-continued (x i) SEQUENCE DESCRIPTION: SEQ ID NO:95: TATATAGGTA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCA 60 TGCCTGTCTC ACCCAGTTCA TGGTATACCC AGTGAATGAG TATCCGGAAG CTTTGCAGGA 1 2 0 AACTCTTACT GAAC 1 3 4 (2) INFORMATION FOR SEQ ID NO:96: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 116 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:96: TATATAGGTA CCAGCTACAA CCAGAAGTTC AAGGGCAGAG TTACAGTTTC TTTGAAGCCT 60 TCATTTAACC AGGCCTACAT GGAGCTCAGT AGTCTGTTTT CTGAAGACAC TGCAGT 116 (2) INFORMATION FOR SEQ ID NO:97: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 116 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY; linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:97: TATATCTAGA GGCCATTCTT ACCTGAGGAG ACGGTGACTA AGGTTCCTTG ACCCCAGTAG 60 TCCATAGAAT AGTCTCGAAA CCCCCGTCTT GTACAGTAAT AGACTGCAGT GTCTTC 116 (2) INFORMATION FOR SEO ID NO:98: (i) SEOUENCE CHARACTERISTICS: (A) LENGTH: 408 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..408 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:98: ATG CAT CAG ACC AGC ATG GGC ATC AAG ATG GAA TCA CAG ACT CTG GTC 4 8 Met His Gln Thr Ser Met Gly Ile Lys Met Glu Ser Gln Thr Leu Val 10 TTC ATA TCC ATA CTG CTC TGG TTA TAT GGT GCT GAT GGG AAC ATT GTT 96 Phc Ilc Scr Ilc Leu Leu Trp Leu Tyr Gly Ala Asp Gly Asn Ilc Val 2 0 1 4 4 4 0 ACC TTG AGC TGC AAG GCC AGT GAA AAT GTG GAT ACT TAT GTA TCC TGG 192 Thr Leu Ser Cys Lys Ala Ser Glu Asn Val Asp Thr Tyr Val Ser Trp 5 5 60 TAT CAA CAG AAA CCA GAG CAG TCT CCT AAA CTG CTG ATA TAT GGG GCA 2 4 0 Tyr Gln Gln Lys Pro Glu Gln Ser Pro Lys Leu Leu Ile Tyr Gly Ala 6 5 7 0 7 5 8.0

тсс	AAC	CGG	TAC	ACT	GGG	GTC	ссс	GAT	CGC	ттс	ACG	GGC	AGT	GGA	тст	288
Scr	Asn	Arg	Туг	Thr 85	Gly	Val	Pro	Asp	Arg 90	Phc	Thr	G 1 y	Ser	G 1 y 9 5	Scr	
GCA	ACA	GAT	ттс	ACT	СTG	ACC	АТС	AGC	AGT	GTG	CAG	GCT	GAA	GAC	СТТ	3 3 6
Ala	Thr	Asp	Phc 100	Thr	Lcu	Thr	Ilc	S c r 1 0 5		V a l	Gln	Ala	G l u 1 1 0	Asp	Lcu	
GCA	GAT	тат	CAC	TGT	GGA	CAG	AGT	TAC	AAC	ТАТ	CCA	ттс	ACG	ттс	GGC	3 8 4
Ala	Asp	Туг 115	His	Суs	Gly	Gln	S c r 1 2 0	Туг	Asn	Туг	Pro	Phc 125	Thr	Рһс	Gly	
ТСG	GGG	ACA	A A G	ттс	GAA	АТА	A A G									4 0 8
Scr	G l y 130	Thr	-		Glu		Lys									

(2) INFORMATION FOR SEQ ID NO:99:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 136 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:99:

Mct 1	H i s	Glni	Thr	S c r 5	Μcι	Gly	I 1 c	L y s	M c t 1 0	Glu	Ser	Gln	Thr	L c u 1 5	V a 1
Phe	Ile	Scr	I 1 c 2 0	Leu	Lcu	Trp	Lcu	T y r 2 5	Gly	Ala	A s p	Gly	A s n 3 0	Ilc	V a 1
Mct	Thr	G 1 n 3 5	Scr	Pro	Lys	Scr	M c t 4 0	Туг	Val	Scr	I 1 c	G 1 y 4 5	Glu	Arg	Val
Thr	L c u 5 0	Scr	C y s	Lys	Ala	S c r 5 5	Glu	A s n	V a 1	A s p	T h r 6 0	Туг	V a l	Ser	Тrр
T y r 6 5	Gln	Gln	Lys	Pro	G 1 u 7 0	Gln	Scr	Рго	Lys	L c u 7 5	Lcu	Ile	Туг	Gly	A 1 a 8 0
Scr	Asn	Arg	Туг	Thr 85	Gly	V a 1	Pro	A s p	Arg 90	Phe	Thr	G 1 y	Ser	G 1 y 9 5	Ser
Ala	Thr	A s p	P h c 1 0 0	Thr	Lcu	Thr	I 1 c	S e r 1 0 5	Ser	Val	Gln	Ala	G 1 u 1 1 0	A s p	Leu
Ala	A s p	T y r 1 1 5	H i s	C y s	G 1 y	Gln	S c r 1 2 0	Туг	A s n	Туг	Pro	P h c 1 2 5	T h r	Phc	Gly
Ser	G 1 y 1 3 0		L y s				•								

($\,2\,$) Information for SEQ ID NO:100:

- ($\,\mathrm{i}\,$) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 456 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA
- (i x) FEATURE:

 - (A) NAME/KEY: CDS (B) LOCATION: 1..456
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:100:
- ATG ACA TCA CTG TTC TCT CTA CAG TTA CCG AGC ACA CAG GAC CTC GCC Met Thr Ser Leu Phe Ser Leu Gln Leu Pro Ser Thr Gln Asp Leu Ala 1 5 ATG GGA TGG AGC TGT ATC ATC CTC TTC TTG GTA GCA ACA GCT ACA GGT Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly 96

-continued

					CAA											1 4 4
Val	Lcu	Ser 35	Gln	Val	Gln	Leu	G 1 n 4 0	Gln	Pro	Gly	Ala	Asp 45	Leu	Val	Mci	
						~							m . a		m m 0	100
					AAG											1 9 2
Pro	5 O	Ala	Pro	vai	Lys	5 5	SCI	Cys	Leu	Ala	60	Gly	1 y 1	116	r II C	
ACC	AGC	тсс	TGG	АТА	AAC	TGG	GTG	AAG	CAG	A G G	ССТ	GGA	CGA	GGC	СТС	2 4 0
					Asn											
6 5			•		7 0	-				7 5					8 0	
GAG	ТGG	АТТ	G G A	A G G	АТТ	GAT	ССТ	тсс	GAT	GGT	GAA	GTT	CAC	TAC	ААТ	2 8 8
Glu	Тгр	Ilc	Gly	_	Ilc	Asp	Pro	Ser	_	Gly	Glu	Val	His	-	Asn	
				8 5					9 0					9 5		
CAA	GAT	ттс	A A G	GAC	A A G	GCC	ACA	CTG	АСТ	GTA	GAC	AAA	тсс	ТСС	AGC	3 3 6
GIn	Asp	Phe	•	Asp	Lys	Ala	Thr		Thr	V a l	Asp	Lys		Ser	Scr	
			100					1 0 5					1 1 0			
ACA	GCC	TAC	АТС	CAA	СТС	AAC	AGC	CTG	ACA	тст	GAG	GAC	тст	GCG	GTC	3 8 4
Thr	Ala	-	Ile	Gln	Lcu	A s n		Lcu	Thr	Scr	Glu		Scr	Ala	Val	
		1 1 5					120					1 2 5				
					GGA											4 3 2
Туг	-	Суs	Ala	Агд	Gly		Lcu	Pro	Тrр	Рһс		Asp	Trp	Gly	Gln	
	1 3 0					1 3 5					1 4 0					
GGG	ACT	CTG	GTC	ACT	GTC	тст	G C A									4 5 6
•	Thr	Leu	Val	Thr	V a l	Scr	Ala									
1 4 5					150											

(2) INFORMATION FOR SEQ ID NO:101:

- $\left(\begin{array}{c} i \end{array} \right)$ SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 152 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:101:

Met 1	Thr	Scr	Leu	Phc 5	Sет	Leu	Gln	Leu	P r o	Ser	Thr	Gln	A s p	L c u 1 5	Ala
Mct	Gly	Тгр	S c r 2 0	C y s	Ile	Ilc	Leu	P h e 2 5	Lcu	V a l	Ala	Thr	A 1 a 3 0	Thr	Gly
V a 1	Lcu	S c r 3 5	G 1 n	V a l	Gln	Leu	G 1 n 4 0	Gln	Pro	G 1 y	Ala	A s p 4 5	Leu	V a 1	Met
Pro	G 1 y 5 0	Ala	Pro	V a l	Lys	L c u 5 5	Scr	C y s	Lcu	Ala	S c r 6 0	G 1 y	Туг	Ilc	Phc
T h r 6 5	Scr	Scr	Тгр	ІІс	A s n 7 0	Тгр	V a 1	Lys	G 1 n	Arg 75	Pro	G I y	Arg	Gly	L c u 8 0
Glu	Тгр	I 1 c	G 1 y	Arg 85	Ilc	A s p	Pro	Sсг	A s p 9 0	Gly	G1 u	Val	H i s	T y r 9 5	Asn
Gln	A s p	Phc	L y s 1 0 0	A s p	L y s	Ala	Thr	L e u 1 0 5	Thr	Val	A s p	Lys	S e r 1 1 0	Scr	Sсг
Thr	Ala	T y r 1 1 5	Ilc	Gln	Leu	Asn	S c r 1 2 0	Lcu	Thr	Scr	Glu	A s p 1 2 5	Scr	Ala	V a l
Туг	Туг 130	C y s	Ala	Arg	Gly	Phc 135	Lcu	Рго	Trp	Phc	A 1 a 1 4 0	A s p	Trp	Gly	Gln
G 1 y 1 4 5	Thr	Leu	V a I	Thr	V a 1 1 5 0	Ser	Ala								

($\,2\,$) Information for SEQ ID NO:102:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids (B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:102:

Asp 1	Ilc	Gln	Μcι	Thr 5	Gln	Scr	Pro	Scr	T h r 1 0	Leu	Scr	Ala	Sег	V a 1 1 5	Gly
A s p	Arg	V a 1	Thr 20	I I c	Thr	Суs	Arg	A 1 a 2 5	Scr	Gln	Ser	Ile	А s п 3 0	Thr	Тгр
Leu	Ala	Т r р 3 5	Туг	Gln	Gln	Lys	Pro 40	Gly	L y s	Ala	Рго	L y s 4 5	Leu	Leu	Mct
Туг	L y s 5 0	Ala	Ser	S c r	Lcu	G 1 u 5 5	Scr	Gly	V a l	Pro	S c r 6 0	Arg	Phe	I 1 c	G 1 y
S c r 6 5	G 1 y	Scr	Gly	Thr	Glu 70	Phc	Thr	Leu	Thr	I 1 c	Ser	Scr	Leu	Gln	Pro 80
A s p	A s p	Phc	Ala	Thr 85	Туг	Туг	C y s	Gln	G 1 n 9 0	Туг	A s n	Ser	A s p	S c r 9 5	Lys
Мει	Phc	Gly	G 1 n 1 0 0	G 1 y	Thr	Lys	V a l	G 1 u 1 0 5	V a l	Lys					

(2) INFORMATION FOR SEQ ID NO:103:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:103:

Asp 1	Ile	Gln	Mct	Thr 5	Gln	Ser	Pro	Sсг	Thr 10	Lcu	Scr	Ala	Ser	V a 1 1 5	G 1 y
Asp	Arg	Val	Thr 20	Ilc	Thr	Суs	Lys	A 1 a 2 5	Scr	Glu	A s n	V a 1	A s p 3 0	Thr	Туг
Val	Scr	Trp 35	Туг	Gln	Gln	Lys	Pro 40	Gly	Lys	Ala	Pro	L y s 4 5	Lcu	Leu	I 1 c
Туг	G 1 y 5 0	Ala	Ser	Asn	Arg	Туг 55	Thr	G 1 y	V a 1	Pro	S c r 6 0	Агд	Phc	Ser	G 1 y
S c r 6 5	Gly	Ser	G 1 y	Thr	A s p 7 0	Phc	Thr	Lcu	Thr	I 1 c	S c r	Ser	Lcu	G 1 n	P r o 8 0
A s p	A s p	Phc	Ala	Thr 85	Туr	Туr	C y s	Gly	G 1 n 9 0	Ser	Туг	A s n	Туг	Pro 95	Phe
Thr	Phc	Gly	G 1 n 1 0 0	Gly	Thr	Lys	V a 1	G l u 1 0 5	V a l	Lys					

(2) INFORMATION FOR SEQ ID NO:104:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids (B) TYPE: amino acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:104:
- Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ser

-continued

 Ala
 Ile
 Ile
 Ile
 Trp
 Val
 Arg
 Gln
 Ala
 Pro
 Gly
 Gln
 Gly
 Leu
 Glu
 Trp
 Met

 Gly
 Gly
 Ile
 Val
 Pro
 Met
 Pro
 Pro
 Pro
 Asn
 Tyr
 Ala
 Gln
 Lys
 Phe

 Gln
 Gly
 Arg
 Val
 Thr
 Ile
 Thr
 Ala
 Asp
 Glu
 Ser
 Thr
 Asn
 Thr
 Ala
 Tyr

 Mcl
 Glu
 Leu
 Ser
 Leu
 Arg
 Ser
 Glu
 Asp
 Thr
 Ala
 Phe
 Cys

 Ala
 Gly
 Tyr
 Gly
 Ile
 Tyr
 Ser
 Pro
 Glu
 Glu
 Tyr
 Asn
 Gly
 Gly
 Leu

 Val
 Thr
 Val
 Ser
 Ser
 Ser
 Pro
 Pro
 Glu
 Glu
 Tyr
 Asn
 Gly
 Gly
 Leu

(2) INFORMATION FOR SEQ ID NO:105:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:105:

(2) INFORMATION FOR SEQ ID NO:106:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 115 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA (oligonucleotide)
- ($\,x\,$ i) SEQUENCE DESCRIPTION: SEQ ID NO:106:

TTTTTTCTAG ACCACCATGG AGACCGATAC CCTCCTGCTA TGGGTCCTCC TGCTATGGGT 60
CCCAGGATCA ACCGGAGATA TTCAGATGAC CCAGTCTCCG TCGACCCTCT CTGCT 115

(2) INFORMATION FOR SEQ ID NO:107:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 120 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

	(D) TOPOLOGY: linear					
(ii) MOL	ECULE TYPE: DNA (oligo	onucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:107:				
T T T T A A G C T T	GGGAGCTTTG	CCTGGCTTCT	GCTGATACCA	GGATACATAA	GTATCCACAT	6 0
TTTCACTGGC	CTTGCAGGTT	ATGGTGACCC	TATCCCCGAC	GCTAGCAGAG	AGGGTCGACG	1 2 0
(2) INFORMATION	FOR SEQ ID NO:108:					
	JENCE CHARACTERISTI (A) LENGTH: 118 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	onucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:108:				
TTTTAAGCTT	Стаатттат	GGGCATCCAA	CCGGTACACT	GGGGTACCTT	CACGCTTCAG	6 0
TGGCAGTGGA	TCTGGGACCG	A T T T C A C C C T	CACAATCAGC	TCTCTGCAGC	CAGATGAT	1 1 8
(2) INFORMATION	FOR SEQ ID NO:109:					
	JENCE CHARACTERISTIC (A) LENGTH: 120 base (B) TYPE: nucleic acid (C) STRANDEDNESS: 9 (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQU	JENCE DESCRIPTION: SE	EQ ID NO:109:				
TTTTTCTAG	AGCAAAAGTC	T A C T T A C G T T	TGACCTCCAC	СТТССТСССС	TGACCGAACG	6 0
T G A A T G G A T A	GTTGTAACTC	TGTCCGCAGT	AATAAGTGGC	GAAATCATCT	GGCTGCAGAG	1 2 0
(2) INFORMATION I	FOR SEQ ID NO:110:					
1	JENCE CHARACTERISTIC (A) LENGTH: 114 base p (B) TYPE: nucleic acid (C) STRANDEDNESS: 8 (D) TOPOLOGY: linear	pairs				
(ii) MOLI	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQU	ENCE DESCRIPTION: SE	Q ID NO:110:				
TTTTTCTAGA	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	тсстсствтс	AGGTACCGCG	6 0
GGCGTGCACT	CTCAGGTCCA	GCTTGTCCAG	TCTGGGGCTG	A A G T C A A G A A	ACCT	1 1 4
(2) INFORMATION I	FOR SEQ ID NO:111:					
(VENCE CHARACTERISTIC (A) LENGTH: 121 base of (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii) MOLI	ECULE TYPE: DNA (oligo	nucleotide)				
(xi)SEQU	ENCE DESCRIPTION: SE	Q ID NO:111:				
ттттбааттс	TCGAGACCCT	GTCCAGGGGC	CTGCCTTACC	CAGTTTATCC	AGGAGCTAGT	6 0
AAAGATGTAG	CCAGAAGCTT	TGCAGGAGAC	CTTCACGGAG	CTCCCAGGTT	TCTTGACTTC	1 2 0
A						1 2 1

(2) INFORMATION FOR SEQ ID NO:112:													
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 137 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:112:													
TTTTGAATTC TCGAGTGGAT GGGAAGGATT GATCCTTCCG ATGGTGAAGT TCACTACAAT	6 0												
CAAGATTTCA AGGACCGTGT TACAATTACA GCAGACGAAT CCACCAATAC AGCCTACATG	1 2 0												
GAACTGAGCA GCCTGAG	1 3 7												
(2) INFORMATION FOR SEQ ID NO:113: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 134 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:113:													
TTTTTCTAGA GGTTTTAAGG ACTCACCTGA GGAGACTGTG ACCAGGGTTC CTTGGCCCCA	6 0												
GTCAGCAAAC CAGGGCAGAA ATCCTCTTGC ACAGTAATAG ACTGCAGTGT CCTCTGATCT	120												
CAGGCTGCTC AGTT	1 3 4												

65

What is claimed is:

1. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from human acceptor immunoglobulin heavy and light chains, which humanized immunoglobulin specifically binds to an antigen with an affinity constant of at least 10⁷ M⁻¹ and no greater than about four-fold that of the donor immunoglobulin, wherein said humanized immunoglobulin comprises amino acids from the donor immunoglobulin framework outside the Kabat and Chothia CDRs, wherein the donor amino acids replace corresponding amino acids in the acceptor immunoglobulin heavy or light chain frameworks, and each of said donor amino acids:

- (I) is adjacent to a CDR in the donor immunoglobulin sequence, or
- (II) contains an atom within a distance of 4 Å of a CDR in said humanized immunoglobulin.
- 2. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from 55 human acceptor immunoglobulin heavy and light chains, which humanized immunoglobulin specifically binds to an antigen with an affinity constant of at least 10⁷ M⁻¹ and no greater than about four-fold that of the donor immunoglobulin, wherein said humanized immunoglobulin comprises 60 amino acids from the donor immunoglobulin framework outside the Kabat and Chothia CDRs, wherein the donor amino acids replace corresponding amino acids in the acceptor immunoglobulin heavy or light chain frameworks, and each of said donor amino acids:
 - (I) is adjacent to a CDR in the donor immunoglobulin sequence, or

- (II) contains an atom within a distance of 5 Å of a CDR in said humanized immunoglobulin.
- 3. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from human acceptor immunoglobulin heavy and light chains, which humanized immunoglobulin specifically binds to an antigen with an affinity constant of at least 10⁷ M⁻¹ and no greater than about four-fold that of the donor immunoglobulin, wherein said humanized immunoglobulin comprises amino acids from the donor immunoglobulin framework outside the Kabat and Chothia CDRs that replace the corresponding amino acids in the acceptor immunoglobulin heavy or light chain frameworks, and each of said amino acids:
 - (I) is adjacent to a CDR in the donor immunoglobulin sequence, or
 - (II) contains an atom within a distance of 6 Å of a CDR in said humanized immunoglobulin.
- 4. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from acceptor immunoglobulin heavy and light chains, which humanized immunoglobulin specifically binds to an antigen with an affinity constant of at least $10^7 \,\mathrm{M}^{-1}$ and no greater than about four-fold that of the donor immunoglobulin, wherein said humanized immunoglobulin comprises amino acids from the donor immunoglobulin framework outside the Kabat and Chothia CDRs that replace the corresponding amino acids in the acceptor immunoglobulin heavy or light chain frameworks, wherein each of these said donor amino acids:
 - (I) is adjacent to a CDR in the donor immunoglobulin sequence, or

- **164**A humanized immunoglobulin
- (II) is capable of interacting with amino acids in the CDRs, or
- (III) is typical at its position for human immunoglobulin sequences, and the replaced amino acid in the acceptor is rare at its position for human immunoglobulin 5 sequences.
- 5. A humanized immunoglobulin according to claim 1, 2 or 3 which specifically binds to an antigen with an affinity of between $10^8~M^{-1}$ and $10^{10}~M^{-1}$.
- 6. A humanized immunoglobulin according to claim 1, 2 10 or 3 which specifically binds to an antigen with an affinity of no greater than about two-fold that of the donor immunoglobulin.
- 7. A humanized immunoglobulin according to claim 1, 2, 3 or 4 which is substantially pure.
- **8**. A pharmaceutical composition comprising a humanized immunoglobulin according to claim **1**, **2**, **3** or **4** in a pharmaceutically acceptable carrier.

- 9. A humanized immunoglobulin according to claim 1, 2 or 3, wherein the antigen is an IL-2 receptor.
- 10. A humanized immunoglobulin according to claim 1, 2 or 3 wherein the donor immunoglobulin is the anti-Tac antibody.
- 11. A humanized immunoglobulin according to claim 1, 2 or 3, further comprising an amino acid outside the Kabat and Chothia CDRs that replaces the corresponding amino acid in the acceptor immunoglobulin heavy or light chain frameworks, wherein said amino acid is typical for its position in human immunoglobulin sequences and said corresponding amino acid in the acceptor immunoglobulin is rare for its position in human immunoglobulin sequences.

* * * * *